

# A gradient analysis of communities of macrofungi and slime moulds on decaying beech logs

En gradientanalyse af samfund af storsvampe og slimsvampe på faldne bøgestammer



## Specialerapport

Jacob Heilmann-Clausen  
Afdeling for alger og svampe  
Botanisk Institut, Københavns Universitet  
1999

Vejleder: Thomas Læssøe (Lektor).

# **A gradient analysis of communities of macrofungi and slime moulds on decaying beech logs**

**En gradientanalyse af samfund af storsvampe (Fungi pp.) og slimsvampe  
(mycomycota) på faldne bøgestammer**

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Skovbillede fra Suserup Skov. I forgrunden ses stammen F3NW-B7 (nedbrydningsstadie 5), bagved ses F3NW-B8 (nedbrydningsstadie 4).

Insite Suserup Skov. In the foreground the log F3NW-B7 (decay class 5), is seen, in the background F3NW-B8 (decay class 4) is situated.

Photo Jacob Heilmann-Clausen, May 1995.

## Forord

Mit specialeforløb har været usædvanligt. Det feltarbejde der ligger til grund for den foreliggende rapport udførte jeg så tidligt som i 1994 og 1995, mens bearbejdelsen af data samt en væsentlig del af bestemmelsesarbejdet er foregået i 1998 og 1999. I 1994, da jeg påbegyndte registreringsarbejdet, var jeg i færd med at afslutte biologistudiets første del. Jeg havde ikke en udviklet kritisk tilgang til projektdesign og metodevalg og havde ikke et speciale i tankerne. Arbejdet udviklede sig imidlertid i omfang og i begyndelsen af 1995 blev det klart for mig, at en systematisk og videnskabelig bearbejdning af det indsamlede materiale var mere end jeg kunne klare ved siden af det almindelige studieliv. Som alternativ til at opgive undersøgelsen valgte jeg at fortsætte feltarbejdet året ud, for derefter at putte hele projektet i skuffen til bedre tider. Samtidigt traf jeg aftale med Thomas Læssøe, som indvilgede i at være vejleder for mig.

De væsentligste ulemper ved forløbet har været at jeg ikke har haft tid og viden nok til at designe feltarbejdet optimalt og at jeg ikke har haft en vejleder inde i billede på et tidligere tidspunkt. Konkret har det medført et mindre optimalt undersøgelsesdesign:

- Registreringen af svampe på stammerne har udelukkende baseret sig på tilstedeværelse kontra fravær af arter. En estimering af forekomstmønstre (areal, mikrohabitater) havde været interessant, og i et vist omfang muligt indenfor forsøgsdesignet.
- Registreringen af miljøvariable er ikke optimal. Bl.a. måtte jeg, efter adskillige arbejdsdage opgive et storstilet forsøg på at måle mikroklima vha. træklodser, pga. tidsmangel og et ugennemtænkt fsøgsdesign. Tilsvarende løb et forsøg på mekanisk at måle træhårdhed i vasken, idet jeg ikke havde tid til at kalibrere og tilpasse det lånte udstyr til nedbrudt ved.
- Min forståelse for betydningen af repræsentative indsamlinger var begrænset, hvilket har medført at adskillige arter ikke er belagt fra undersøgelsen.

Forløbet har dog også haft klare fordele, ikke mindst tidsmæssigt. Projektet havde næppe fået det nuværende omfang indenfor et "standardiseret" specialeforløb, da et sådant ikke tillader to hele feltsæsoner.

Jeg har valgt i min bearbejdning af det indsamlede materiale at koncentrere indsatsen om at skrive et artikelmanuskript, så vidt muligt i en form der gør det umiddelbart klart til indsendelse til et internationalt anderkejendt og "peer-reviewed" tidsskrift. Det her præsenterede manuskript foreligger dog i en "brutto-version" som skal beskæres inden det indsendes.

Konkret har jeg skrevet manuskriptet med henblik på publicering i det engelske tidsskrift "Mycological Research", med de krav til tekstudformning det indebærer. Den lidt besynderlige citering af artikler med tre forfattere (1ste gang nævnt f.eks. Chapela, Boddy & Rayner 1988; 2den gang nævnt Chapela et al 1988), er en konsekvens af disse krav.

En del personer er takket i artikel-manuscriptet og vil ikke blive yderligere individuelt takket her. De skal dog alle vide at deres hjælp har været uvurderlig. Yderligere vil jeg gerne takke ansatte og studerende ved AAS, Københavns Universitet for hjælp med diverse opgaver, diskussioner mv. En særlig tak skal lyde til Chris B. Jakobsen, for hjælp med diverse computerproblemer. Desuden en stor tak til Christian Lange, Århus Universitet, som har været til uvurderlig hjælp, da det brændte på med de sidste figurer. Sidst, men ikke mindst vil jeg takke Malene, Silja og Sofia for god opbakning gennem hele forløbet

Den her foreliggende rapport er ikke den første der belyser samfund af ved-nedbrydende svampe i Suserup Skov. To projekthold under fagmodulet Thalofytter (Svampe) / a-modulet Svampetaxonomi vejledt af Thomas Læssøe har barslet med rapporter indenfor området. Ikke mindst den sidstnævnte er relevant i forhold til den her foreliggende rapport:

Hellensten, H. S., Closter, R. M. & Wamberg, C. (1997). Fungaen på egestammer i tre nedbrydningsstader. - Projetrapport, a-modulet svampetaxonomi, AAS, Københavns Universitet.

Madsen, B. & Johansen, C. (1994). Succession af kernesvampe på væltede bøgestammer i Suserup Skov. – Projetrapport, fagmodulet thalofytter (svampe), AAS, Københavns Universitet.

Begge rapporter er upublicerede, men tilgængelige via Thomas Læssøe, AAS, Botanisk Institut, Københavns Universitet.



Myceliestrenge af *Phanerochaete velutina* i kontakt med et frugtlegeme af *Hyphoderma argillaceum*.  
Mycelial cords of *Phanerochaete velutina* makes contact to a sporocarp of *Hyphoderma argillaceum*.  
Photo: Jacob Heilmann-Clausen.

## Dansk sammenfatning.

### En gradientanalyse af samfund af storsvampe og slimsvampe på faldne bøgestammer.

Svampe er en dominerende organismegruppe i nedbrydningen af dødt ved, og spiller derved en uhyre vigtig rolle i alle skov-økosystemer. Nedbrydningen af dødt ved af store dimensioner, f.eks. hele stammer, er en kompliceret og langvarig proces, som typisk involverer mange forskellige svampearter, foruden repræsentanter fra andre organismegrupper, heriblandt slimsvampe. Generelt regnes mikroklima, substratkvalitet (træart, nedbrydningsstadie) og den tilgængelige svampe-artspulje som de vigtigste parametre i forhold til udformningen af svampesamfund på ved under nedbrydning. Tidlige undersøgelser har undersøgt effekterne af disse parametre hver for sig, men kun meget få undersøgelser har forsøgt en mere overordnet analyse. Hovedformålet med denne rapport er et forsøg på en sådan analyse.

Mit speciale baserer sig på registreringer af svampefrugtlegemer på 70 systematisk udvalgte faldne bøgestammer i Suserup Skov, syd for Sorø. Alle de undersøgte stammer er store. De mindste har en brysthøjdediameter på 70 cm, mens den største når en diameter på 168 cm. (Til sammenligning hugges bøg i Danmark normalt ved en diameter på maksimalt 70 cm). Udenfor svampe indgår slimsvampe (også knedt som svampedyr) i undersøgelsen. Forekomsten (+/-) af frugtlegemer blev undersøgt på hver stamme ved i alt 9 besøg i løbet af 1994 og 1995, og er opsummeret i en stamme/arts datamatrix (se appendix 3). En del materiale blev bestemt i felten, men i mange tilfælde var det nødvendigt at indsamle materiale til mikroskopisk bestemmelse. I tilknytning til hver undersøgt stamme har jeg målt eller estimeret en række miljø-parametre (se appendix 4) som godt kan deles i tre grupper: 1. Stamme-variable (dimension, kompleksitet, nedbrydningsgrad, alder mv.) 2. Mikroklimatiske variable (vindexponering, jordfugtighed, afstand til skovbryn mv.). 3. Floristiske variable (antal plantearter omkring stammen, frekvens af enkelt-plantearter). Strukturen i stamme/arts datamatrix'en blev undersøgt ved hjælp af DCA\* (detrended correspondance analysis). De resulterende DCA-akser blev herefter analyseret i forhold til de undersøgte miljøparametre, dels ved hjælp af simpel korrelations analyse, dels ved hjælp af multibel regressions analyse.

I alt er 302 arter erkendt i materialet, heraf 25 arter af slimsvampe og 277 arter af svampe. En del af de fundne arter er ikke tidligere rapporteret fra Danmark (se appendix 2). For både svampe og svampedyr blev det fundet at artsantallet er højest på intermediaert nedbrudte stammer.

Den første DCA-akser (DCA1) repræsenterer langt den stærkeste gradient i materialet og fremstår som en uveydig successionsgradient, beskrivende nedbrydningsforløbet fra det friskfaldne stamme til den stærkt nedbrudte. DCA2 er mere kompleks men afspejler især graden af mikroklimatisk stress (udtørnings-stress) og stammens nedbrydningshastighed. Endelig fremstår DCA3 som en svag og noget tvivlsom jordbundsgradient (fra fattig til rig bund). På grund af den svage gradientstruktur langs DCA3 blev yderligere DCA-akser ikke undersøgt.

De fundne resultater stemmer generelt godt overens med tidligere undersøgelser og teorier på området. En markant dominans af en successionsgradient er flere gange beskrevet fra studier af svampesamfund på nåletræ. Den konkrete samfundsudvikling langs aksen afspejler navnlig en tilpasning til øget konkurrence efterhånden som nedbrydningsforløbet skrider frem, men synes også at afspejle, at svampearter der forekommer sent i forløbet ofte er særligt tilpassede til at videreføre nedbrydningen af ved, der allerede er delvist nedbrudt.

Koblingen mellem nedbrydningshastighed og mikroklimatisk stress langs DCA2 stemmer godt overens med laboratorieforsøg, men er ikke tidligere beskrevet fra naturlige forhold. Koblingen mellem de to faktorer er kompleks og det synes sandsynligt at nedbrydningshastigheden afspejler graden af mikroklimatisk stress påført

både udefra (vind mv.) men også indefra, idet visse kærnesvampe tilsyneladende udtrørre erobret ved aktivt for at holde andre nedbrydersvampe på afstand. Aksen er stærkt negativt korreleret med diversiteten (antal arter pr. stamme) af både svampe og svampedyr, dvs. at langsomt nedbrudte/mikroklimatisk stressede stammer generelt er artsfattige. Kun barksvampe er ligeligt repræsenteret langs aksen og fremstår derved som en generelt udtrørringstolerant gruppe.

Jordbundsgradienten udtrykt langs DCA3 er som nævnt noget svag og tvivlsom, men ville formodentlig blive mere klart udtrykt, hvis stammer fordelt over et bredere spektrum af jordbundstyper end repræsenteret i Suserup Skov, blev inkluderet i analysen.

Som helhed giver undersøgelsen et overordnet indblik i svampe-samfundsudviklingen på faldne bøge-stammer og vil være et godt udgangspunkt for mere detaljerede studier, f.eks på mikrohabitateniveau, eller mht. at undersøge samspil med andre organismegruppers betydning for nedbrydningsforløbet.

\*DCA er en metode til indirekte gradientanalyse, dvs. at metoden fremdrager grader fra et givet materiale udelukkende udfra de enkelte arters fordeling på prøveflader (i dette tilfælde bøgestammer). De fremkomne grader kan derefter tolkes udfra målte miljøvariable. Til forskel er f.eks. CCA (canonical correspondance analysis) en metode til direkte gradient analyse, der direkte indrager undersøgte miljøvariable i udtrækningen af akser.

Den første akse i en DCA (DCA1) udtrækkes således, at de undersøgte enkeltarters optima bliver bedst muligt spredt langs aksen, samtidigt med at prøvefladerne bliver fordelt således, at dissimilariteten imellem disse bliver bedst muligt bevaret. Successive akser udtrækkes på tilsvarende vis, dog således at den variation der er udtrykt langs allerede udtrukne akser holdes udenfor analysen.

Fremgangsmåden gør at DCA1 altid er den akse, der forklarer mest variation. "Eigen-værdien" fortæller hvor meget variation efterfølgende akser forklarer. Når der, som i den her præsenterede undersøgelse, er et stort spring i eigen-værdi mellem DCA1 og DCA2-3, indikerer dette, at DCA1 forklarer betydeligt mere variation end de to andre akser.

Gradientlængden er et udtryk for mængden af  $\beta$ -diversitet (arts-turnover) udtrykt langs en given akse. En gradientlængde på 3.5 SD enheder indikerer populært sagt at prøvefelter (stammer) placeret i hver sin ende af akseren ikke har nogen arter til fælles.



Mycelieskjolde (zone-lines) opdeler en bøgestamme i territorier, hver optaget af en bestemt svampe-individ. Mycelieskjolde dannes af mange svampe som barrierer imod indtrængende konkurrenter.  
Pseudosclerotial plates (zone-lines) dividing a beech log in compartments, each occupied by one fungal individual.

Photo: Jacob Heilmann-Clausen, May 1995.

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## A gradient analysis of communities of macrofungi and slime moulds on decaying beech logs

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Jacob Heilmann-Clausen

Department of mycology, University of Copenhagen, Øster Farimagsgade 2D, DK-1353 København K., Denmark.

The occurrence of fungi and slime moulds on 70 decaying beech logs was surveyed based on the presence/absence of sporocarps. In total 275 species of fungi and 25 of slime moulds were recognised. The recordings were summarised in a log/species datamatrix, the structure of which was investigated using detrended correspondance analysis (DCA). The ecological nature of the gradients, expressed by the first three DCA-axes were subsequently investigated by considering a number of environmental and log related variables. The first and strongest gradient was found to reflect the community development from newly dead logs, through the complete process of decay. The second gradient was found to be complex, reflecting both the rate of decay and the amount of microclimatic stress. The third rather weak gradient was found to reflect an influence of soil conditions.

Finally the gradients are discussed in a context of fungal ecological strategy theories, and a generalised model of the community development on the studied logs is proposed.

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Running title: Macrofungi and slime moulds on beech logs

Keywords: DCA, *Fagus*, saprotrophic, succession, wood.

The perception and acceptance of dead wood as a key element in forest ecosystem functioning have increased dramatically in recent times (e.g. Harmon et al. 1986, Franklin, Shugart & Harmon 1987, Samuelsson, Gustafsson & Ingelög 1994), and so has the need to understand the associated biological processes. Dead wood is of crucial importance for biodiversity (Samuelsson et al. 1994, Christensen & Emborg 1996, Ohlson et al. 1997) and may constitute an important resource and sink of nutrients, which are gradually released from the wood during decomposition (Harmon et al. 1986, 1994). The process of wood decomposition involves several groups of organisms, such as bacteria, protozoa, nematoda, ar-

thropods and insects, but under normal conditions fungi are the primary agents of decay (Rayner & Boddy 1988, Boddy 1992). The decay of large woody units, in particular whole logs, is a slow process which may take decades or even centuries to complete (Harmon et al. 1986, Rayner & Boddy 1988). The associated decomposer community development is very complex even when only fungi are considered (Boddy 1992, Renvall 1995).

In general the available species pool, microclimate and substrate quality (as determined i.e. by tree species and decay stage) have been denoted as the most important variables determining the fungal community structure in

decaying wood (Gilbertson 1980, De Vries & Kuyper 1988, Rayner & Boddy 1988).

Several studies have dealt with the effects of these factors separately. For instance Coates & Rayner (1985a-c), Chapela, Boddy & Rayner (1988), Lange (1992) and Renvall (1995) have investigated the community changes occurring as decay proceeds. Butin & Kowalski (1983, 1986), Griffith & Boddy (1990) and Keizer & Arnolds (1990) have compared the wood decaying fungi of several different tree species. Boddy (1983), Boddy, Gibbon & Grundy (1985) and Griffith & Boddy (1991a,b) have surveyed the effects of microclimate, and De Vries & Kuyper (1988) and Lindblad (1996) have surveyed the impacts of the available species pool. On the other hand very few studies have tried to evaluate the relative importance of these variables in a multivariate context.

Several kinds of multivariate methods are available for ecological studies (Økland 1990, Kenkel & Booth 1992, Legendre & Legendre 1998). In vegetation ecology ordination methods have been widely applied during recent years, and in several cases new insight in complex vegetation processes and structures have been gained (Økland 1990). In studies of fungal ecology ordination methods have been applied much more occasionally, and has yet to gain popularity. Høiland & Bendiksen (1996) and Lindblad (1997) applied detrended correspondance analysis (DCA) in their studies of fungal communities of decaying conifer logs. However, the main subject of both studies were to investigate the impacts of forestry on the wood fungi, hence less attention was given to the understanding of the general community processes of the studied logs.

In the present study detrended correspondance analysis (DCA) was applied in order to reveal the fungal community structure on decaying logs of beech (*Fagus sylvatica* L.). Apart from fungi, slime moulds were included in the study. Slime moulds are not known as active wood decayers, but seems mainly to be phagotrophic, feeding on bacteria and fungi (Ing 1994). The main questions addressed by the study are:

1. Which coenoclines are the most important in the material?

2. Can these be interpreted to reflect underlaying ecological gradients?

3. How do the observed coenoclines/ecological gradients relate to current theories of fungal ecological strategies and decay development?

The study is entirely based on registration of sporocarps, a method which is considered in detail in the discussion.

## MATERIAL AND METHODS

Suserup Skov (19.2 ha, 7-31 m a.s.l., 55°22'N, 11°34'E) is a near-natural temperate deciduous forest situated in the eastern part of Denmark on the island Zealand. The climate is cool-temperate and suboceanic with an annual mean temperature of 8.10 °C and an annual mean precipitation of 635 mm with maximum occurring in July to December (Emborg, Christensen & Heilmann-Clausen 1996).

The forest borders open land with scattered woody vegetation to the north and east, and pasture-land with a fairly well-developed woody vegetation to the west. To the south, the forest borders lake Tystrup.

The forest is situated on an undulating elevated plateau to the north and some +/- south-facing slopes towards the lake, intermixed with smaller flat areas. The soils are generally deep mull or moder-soils, developed on various types of glacial tills in the elevated parts and on lacustrine deposits towards the lake (Vejre & Emborg 1996). The pH in the topsoil generally varies from 3.9 to 4.5, but locally values above 7 are reached (Feilberg 1993, Møller 1997).

The growth conditions are generally favorable and the typical maximum tree-height is about 40 m. The dominant tree-species are beech (*Fagus sylvatica* L.), ash (*Fraxinus excelsior* L.), oak (*Quercus robur* L.) and elm (*Ulmus glabra* L.), which in most parts of the forest forms a complete and dense canopy. The herb-layer is well developed, but is generally completely dominated by early flowering herbs such as *Anemone* spp., *Mercurialis perennis* L. and *Corydalis bulbosa* (L.)DC.

The cultural history of the forest is well-studied (Fritzbøger & Emborg 1996, Emborg et al. 1996). The impact of forestry has been very limited, at least since 1854, with only minor, selective cuttings having been

made, especially during war time. Since 1960 the forest has been kept as a strict non-intervention forest, and it is now close to a dynamic steady-state (Embørg, Christensen & Heilmann-Clausen 1999), with plenty of standing and fallen dead wood, and equilibrium between regeneration, mature stands and breakdown. Further details on the forest structure and -composition have been published by Embørg et al.(1996).

#### *Dating and selection of logs.*

In total 70 logs were selected for the study. Prior to this an attempt was made to establish the time of death (log age) of all dead beech trees in the forest with a diameter in breast height (dbh) above ca. 60 cm. For this purpose a sequence of high resolution aerial photos were used. All dead beech trees known by 1994 were tracked backwards in the photo sequence, until they were identified as standing, living trees.

The log age was successfully determined for 130 beech logs, which were subsequently divided into seven age-classes (1960-67, 1967-72, 1972-78, 1978-81, 1981-1985, 1985-90, 1990-93), and four dbh classes (70-89 cm, 90-109 cm, 110-129 cm, >130 cm). The inequality in the age classes mirrors the unequal distribution of available aerial photos.

The seven age classes were used as basis for the selection of logs for the study. - From each age class ten logs were selected with the attempt to obtain an equal representation of the dbh classes within each. In cases of multiple possible choices to fulfill this aim, logs were selected randomly.

#### *Registration of log variables*

In addition to the log variables mentioned above (log age and dbh) the following variables were measured or recorded for each log:

**Log type** (uprooted with a distinct root plate, broken at root neck, broken 2-7 m above ground-level, broken 8-15 m above ground-level). Number of **fractures** and **forks** on the log, incl. major branches, with diam. equal to or larger than 50 cm. Medium **decay class**, **soil humidity**, **wind exposure** and **sun exposure**, subjectively judged according to the parametres given in Tables 1 & 2. **Bark cover**, **moss cover** and distribution of stromata and black-stained wood of the pyrenomycete *Eutypa spinosa* (**Eutypa cover**) on the log surface, estimated to the nearest 10 % of the total log surface. Degree of **soil contact**, estimated to the nearest 10 % of the log-length. **Distance to the forest edge** measured to the next 5 m.

The occurrence of the plants *Allium ursinum* L., *Anemone*

Characteristics	
Class 1	Wood hard, a knife (with a thin blade) penetrates only a few mm into the wood, bark intact, twigs (diam. < 1 cm) intact.
Class 2	Wood rather hard, a knife penetrates less than 1 cm into the wood, bark starting to break up, twigs +/- lost, branches (diam. 1-4 cm) intact
Class 3	Wood distinctly softened, knife penetrates ca. 1-4 cm into the wood, except for parts colonized by certain pyrenomycetes (in particular <i>Eutypa spinosa</i> , <i>Kretzschmaria deusta</i> and <i>Xylaria hypoxylon</i> ), bark partly lost, branches +/- lost, original log circumference intact
Class 4	Wood strongly decayed, knife penetrates ca. 5-10 cm into the wood, except for parts colonized by certain pyrenomycetes (see above), bark lost in most places, original log circumference disintegrating
Class 5	Wood very strongly decayed, either to a very soft crumbly substance, or being flaky and fragile with numerous remnants of pseudosclerotial plates, these defining the log surface, knife penetrates in most places more than 10 cm into the wood, original log circumference not or hardly recognizable.

Table 1. Characteristics used for the classification of logs in decay classes.

	Soil humidity	Wind exposure	Sun exposure
Class 1	Well drained forest soils on hill tops.	Exposed forest at edges and on hills.	Forest edges and open forest on south facing slopes.
Class 2	Other high, well drained forest soils.	Open forest, without a proper understorey.	Open, sunlit forest.
Class 3	Normal, well drained forest soils.	Normal forest, with sparse understorey.	Normal forest on level areas.
Class 4	Low, +/- waterlogged soils.	Normal forest, with a well developed understorey.	Normal forest, either with a dense canopy or on north facing slopes.
Class 5	Swamp soils, +/- inundated during wintertime.	Well sheltered places, protected by a dense understorey.	High forest with a dense canopy and on north facing slopes.

Table 2. Characteristics used for the classification of logs in classes of soil humidity, wind exposure and sun exposure.

*nemorosa* L., *A. ranunculoides* L., *Arum maculatum* L., *Corydalis bulbosa*, *C. intermedia* (L.) Merat, *Ranunculus ficaria* L., *Gagea spathacea* (Hayne) Salisb., *Galium odoratum* (L.) Scop., *Lamiastrum galeobdolon* (L.) Ehrend. & Polatschek, *Mercurialis perennis*, *Pulmonaria obscura* Dumont., *Sanicula europaea* L. and *Viola riviana* Reichenb./reichenbachiana Jordan, were recorded around each log in May 1995. The relative distribution of each species, and of vegetation-less areas were estimated to the nearest 10 % of the forest-floor area surrounding the individual log. The variable floristic deviation was derived by considering the forest floor area covered by plants other than the generally dominant *A. nemorosa*, whereas plant diversity was obtained by summing up the number of plant species occurring in the vicinity of each log.

The independent registration of log age and decay rate made it possible to estimate the log specific decay rate, by considering the deviation from the mean decay class within each age class. To obtain a more precise comprehension of the log specific decay development, the logs were re-investigated in 1999, and the decay stage were re-estimated. In this estimation an additional decay class (class 6) were introduced for almost completely decom-

posed logs.

In Table 3 all log variables are summarized.

#### *Registration of sporocarps*

Sporocarps of fungi and slime moulds were recorded on all logs at nine occasions between May 1994 and December 1995. (8-9 May 1994, 10 July 1994, 23-24 Aug. 1994, 4 Oct. 1994, 24 Feb. 1995, 25 Sep. 1995, 1 Oct. 1995, 15 Oct. 1995, 14+17 Dec. 1995). The whole log were investigated, including the stump and branches thicker than 10 cm. Sporocarps were identified *in situ* or collected for later identification. The visits in September and October 1995 were rather superficial and included only species determinable in the field.

Within the basidiomycetes and slime moulds all taxonomic groups are included in the study, within the ascomycetes, non-stromatic pyrenomycetes are excluded. However, due to the project-scale it was not intended to make a complete registration of all species on all logs. In all likelihood genera with very inconspicuous sporocarps (e.g. *Basidiiodendron*, *Botryobasidium*, *Sistotrema* and *Tulasnella* within the basidiomycetes and *Arachnopeziza* and *Hyaloscypha* within the ascomycetes) are underrepresented. Some common species were treated in an un-

Variable	Data type	Scale	Range	Median	Unit
Log type	Nominal	4 levels			
Dbh.	Ratio	continuous	70-168	110	cm
Fractures	Ratio	count	0-5	2	Number
Bole forks	Ratio	count	0-4	1	Number
Decay stage	Ordinal	5 levels	1-5	3	Class
Log age	Ratio	continuous	2-31	15	Years
Bark cover	Ratio	12 levels	0-100	10	%
Moss cover	Ratio	12 levels	0-60	5	%
Eutypa cover	Ratio	12 levels	0-70	20	%
Soil contact	Ratio	11 levels	10-100	90	%
Soil type	Nominal	3 levels			Class
Soil humidity	Ordinal	5 levels	1-5	4	Class
Wind exposure	Ordinal	5 levels	1-5	4	Class
Sun exposure	Ordinal	5 levels	1-5	4	Class
Dist. to edge	Ratio	continuous	5-135	70	m
Plant diversity	Ratio	count	1-6	2	Number
Plant cover	Ratio	11 levels	20-100	90	%
Floristic deviance	Ratio	12 levels	0-100	*	%
Decay rate	Ordinal	5 levels	-2 - +2	0	Class

Table 3. Log variables measured or estimated for each log. \*The variable, floristic deviance, is further subdivided in eight variables, each representing a plant species; *Anemone nemorosa* (Ane nem), *Anemone ranuculoides* (Ane ran), *Corydalis bulbosa* (Cor bul), *Galium odoratum* (Gal odo), *Lamiastrum galeobdolon* (Lam gal), *Mercurialis perennis* (Mer per), *Ranunculus ficaria* (Ran fic), *Viola riviana/reichenbachiana* (Vio riv) and a ninth variable (Other species) with the remaining species (*Allium ursinum*, *Arum maculatum*, *Corydalis intermedia*, *Gagea lutea*, *Pulmonaria obscura* and *Sanicula europaea*)

critical or broad sense, in order to reduce microscopical work. This was the case for *Galerina marginata* (Batsch) Kühn., *Scutellinia scutellata* (L. ex St. Amans) Lambotte, *Mollisia cinerea* (Batsch ex Mérat) Karst., *Athelia epiphylla* Pers.:Fr., *Botryobasidium conspersum* J. Erikss., and *Peziza micropus* Pers. which were all determined using macroscopical characters, although a microscopical examination is necessary for a critical determination of these taxa. This uncritical approach was chosen after microscopical examinations of dozens of specimens in each species-group, which failed to detect more than one species in each case. Also the concept of *Bisporella citrina* is broad and seems to include an undescribed taxon, microscopically defined by asci regularly shorter than 100 um.

#### Nomenclature

The nomenclature and species-concepts of basidiomycetes follows Hansen & Knudsen (1992, 1997), except for *Sistotrema* pp. (follows Hallenberg 1984), Tomen

tella (follows Kóljalg 1996) and *Stypella* and *Exidia* pp. (follows Roberts 1998). Within the ascomycetes, nomenclature and species-concepts in general follows Hansen & Knudsen (1999). However, within the discomycetes reference to special literature has been necessary in several cases. The nomenclature of slime moulds follows Nannenga-Bremekamp (1991).

#### Data Treatment

Detrended correspondance analysis (DCA) was chosen as ordination method for several reasons. It is easy to access and assumes unimodal species/gradient response curves, in contrast to principal component analysis (PCA) which assumes linear responses. Linear species/gradient responses appears to be rare in nature, and PCA has in general appeared to perform badly on vegetation data (Økland 1990). In DCA, samples (logs) and species are ordinated simultaneously in the same ordination space following an iterative procedure, which aims to optimise the dispersion of species optima. Sample

scores are derived from species scores by a process of weighted averaging. For more details on DCA see Hill & Gauch (1980) or Økland (1990).

The relative powers of DCA axes can be established by comparing eigenvalues and gradient lengths. The gradient length express the amount of β-diversity (species turnover) expressed along the individual axis, whereas the eigenvalue tells how much of the sample variation which can be explained. The eigenvalue is most informative in comparison with eigenvalues of other axes of the same ordination. Økland (1999) has shown that the much used measures of total inertia and percent explained variation have little significance. Hence these measures are omitted in the present paper.

DCA was run using the program PC-ORD 3.0 (McCune & Mefford 1997) using default options. Downweighting of rare species was avoided in the final ordinations, since it tended to weaken correlations with environmental variables. However species occurring on two logs or less were excluded from the ordination.

The consistence of the found DCA axes were evaluated by ordination of subsets as suggested by Laweson (1997) and Ejrnæs & Bruun (1999). Three subsets derived from the full data set, were analysed:

1. slime moulds excluded.
2. Common species (Species occurring on 6 logs or less excluded).
3. Subordinate species (Species occurring on 20 logs or more excluded).

Relationships between DCA-axes and environmental variables were evaluated by Kendall rank correlation. Rank correlation was chosen in favour of its parametric alternatives, since several variables are on ordinal scale while others deviate strongly from normality. Kendall rank correlation was favoured over Spearman rank correlation, since the latter method is less efficient in the presence of many tied ranks (Legendre & Legendre 1998). Subsequently multiple regression was applied in order to clarify the structures of the ecological gradients expressed along the DCA-axes. Backward elimination (see Zar 1999) was used to evaluate which variables

should be selected for the multiple regression models. In the case of curvi-linear relationships polynomial regression was applied. The two nominal variables, soil type and log type were coded as 'dummy' variables (see Zar 1999).

To ease the interpretation of the DCA-axes in respect to fungal community development, the reported fungal species were sub-divided into six morpho-groups (Table 4). For each group the relative contribution to overall species richness were calculated per log, prior to correlation analysis.

All regression and correlation analysis was run using the program JMP for MacIntosh (SAS Institute Inc. 1997).

## RESULTS

In total 302 taxa, 277 species of fungi and 25 species of slime moulds, were recorded during the study, (Table 4). More than one third (124) of the recorded species were found on one or two logs only, and were hence omitted from the ordination. 15 species were found on 35 logs or more.

The number of fungal species per log averaged 30.6, with the highest numbers obtained on logs in intermediate age classes, Fig. 1. At the single most species rich log 78 species were recorded. Logs 7-20 years old were

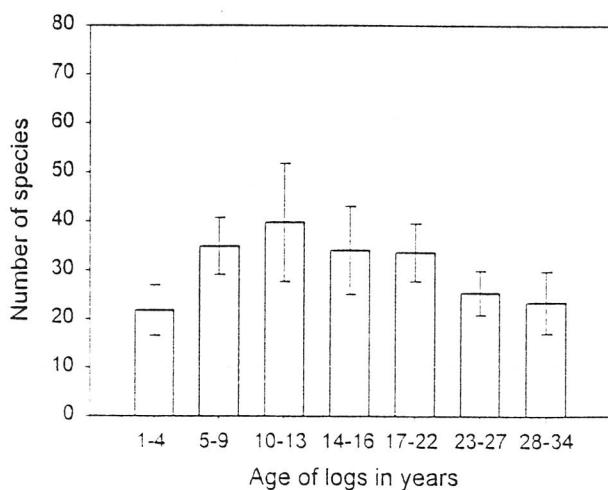


Fig 1. Average number of fungal species per log in the seven age classes. The bars represent 95% confidence limits. Number of logs = 10 in each class.

Abbreviation in DCA-diagrams	Species	Number of logs	Collection-numbers of selected voucher specimens preserved at the Botanical Museum, Copenhagen (C.)
<b>PYRENOMYCETES</b>			
cam poly	Camarops polysperma (Mont.) Miller	9	JHC94-350
	Camarops tubulina (Alb. & Schw.) Shear	2	C
dia disc	Diatype disciformis (Hoffm.: Fr.) Fr.	5	C
	Diatype flavovirens (Pers.: Fr.) Fr.	1	JHC94-015
	Diatype stigma (Hoffm.: Fr.) Fr.	1	
eut spin	Eutypa spinosa (Pers.: Fr.) Tul. & C.Tul.	62	JHC94-139; JHC94-144
	Eutypella quaternata (Pers.: Fr.) Rappaz	2	JHC94-351
hyp gela	Hypocrea gelatinosa (Tode: Fr.) Fr.	4	C
	Hypomyces aurantius (Pers.: Fr.) Tul. & C.Tul.	1	C
hyp coha	Hypoxyylon cohaerens (Pers.: Fr.) Fr.	3	JHC94-146
hyp frag	Hypoxyylon fragiforme (Pers.: Fr.) Kickx	15	C
hyp rubi	Hypoxyylon rubiginosum (Pers.: Fr.) Fr.	15	C
kre deus	Kretzschmaria deusta (Hoffm.: Fr.) P.M.D. Martin	45	C
	Lopadostoma turgidum (Pers.) Traverso	1	JHC94-009
nec cocc	Nectria coccinea (Pers.: Fr.) Fr.	4	C
	Nectria episphaeria (Tode: Fr.) Fr.	1	C
	Nectria peziza (Tode: Fr.) Fr.	2	JHC94-159; JHC94-160
nem atro	Nemania atropurpurea (Fr.: Fr.) Pouzar	4	JHC94-019; JHC94-062; JHC94-356
nem ches	Nemania chestersii (J. D. Rogers & Whalley) Pouzar	9	JHC94-140; JHC94-143; JHC94-162
nem serp	Nemania serpens (Pers.: Fr.) Gray	6	JHC94-352; JHC94-354; JHC94-357
xyl hypo	Protocrea cfr. farinosa (Berk. & Broome) Petch	1	JHC94-010
xyl poly	Xylaria hypoxylon (L.: Fr.) Grev.	42	C
	Xylaria polymorpha (Pers.: Fr.) Grev.	17	C
<b>DISCOMYCETES</b>			
	Arachnopeziza aurata Fuckel	3	JHC94-420
	Arachnopeziza variepilosa (Galán & Raitvii) Huhtinen	1	JHC94-035
asc cyli	Ascocoryne cylindrium (Tul.) Korf	8	C
bis citr	Bisporella citrina (Batsch : Fr.) Korf & S.Carp. s.l.	10	JHC95-224; JHC95-253; JHC95-254
	Cystopezizella conorum (Rehm) Svrcek	1	JHC94-359
	Dasyscyphella nivea (Hedw.: Fr.) Raity.	1	JHC95-202
	Hyaloscypha fuckelii Nanmf.	1	JHC94-036; JHC95-210
lac brev	Lachnum brevipilosum Baral	5	JHC95-252
lac impu	Lachnum impudicum Baral	22	JHC94-069; JHC95-201
lac virg	Lachnum virgineum (Batsch: Fr.) P.Karst.	4	JHC94-360
mol cine	Mollisia cinerea (Batsch) P.Karst.	49	JHC94-432
	Mollisia ligni (Desm.) P.Karst.	1	JHC95-206
neo pura	Neobulgaria pura (Fr.) Petrik	6	C
	Ombrophila sp.	1	JHC94-421
	Orbilia cfr. alnea Velen.	2	JHC94-430; JHC94-150
orb deli	Orbilia delicatula (P.Karst.) P.Karst.	19	JHC94-101b; JHC94-423
	Orbilia epipora (Nyl.) P.Karst.	2	JHC94-037; JHC94-102; JHC94-103
pez micr	Peziza micropus Pers.: Fr.	26	C
pol prui	Polydesmia pruinosa (Jerdon in Berk. & Broome) Boud.	12	JHC95-221
scu scut	Scutellinia scutellata (L.: Fr.) Lambotte	29	C
	Tapesia lividofusca (Fr.: Fr.) Rehm	1	JHC95-209
<b>CORTICOID FUNGI</b>			
ath epip	Athelia epiphylla Pers.	37	JHC95-248; JHC95-249
	Athelopsis glaucina (Bourd. & Galz.) Parm.	1	JHC94-057
	Basidiobolus caesiocinereum (Höhn. & Litsch.) Luck-Allen	1	JHC95-207
	Botryobasidium botryosum (Bres.) J.Erikss.	1	JHC94-028
bot cons	Botryobasidium conspersum J.Erikss.	29	C
	Botryobasidium laeve (J.Erikss.) Parm.	2	JHC94-124
bot prui	Botryobasidium pruinatum (Bres.) J.Erikss.	6	JHC94-055
bot subc	Botryobasidium subcoronatum (Höhn. & Litsch.) Donk	5	C
bre oliv	Brevicellicium olivascens (Bres.) Larss. & Hjortst.	9	JHC95-230; JHC95-231
bys cori	Byssomerulius corium (Fr.) Parm.	3	JHC95-240
cer exce	Ceriporia excelsa (Lund.) Parm.	8	JHC94-198; JHC94-344; JHC95-250
cer reti	Ceriporia reticulata (Hoffm.: Fr.) Dom.	38	C
	Coniophora arida (Fr.) P.Karst.	2	JHC95-195
con pute	Coniophora puteana (Schum.: Fr.) P.Karst.	17	C
	Cristinia gallica (Pilát) Jülich	2	JHC94-183
	Cristinia helvetica (Pers.) Parm.	2	JHC94-081
cyl laev	Cylindrobasidium laeve (Pers.: Fr.) Chamuris	7	JHC95-241

Table 4. Total species list subdivided in morphological groups. Note that several heterobasidiomycetes are assigned as corticioid fungi, due to their thin, apressed sporocarps. A 'C' in the voucher specimen column indicates that the species is preserved at C from the locality, but not from the present investigation.

Table 4 continued.

<b>POLYPORES</b>			
bje adus	Antrodiella hoehnelii (Bres.) Niemelä	2	C
	Antrodiella semisupina (Berk. & Curt.) Ryvarden & Johans.	2	C
	Bjerkandera adusta (Willd.: Fr.) P.Karst.	17	C
cer gilv	Bjerkandera fumosa (Pers.: Fr.) P.Karst.	1	JHC95-197
dat moll	Ceriporiopsis gilvescens (Bres.) Dom.	9	JHC95-251
	Datronia mollis (Sommerf.: Fr.) Donk	6	C

Table 4. continued.

Fom fome	Fomes fomentarius (L.: Fr.) Fr.	38	
fom pini	Fomitopsis pinicola (Swartz: Fr.) P.Karst.	6	C
fus ferr	Fuscoporia ferrea (Pers.) G. Cunn.	5	JHC95-194
gan lips	Ganoderma lipsiensis (Batsch) Atk.	22	C
	Ganoderma pfeifferi Bres.	1	C
ino nodu	Inonotus nodulosus (Fr.) P.Karst.	6	C
isc resi	Ischnoderma resinosum (Schrad: Fr.) P.Karst.	14	C
mer giga	Meripilus giganteus (Pers.: Fr.) P.Karst.	18	C
phy sang	Physisporinus sanguinolentus (Alb. & Schw.: Fr.) Pilát	16	C
phy vitr	Physisporinus vitreus (Pers.: Fr.) P.Karst.	14	JHC94-199
	Polyporus badius (Pers.) Schw.	1	C
pol brum	Polyporus brumalis (Pers.) Fr.: Fr.	3	C
pol cili	Polyporus ciliatus Fr.: Fr.	3	C
pol squa	Polyporus squamosus (Huds.: Fr.) Fr.	3	C
pol tube	Polyporus tuberaster (Pers.: Fr.) Fr.	3	C
pol vari	Polyporus varius (Pers.) Fr.: Fr.	13	C
	Skeletocutis nivea (Jungh.) Keller	2	JHC94-349
tra hirs	Trametes gibbosa (Pers.: Fr.) Fr.	2	C
tra vers	Trametes hirsuta (Wulfen Fr.) Pilát	7	C
	Trametes versicolor (L.: Fr.) Quél.	20	C
	Tyromyces chioneus (Fr.: Fr.) P.Karst.	1	JHC95-093
	Tyromyces wynnei (Berk. & Broome) Donk	1	C
<b>AGARICS</b>			
arm gall	Armillaria gallica Marxm. & K.Korh.	44	C
	Armillaria mellea (Vahl: Fr.) P.Kumm.	1	C
bol reti	Bolbitius reticulatus (Pers.: Fr.) Rick.	7	C
cli diat	Clitocybe diatreta (Fr.: Fr.) P.Kumm.	21	JHC94-166
cli nebu	Clitocybe nebularis (Batsch: Fr.) P.Kumm.	9	C
cli phyl	Clitocybe phyllophilia (Pers.: Fr.) P.Kumm.	3	C
cli hobn	Clitopilus hobsonii (Berk.) P.D.Orton	10	JHC94-424
col buty	Collybia butyracea (Bull.: Fr.) P.Kumm.	8	C
	Collybia fusipes (Bull.: Fr.) Quél.	1	C
	Collybia peronata (Bolt.: Fr.) P.Kumm.	1	
	Collybia tuberosa (Bull.: Fr.) P.Kumm.	2	C
con brun	Conocybe brunnea (J.E.Lange & Kühner)ex Watling	3	JHC94-183
con tene	Conocybe sordida (Kühner)ex Kühner & Watling	1	JHC94-185
cop dome	Coprinus domesticus (Bolt.: Fr.) Gray	3	C
	Coprinus echinosporus Buller	1	JHC94-054
	Coprinus lagopus (Fr.: Fr.) Fr.	1	C
cop mica	Coprinus micaceus (Bull.: Fr.) Fr.	36	C
	Coprinus xanthothrix Romagn.	1	JHC94-426
	Cystolepiota adulterina (F.H.Møller) Bon	1	C
	Cystolepiota hetieri (Boud.) Singer	1	JHC94-187
	Entoloma dichroum (Pers.: Fr.) P.Kumm.	1	JHC94-361
	Flammulaster muricatus (Fr.: Fr.) Watling	1	JHC94-370; JHC95-092
	Galerina nana (Petri) Kühner	1	JHC94-181
gal unic	Galerina unicolor (Fr.) Singer s.l.	24	C
gym junio	Gymnopilus junonioides (Fr.: Fr.) P.D.Orton	3	C
hem cucc	Hemimycena cucullata (Pers.: Fr.) Singer	3	JHC94-167
hen cand	Henningsomyces candidus (Pers.: Fr.) Kuntze	5	C
hyd suba	Hydropus subalpinus (Höhn.) Singer	3	C
hyp fasc	Hypholoma fasciculare (Huds.: Fr.) P.Kumm.	35	C
	Inocybe petiginosa (Fr.: Fr.) Gillet	1	JHC94-186
kue muta	Kuehneromyces mutabilis (Schaeff: Fr.) Sing. & Smith	15	C
	Laccaria laccata (Scop.: Fr.) Berk. & Broome	1	C
lac subd	Lactarius subdulcis (Bull.: Fr.) Gray	4	C
lep aspe	Lepiota aspera (Pers.: Fr.) Quél.	6	C
	Lepiota boertmannii Knudsen	1	JHC95-261
lep cris	Lepiota cristata (Bolt.: Fr.) P.Kumm.	7	C
lep fulv	Lepiota fulvella Rea	4	C
	Lepiota jacobii Vellinga & Knudsen	1	C
	Lepiota ochraceofulva P.D.Orton	2	C
lep flacc	Lepista flaccida (Sow.: Fr.) Pat.	14	C
	Lepista nuda (Bull.: Fr.) Cooke	1	C
mac rhac	Macrolepiota rhacodes (Vitt.) Singer	12	C
mar alli	Marasmius alliaceus (Jacq.: Fr.) Fr.	40	C
	Marasmius cohaerens (Alb. & Schw.: Fr.) Cooke & Quél.	1	C
mar rotu	Marasmius rotula (Scop.: Fr.) Fr.	27	C
mar torq	Marasmius torquescens Quél.	9	C
	Marasmius wynnei Berk. & Broome	1	C
meg plat	Megacollybia platyphylla (Pers.: Fr.) Kotl. & Pouz.	22	C
	Melanophyllum aimatospermum (Bull.: Fr.) Kreisel	2	C

Table 4 continued.

	Melanotus horizontalis (Bull.) P.D.Orton	1	JHC95-191
	Merismodes anomalus (Pers.: Fr.) Singer	2	JHC95-198
	Micromphale brassicolens (Romagn.) P.D.Orton	1	C
myc amic	Mycena amicta (Fr.) Quél.	3	JHC94-053
myc croc	Mycena crocata (Schrad.: Fr.) P.Kumm.	35	C
myc dios	Mycena diosma Kriegsteiner & Schwöbel	6	C
	Mycena erubescens Höhn.	2	C
myc gale	Mycena galericulata (Scop.: Fr.) Quél.	28	C
myc gallo	Mycena galopus (Pers.: Fr.) P.Kumm.	4	C
myc haem	Mycena haematopus (Pers.: Fr.) P.Kumm.	50	C
myc hiem	Mycena hiemalis (Osb.: Fr.) Quél.	3	C
myc peli	Mycena pelianthina (Fr.) Quél.	5	C
	Mycena polygramma (Bull.: Fr.) Gray	2	C
myc pura	Mycena pura (Pers.: Fr.) P.Kumm.	17	C
myc rena	Mycena renati Quél.	11	C
myc rose	Mycena rosea Gramberg	4	C
myc spei	Mycena speirea (Fr.: Fr.) Gillet	4	C
myc tint	Mycena tintinabulum (Fr.) Quél.	7	C
omp epic	Omphalina epichysium (Pers.: Fr.) Quél.	8	C
oud muci	Oudemansiella mucida (Schrad.: Fr.) Höhn.	14	
pan sero	Panellus serotinus (Pers.: Fr.) Kühn.	8	C
pho auri	Pholiota aurivellus (Fr.) P.Kumm.	6	C
pho squa	Pholiota squarrosa (Weigel: Fr.) P.Kumm.	2	JHC95-155
	Pleurotus dryinus (Pers.: Fr.) P.Kumm.	1	C
ple oste	Pleurotus ostreatus (Jacq.: Fr.) P.Kumm.	10	
plu cerv	Pluteus cervinus (Batsch) Singer	37	C
plu chry	Pluteus chrysophaeus (Schaeff.) Quél.	11	C
	Pluteus ephabeus (Fr.: Fr.) Gillet	2	JHC94-362
plu inqu	Pluteus godeyii Gill. ss. Lange	1	JHC94-176
plu nanu	Pluteus inquinilinus Romagn.	4	JHC94-174; JHC95-094
plu phle	Pluteus nanus (Pers.: Fr.) P.Kumm.	5	JHC94-178; JHC94-363
	Pluteus phlebophorus (Dittm.: Fr.) P.Kumm.	22	JHC94-050
plu sali	Pluteus podospileus Sacc. & Cub.	1	JHC94-175
plu semi	Pluteus salicinus (Pers.: Fr.) P.Kumm.	8	C
plu thom	Pluteus semibulbosus (Lasch)Gill.	4	JHC94-171; JHC94-172; JHC95-154
plu umbr	Pluteus thomsonii (Berk. & Broome) Dennis	3	JHC95-153
	Pluteus umbrosus (Fr.) P.Kumm.	5	C
	Psathyrella candolleana (Fr.: Fr.) Maire	1	JHC94-098
	Psathyrella conopilus (Fr.: Fr.) A.Pears. & Dennis	1	JHC94-188
	Psathyrella cortinarioides P.D.Orton	1	JHC94-191
	Psathyrella fusca (Schum.) A.Pears.	1	JHC94-192
	Psathyrella lacrymabunda (Bull.: Fr.) Moser	1	
psa obtu	Psathyrella obtusata (Pers.: Fr.) A.H.Smith	19	JHC94-425
psa pilu	Psathyrella piluliformis (Bull.: Fr.) P.D.Orton	8	JHC95-148; JHC95-212
	Psathyrella populina (Britz.) Kits.	1	JHC94-365
psa rost	Psathyrella pygmaea (Bull.: Fr.) Singer	1	JHC94-193
	Psathyrella rostellata Örstadius	4	JHC94-194; JHC94-195
	Psathyrella spadiceogrisea (Schaeff.) Maire	1	JHC94-364
	Psathyrella tephrophylla (Romagn.) Bon	1	C
pse cyat	Pseudoclitocybe cyathiformis (Bull.: Fr.) Singer	9	C
	Ramicola centunculus (Fr.: Fr.) Wattl.	1	JHC94-368
	Resupinatus trichotis (Pers.) Singer	1	C
	Ripartites tricholoma (Alb. & Schw.: Fr.) P.Karst.	1	C
str cyan	Stropharia cyanea (Bull.) Tuomikoski	3	C
	Tricholoma lascivum (Fr.) Gillet	1	C
	Tubaria furfuracea (Pers.: Fr.) Gillet	2	C
	Volvariella hypopithys (Fr.) Shaffer	1	JHC95-086
xer radi	Xerula radicata (Rehman: Fr.) Dörfelt	10	C
<b>OTHER BASIDIOMYCETES</b>			
	Auricularia auricula-judae (Bull.: Fr.) Wettst.	1	C
cal corn	Calocera cornea (Batsch: Fr.) Fr.	6	
	Dacryomyces stillatus Nees: Fr.	1	JHC95-200
exi glan	Exidia glandulosa (Bull: Fr.) Fr.	4	JHC95-199
exi nucl	Exidia nucleata (Schwein.) Burt.	3	C
her cora	Hericium coralloides (Scop.: Fr.) Pers.	4	C
	Lycoperdon echinatum Pers.: Pers.	1	C
lyc perl	Lycoperdon perlatum Pers.: Pers.	13	C
lyc pyri	Lycoperdon pyriforme Schaeff.: Pers.	36	C
mut cani	Mutinus caninus (Huds.: Pers.) Fr.	3	
pha impu	Phallus impudicus L.: Pers.	5	
phl fagi	Phleogena faginea (Fr.: Fr.) Link	11	C
ram stri	Ramaria stricta (Pers.: Fr.) Quél.	15	C
sch comm	Schizophyllum commune Fr.: Fr.	5	C

Table 4 continued.

tre mese	Tremella mesenterica Retz: Fr.	7	C
<b>SLIME MOULDS</b>			
arc cine	Arcyria affinis Rostr.	1	JHC95-260
arc denu	Arcyria cinerea (Bull.) Pers.	3	C
arc denu	Arcyria denudata (L.) Wettst.	4	C
arc fruc	Arcyria incarnata (Pers.) Pers.	2	JHC94-099
cer fruc	Ceratiomyxa fruticulosa (Müll.) Macbr.	24	C
cri argi	Comatricha alta Preuss	1	JHC94-002
cri argi	Comatricha nigra (Pers.) Schroet.	2	JHC94-100
ent lyc	Cribaria argillacea (Pers.) Pers.	5	JHC94-082
ful sept	Enteridium lycoperdon (Bull.) Farr.	18	JHC94-361
ful sept	Fuligo septica (L.) Wiggers	19	C
lam arcy	Lamproderma arcyrioides (Sommerf.) Rost.	3	JHC95-258; JHC95-259
lyc epid	Lycogala epidendrum (L.) Fr.	35	C
phy cine	Metarichia floriformis (Schw.) Nann.-Brem.	1	C
ste axif	Metarichia vesparium (Batsch) Nann.-Brem.	2	JHC95-257
ste fusc	Physarum cinereum (Batsch) Pers.	1	JHC94-094
ste typh	Stemonitis axifera (Bull.) Macbr.	7	C
ste typh	Stemonitis fusca Roth	6	C
ste typh	Stemonitopsis hyperopta (Meylan) Nann.-Brem.	1	JHC94-090
ste typh	Stemonitopsis typhina (Wiggers) Nann.-Brem.	6	C
tri scab	Sympylocarpus amaurochaetoides Nann.-Brem.	2	JHC94-101
tri vari	Trichia contorta (Ditm.) Rost.	1	JHC95-256
tri vari	Trichia persimilis P.Karst.	1	C
tri vari	Trichia scabra Rost.	10	JHC94-429
tri vari	Trichia varia (Pers.) Pers.	29	JHC94-427; JHC94-428
	Tubifera ferruginosa (Batsch) J.F.Gmel.	1	C

found to be significantly more species-rich than both older and younger logs (ANOVA on ranks; Newman-Keuls test,  $P<0.05$ ).

With respect to slime moulds, the largest diversity were found somewhat later in the succession, Fig. 2. Logs 12–20 years old were found to be most species-rich (ANOVA on ranks; Newman-Keuls test  $P<0.05$ ).

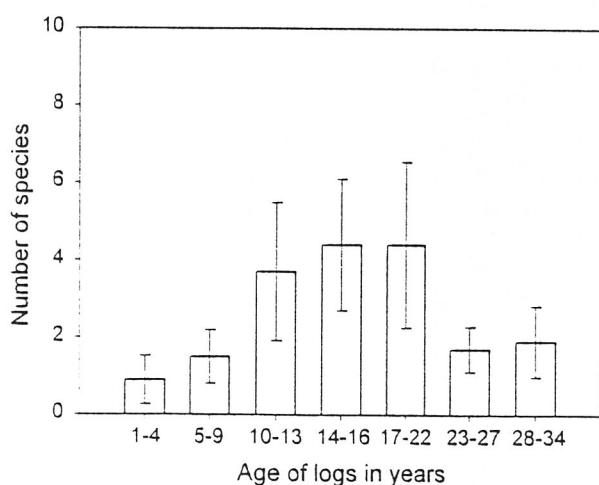


Fig 2. Average number of slime mould species per log in the seven age classes. The bars represent 95% confidence limits. Number of logs = 10 in each class.

Table 5 shows correlation coefficients between the investigated variables. Basically three groups of intercorrelated variables are distinguishable. The first group consists of decay related variables (log age, decay class, bark cover and soil contact), the second of log size and outline variables (dbh, fractures, bole forks) whereas the third, complex set, is constituted by variables related to microclimatic conditions (distance to edge, sun exposure, wind exposure, soil humidity, moss cover).

#### Ordination

After a visual inspection of the initial DCA scattergrams, one distinctly outlying log was omitted from further analysis. Outliers are potentially interesting but may cause serious distortions overruling the general patterns of the ordination. The excluded log was newly dead, but with extensively decayed parts. Hence the associated fungi was a mixture of species associated with initial phases of decay and typical late-stage decayers, which may explain why the log behaved aberrantly.

Table 6 shows the overall properties of the full data sets after exclusion of the outlier, as well as the properties of the three subsets. Table 7 shows the properties of the DCA-axes in the ordination of the full set.

	Dbh.	Bole forks	Frac- tures	Log age	Decay stage	Bark cover	Soil- contact	Dist. to edge	Wind exposure	Sun exposure	Soil- humidity	Moss cover	Eutypa cover	Plant cover	Plant diversity	Floristic deviance	Decay rate
Dbh.	1																
Bole forks	0.41****	1															
Fractures	0.35***	0.46****	1														
Log age	0.03	-0.18	-0.02	1													
Decay stage	0.12	-0.17	0.06	0.80****	1												
Bark cover	-0.03	0.15	0.01	-0.77****	-0.81****	1											
Soil-contact	0.16	-0.01	0.19	0.38****	0.50****	-0.43****	1										
Dist. to edge	0.22**	0.06	0.06	-0.07	-0.02	0.06	0.07	1									
Wind exposure	0.19*	0.04	0.12	-0.17	-0.09	0.12	0.02	0.51****	1								
Sun exposure	0.12	0.04	0.14	0.19*	0.25*	-0.19	0.16	0.13	0.31**	1							
Soil-humidity	0.07	-0.01	0.12	0.08	0.16	-0.09	0.20*	-0.09	0.20*	0.14	1						
Moss cover	0.24*	0.17	0.32**	0.18	0.19	-0.19	0.25*	0.12	0.35***	0.26*	0.34**	1					
Eutypa cover	-0.04	0.99	0.13	0.21*	0.11	-0.18	-0.08	-0.03	-0.11	0.05	-0.13	0	1				
Plant cover	0.01	-0.07	-0.03	0	-0.06	0.02	0.03	0.21*	0.19	0.05	-0.01	-0.02	0.05	1			
Plant diversity	-0.12	-0.01	-0.14	-0.08	0.21*	0.1	-0.15	-0.23*	-0.01	-0.11	0.08	0.08	-0.01	0.08	1		
Floristic deviance	-0.06	-0.07	-0.07	-0.12	0.04	-0.08	0.22*	0.1	-0.04	0.09	0.14	0.01	0.08	0.63****	1		
Decay rate	0.07	0.06	0.21*	-0.21*	0.03	0.06	0.28*	0.15	0.13	0.09	0.07	-0.07	-0.27**	-0.07	-0.29**	0.21*	1

Table 5. Kendall rank correlations between variables. \*\*\*\* P <0.0001; \*\*\* P= 0.0001 - <0.001; \*\* P= 0.001 - <0.01; \* P= 0.01 - <0.05.

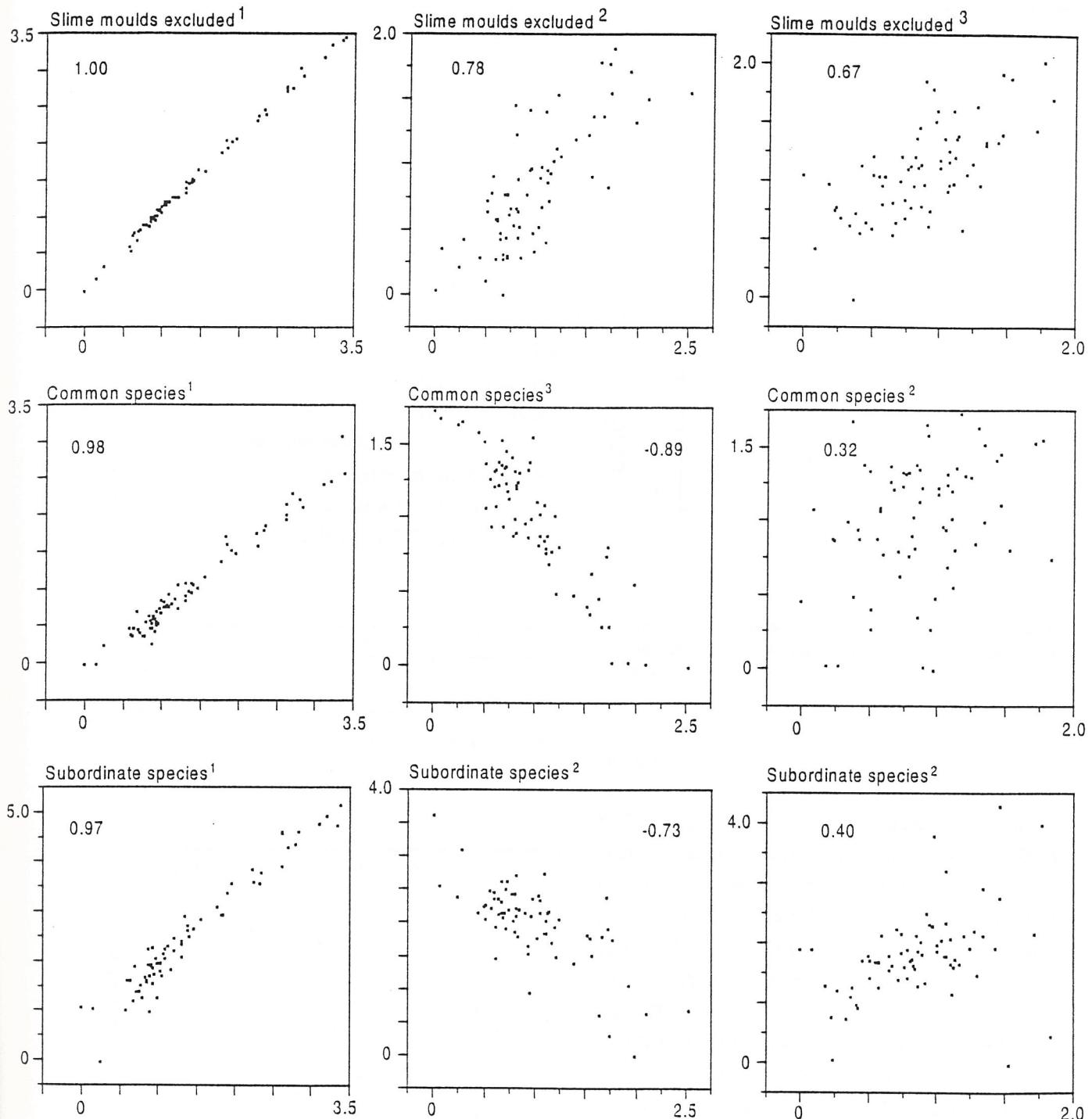
DCA<sup>1</sup>DCA<sup>2</sup>DCA<sup>3</sup>

Fig. 3. Evaluation of DCA coenoclines by ordinations of subsets. The three columns represent scatterplots of the three DCA-axes of the full set against the best correlated of the DCA-axes within the three subsets. Coefficients of the Pearson product moment correlation are indicated within the plots. Axes are scaled in SD units.

	Full set	Slime moulds excluded	Common species	Subordinate species
Number of samples (logs)	69	69	69	69
Number of species	179	164	96	146
Number of non-zero data items	2041	1957	1794	1082

Table 6. Properties of the full data set and of the three investigated subsets.

A scatterplot matrix of DCA1-3 in the full data set against the best correlated axes in three sub-sets (Fig. 3) reveals DCA1 to be very consistently expressed. Also DCA2 is reproduced fairly well in the subset ordinations, whereas DCA3 is clearly expressed only in the subset obtained by excluding slime moulds.

Figs. 4 and 5 shows log based plots of DCA1 against DCA2 and DCA3 in the full data set. A pronounced 'tongue' (cf. Økland 1990) relating to DCA2 is evident

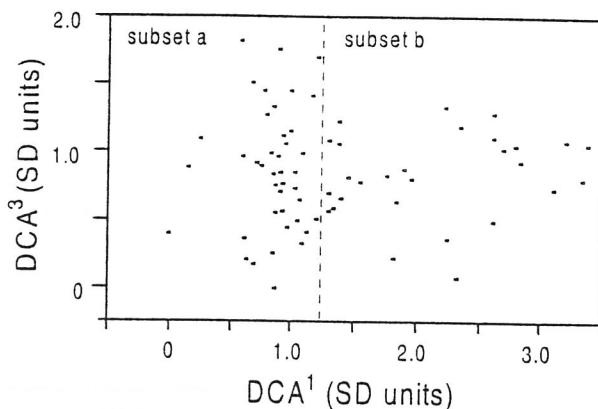


Fig. 5. DCA ordination diagram of the logs on axes 1 and 3. The dotted line shows the cut-level dividing subset a from subset b.

Table 7. Properties of the DCA-axes in the ordination of the full data set.

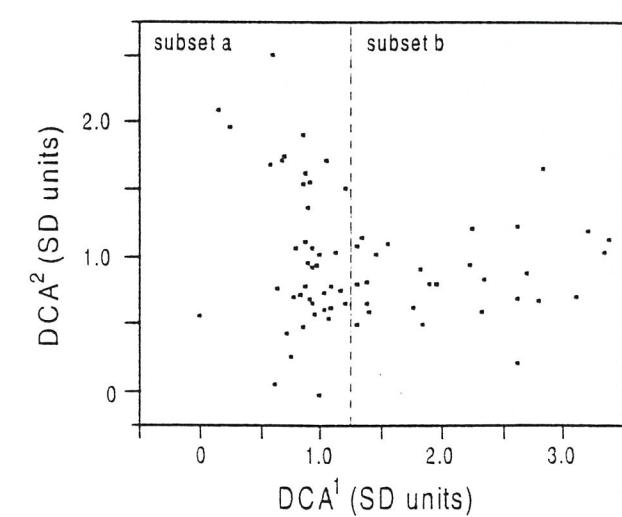


Fig. 4. DCA ordination diagram of the logs on axes 1 and 2. The dotted line shows the cut-level dividing subset a from subset b.

along DCA1. The nature of this was evaluated by departing the samples (logs) in two sub-sets as suggested by Økland (1990), followed by a careful examinations of both subsets. A cut level at 1.25 SD units along DCA1 was found to be superior in the attempt to interpret DCA2 from the recorded variables. Logs with DCA1 scores below 1.25 SD units are referred to as subset a, whereas subset b contains logs with DCA1 scores above 1.25 SD units.

If only subset b is considered, correlations with the variables correlating with DCA2 in the full data set are mostly non-significant (Table 8). This indicate that the tongue represents a true structure in the data set and that DCA2 express an ecological gradient that is only expressed at the lower end of DCA1 (Økland 1990, type c.). Accordingly logs with high DCA1 scores represent noise in the interpretation of DCA2.

A separate ordination of the subsets, as suggested in similar cases by Peet (1980) and Økland (1990), was not

	Subset a. (n=41)	Subset b (n=28)	Full set (n=69)
Log age	0.5156****	-0.0770	0.2346**
Decay rate	-0.4976****	-0.0193	-0.3544***
Wind exposure	-0.4661***	-0.2320	-0.3647****
Dist. to dge	-0.3862***	-0.1853	-0.3134***
Fractures	-0.3768**	0.1144	-0.2162*
Other plants	0.3447**	0.0176	0.1963
Bole forks	-0.3059*	-0.0156	-0.2337*
Decay stage	0.3004*	-0.1210	0.1413
Bark cover	-0.2915*	0.1089	-0.1467
Ane nem	-0.2749*	-0.1490	-0.2150*
Sun exposure	-0.2649*	-0.3046*	-0.2290*
Floristic deviance	0.2613*	0.0958	0.1830*
Plantdiv	0.2548*	0.1902	0.2348**

Table 8. Kendall rank correlations for DCA<sup>2</sup> in subset a and subset b. compared to the full data set. Only variables with significant correlation quotients in a set are shown. \*\*\*\* P < 0.0001; \*\*\* P = 0.0001 - < 0.001; \*\* P = 0.001 - < 0.01; \* P = 0.01 - < 0.05.

	DCA1	DCA1#	DCA2	DCA2#	DCA3	DCA3#
DCA1	1.00	1.00	-0.08	-0.17	0.00	-0.04
DCA2	-0.08	-0.17	1.00	1.00	-0.04	-0.04
DCA3	0.00	-0.04	-0.04	-0.10	1.00	1.00
Decay stage	-0.68****	-0.33**	0.14	0.30*	-0.11	-0.24
Log age	-0.66****	-0.27*	0.23*	0.52****	-0.10	-0.17
Bark cover	0.65****	0.30*	-0.15	-0.29*	0.05	0.12
Soil contact	-0.41****	-0.27*	-0.09	-0.05	-0.02	0.07
Moss cover	-0.23*	-0.04	-0.12	-0.12	0.12	0.22
Sun exposure	-0.22*	0.10	-0.23*	-0.26*	0.02	-0.04
Soil humidity	-0.20*	-0.36**	-0.07	-0.04	0.09	0.16
Decay rate	-0.01	-0.17	-0.35***	-0.50****	0.01	-0.04
Wind exposure	0.00	0.02	-0.36****	-0.45***	0.05	0.14
Dist. to edge	-0.00	0.01	-0.31***	-0.39***	-0.07	0.01
Fractures	-0.06	0.08	-0.22*	-0.38***	0.12	0.12
Other plants	-0.08	-0.33**	0.20*	0.34**	0.11	0.16
Bole forks	0.14	0.20	-0.23*	-0.31*	-0.09	-0.07
Ane nem	-0.00	0.10	-0.22*	-0.27*	0.00	-0.04
Ran fic	-0.17	-0.23	0.14	0.12	0.37***	0.43***
Ane ran	0.03	-0.01	0.13	0.16	0.28**	0.41***
Plant diversity	0.07	0.01	0.23*	0.25*	0.26**	0.42***
Floristic deviance	0.04	-0.03	0.18*	0.26*	0.24**	0.38**
Plant cover	-0.06	-0.06	-0.03	-0.03	0.23*	0.25*
Mer per	0.06	-0.05	0.19	0.16	0.10	0.25*

Table 9. Kendall rank correlations between DCA axes and log variables in the full set and subset b (#) respectively. Only variables with significant correlation quotients in a set are shown. \*\*\*\* P < 0.0001; \*\*\* P = 0.0001 - < 0.001; \*\* P = 0.001 - < 0.01; \* P = 0.01 - < 0.05.

found to increase the informative power, compared to a separate analysis of subset a of the full ordination. Probably because of a decrease in the amount of informative data in the diminished data set. Table 9 shows correlation coefficients between DCA axes and variables, both for the full data set and subset a.

DCA1 correlates strongly with all decay-associated variables, but below 1.25 SD units (subset a) the importance of log age, bark cover and soil contact ceases. Decay stage is however still distinctly expressed. The same is the case for soil humidity, in contrast to the case in the full data set.

DCA2 correlates both in the full data set and in sub-set 1 significantly with several variables, most notably wind exposure, distance to edge and decay rate. In subset a, log age is the best correlated variable, in contrast to the case in the full data set.

DCA3 is associated mainly with a turn-over in the floristic variables (plantcover, plant diversity and floristic deviation). More specific the frequency of *Anemone ranunculoides* and *Ranunculus ficaria* ssp. *biflora* is positively correlated with the axis.

### Modelling

The strong intercorrelation between several variables in the material (Table 5) obscures the structures of the complex environmental gradients behind the axes, and prevents a quantitative interpretation of these, since the

correlation quotients are in no way additive. Multiple regression was applied in an attempt to overcome these shortcomings. Properties of the resulting models are shown in Table 10 and Figs. 6 and 7. Due to the bad recover of DCA3 in the ordination of subsets, models of this axis are not presented.

The model of DCA1 is dominated by a single variable, log age, which explain as much as 84% of the sample variation. Inclusion of other decay related variables did not strengthen the model, but a weak improvement was obtained by including distance to edge and soil contact. The model of DCA2(subset a) is more complex and has, compared with the model of DCA1, a lower coefficient of determination. The model contains four variables which all contribute significantly in the explanation of sample variation.

The attempt to model for DCA3 was not very successful for the full data set. Obtained models were not very powerful ( $r^2$  about 0.3), or appeared more or less spurious by the inclusion of variables which independently are not correlated with the axis. Therefore a model based on subset a is presented. The model has a even lower coefficient of determination than the model of DCA2. Plant diversity is the variable most strongly expressed in the model, but soil contact and the 'dummy variable' soil type contributes as well. With respect to the latter variable the model implies that logs with high DCA3 scores tends to be situated on the richer soil types, viz. clay tills

Terms	Res. Df.	Sum of Squares	Prob. (F)	$R^2$
DCA <sup>1</sup>				
log age (polynomial, 2. degree)	67	362726	<0.0001	0.83
+ edge (polynomial, 2. degree)	65	15924	<0.0001	0.87
+ soil contact	64	7529	0.003	0.89
DCA <sup>2</sup>				
decay rate	68	44335	0.0019	0.33
+ decay stage (polynomial, 2. degree)	66	26267	0.0113	0.53
+ wind exposure	65	12564	0.0014	0.62
- floristic deviance	64	10428	0.0046	0.70
DCA <sup>3</sup>				
plant diversity	68	22815	<0.0001	0.28
- soil type	67	13641	0.0048	0.44
+ soil contact	66	5442	0.032	0.51

Table 10. ANOVA's comparing successively more complicated multiple regression models predicting DCA<sup>1-3</sup>. All values except  $R^2$  and residual degrees of freedom (Res. Df) refer to the added variable.

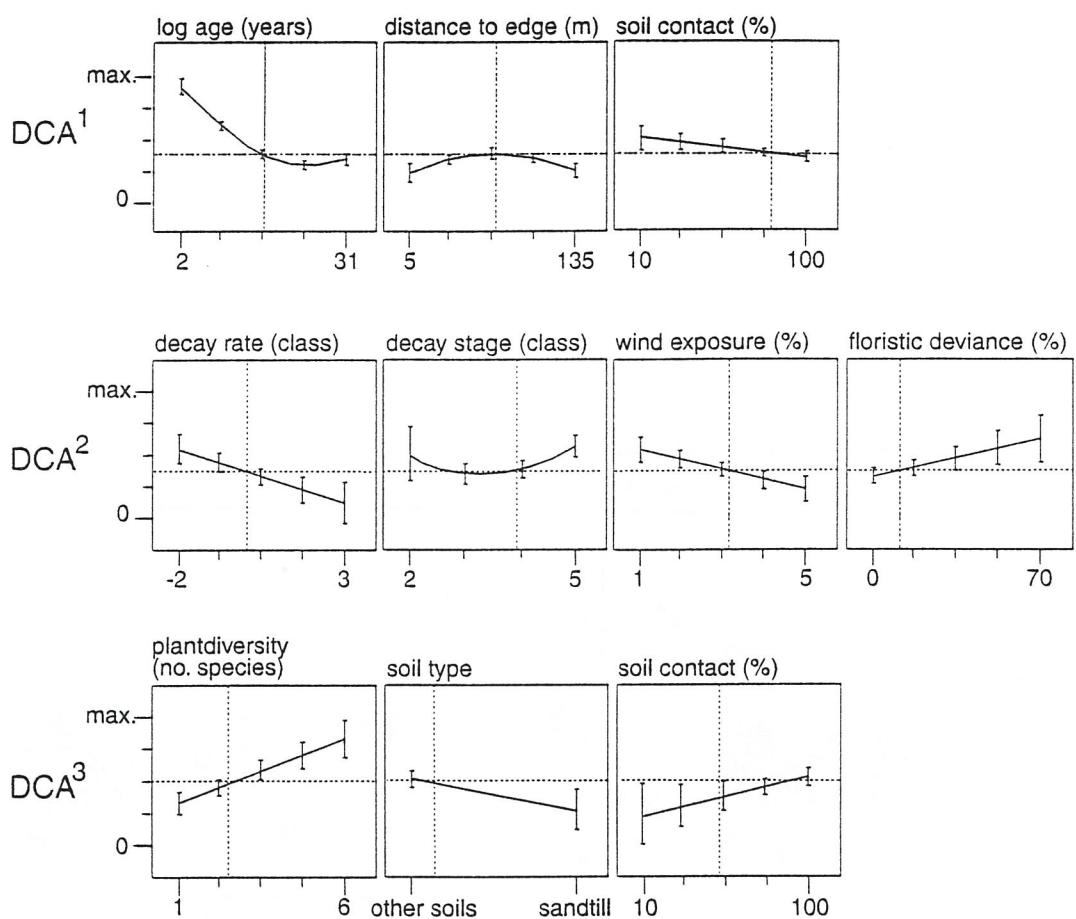


Fig. 6. Prediction profiles showing the roles of individual variables in the multiple regression models obtained for DCA<sup>1-3</sup>.

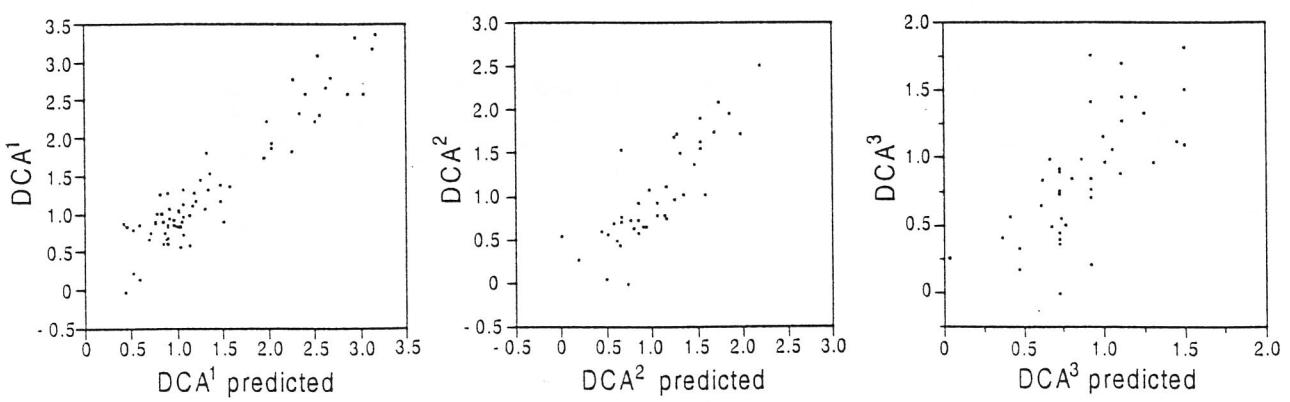


Fig. 7. Plots of the three DCA-axes against the respective fitted multiple regression models. Axes are scaled in SD units.

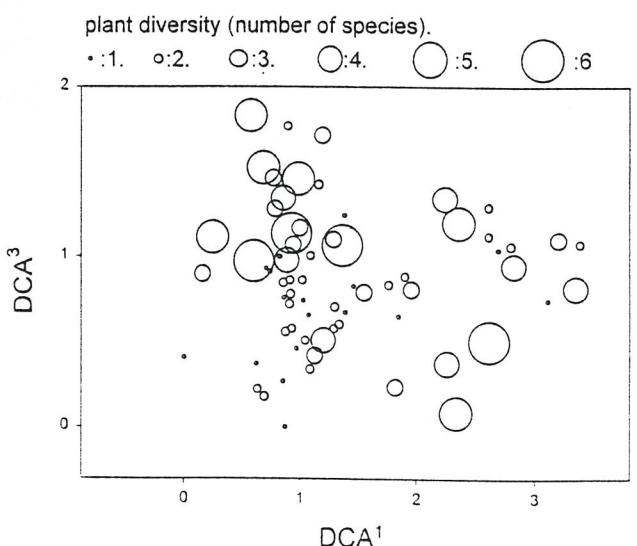
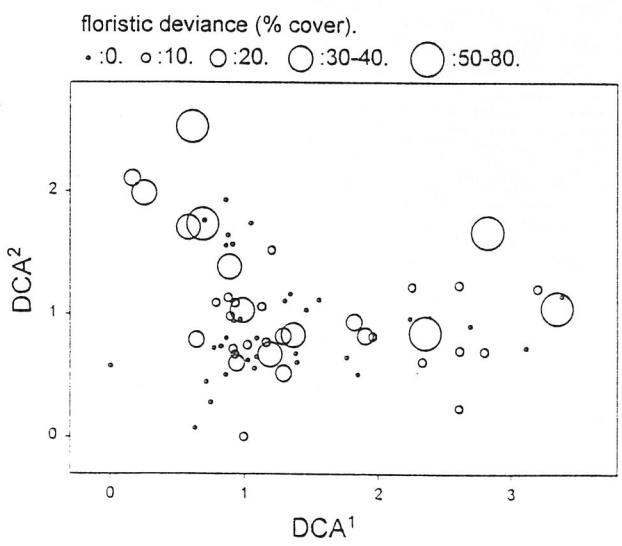
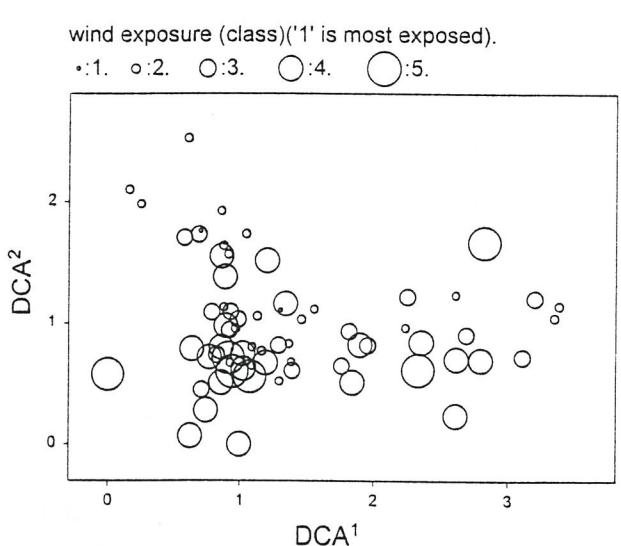
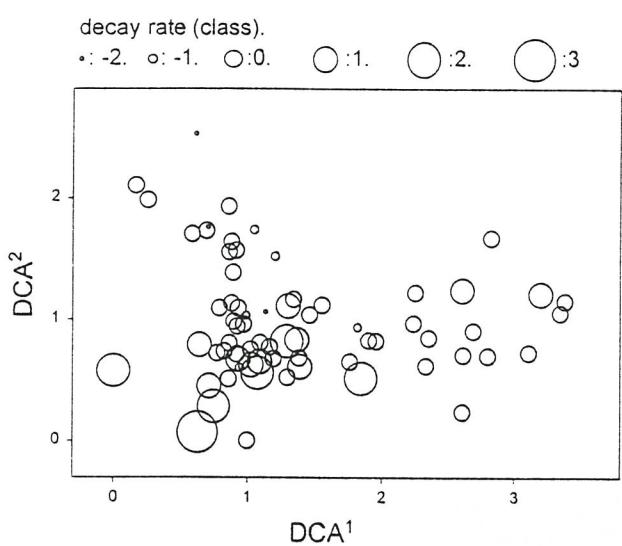
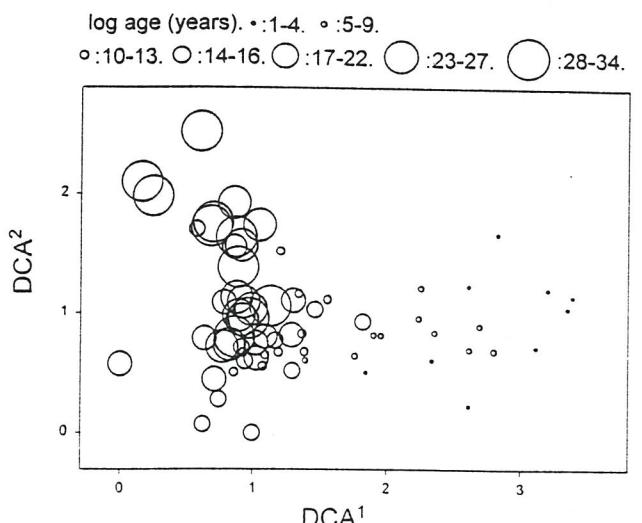
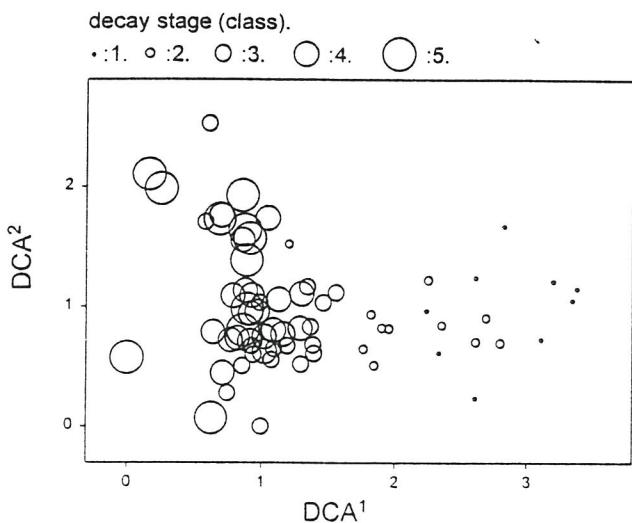


Fig. 8. Biplots showing the log specific scores in the DCA ordination diagrams with respect to various key-variables. Axes are scaled in SD units.

and lacustrine deposits, whereas logs on sandy tills tends to have low DCA3 scores.

#### Distribution of species along DCA-axes

Figs. 9 and 10 shows the distribution of species in the log based DCA ordination space. The pattern is obviously difficult to overview, but some help for the interpretation may be gained from the correlation quotients given in Table 11, and from the accompanying overlay plots (Fig. 11).

The diversity of slime moulds is negatively correlated with DCA1, and a distinct turnover in fungal morpho-groups is evident, except for the corticioid fungi. Polypores and pyrenomycetes are very strongly positively correlated with the axis, whereas agarics are negatively correlated. Weaker correlations are found for discomycetes and other basidiomycetes.

DCA2 correlates strongly negative with both fungal and slime mould diversity. Only within the corticioid fungi the tendency is not evident, and the relative contribution

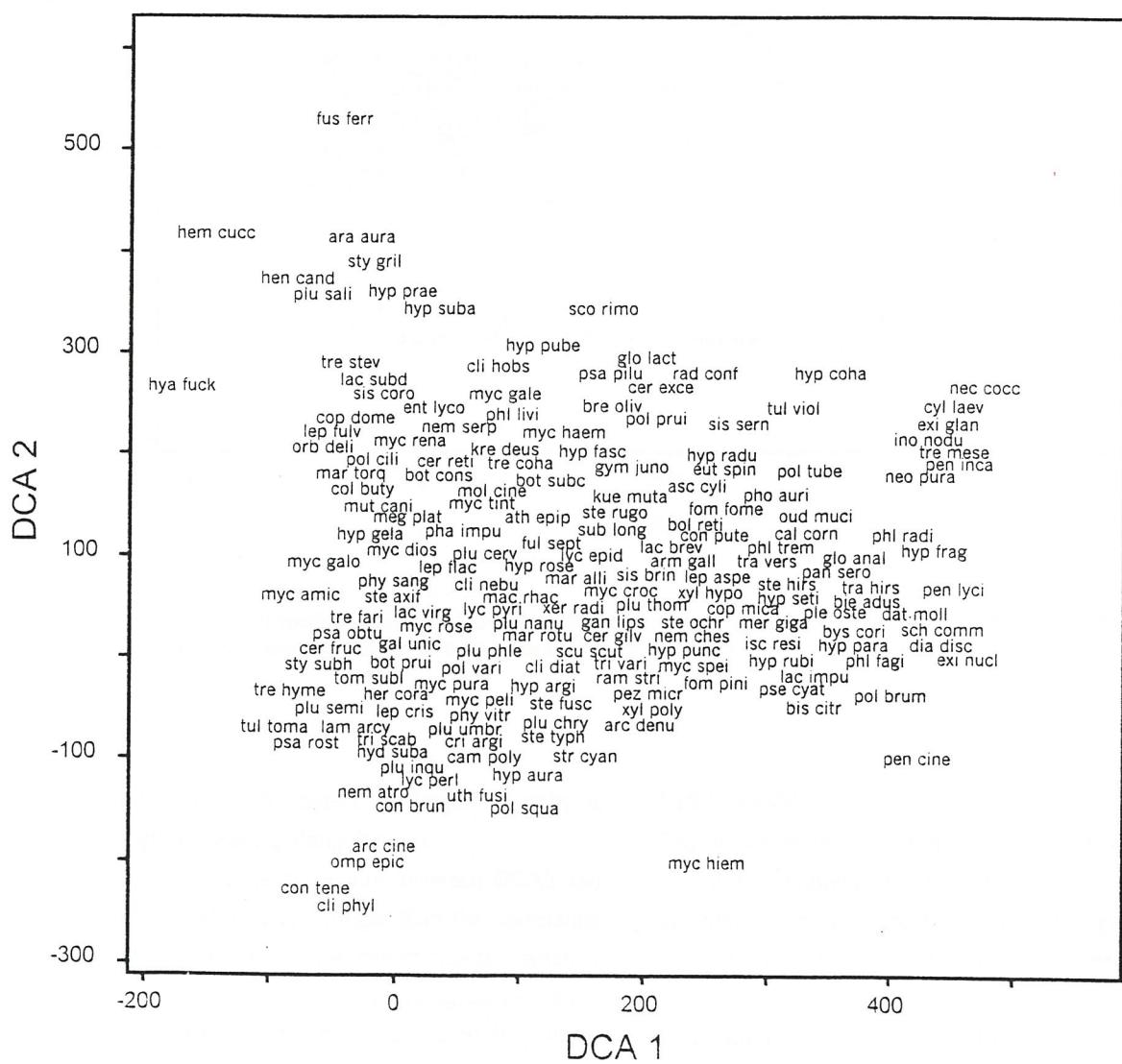


Fig. 9. The distribution of species in the DCA1/DCA2 coenoplane. Names of species are abbreviated according to Table 4. Some species have been moved slightly to give place for all names. Axes are scaled in SD units x 100.

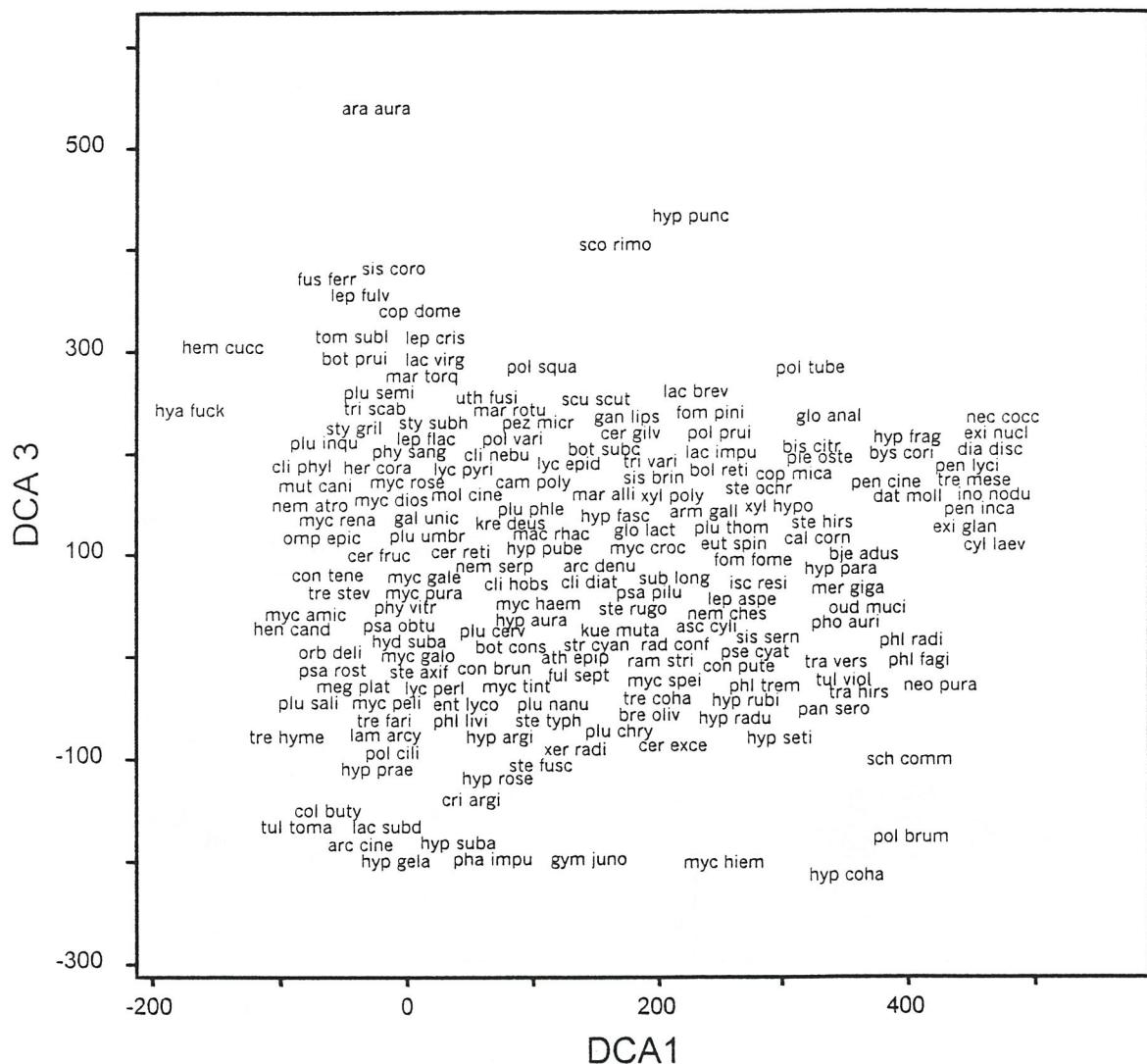


Fig. 10. The distribution of species in the DCA1/DCA2 coenoplane. Names of species are abbreviated according to Table 4. Some species have been moved slightly to give place for all names. Axes are scaled in SD units x 100.

of corticoid fungi with respect to fungal diversity is hence strongly increasing along the axis.

It is noteworthy that the correlation between DCA2 and overall species richness is stronger than the correlation between DCA2 and any of the environmental variables (Table 9). Apart from describing an ecological gradient the axis is in other words representing a gradient in diversity.

DCA3 correlates positively with the relative importance of discomycetes, but apart from this no evident trends are expressed along the axis.

## DISCUSSION

The presented results clearly show that DCA1 responds to a decay gradient, which is dominant in determining the community structure of the studied logs. Compared to DCA2 and DCA3, DCA1 express considerably more sample variation (Table 7), the axis is highly insensitive to modifications of the data set (Fig. 3) and has a very clear model structure (Table 10, Figs. 6 and 7).

Several previous investigations based on registrations of fungal sporocarps have yielded comparable results. A distinct species turnover during decay has been described

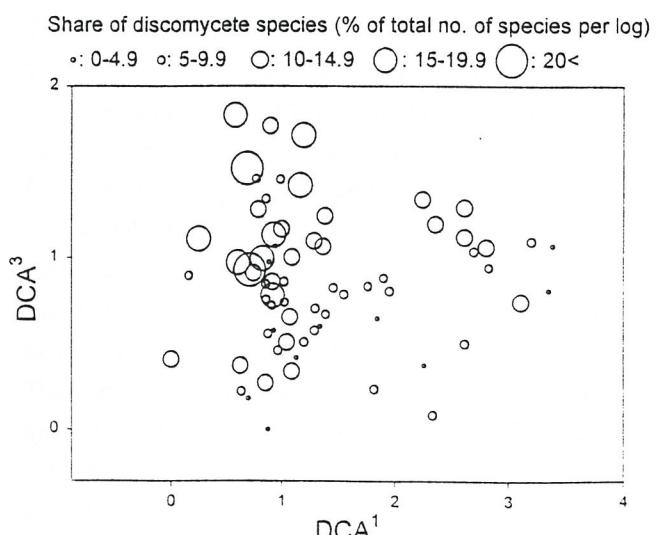
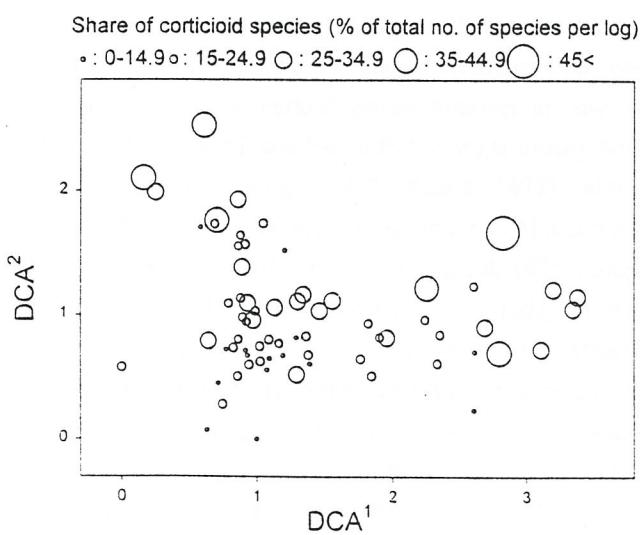
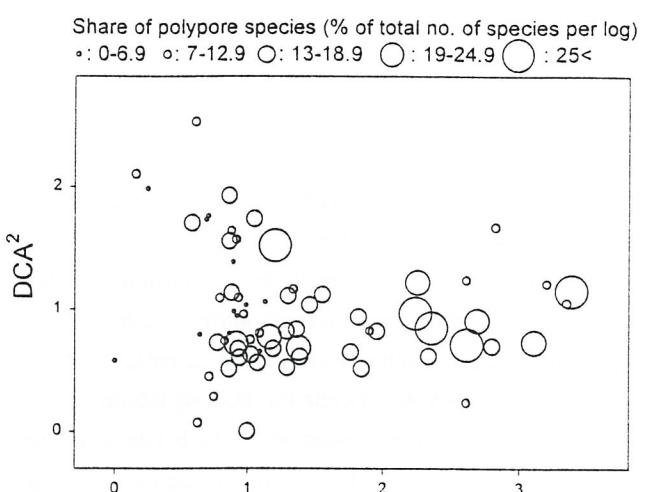
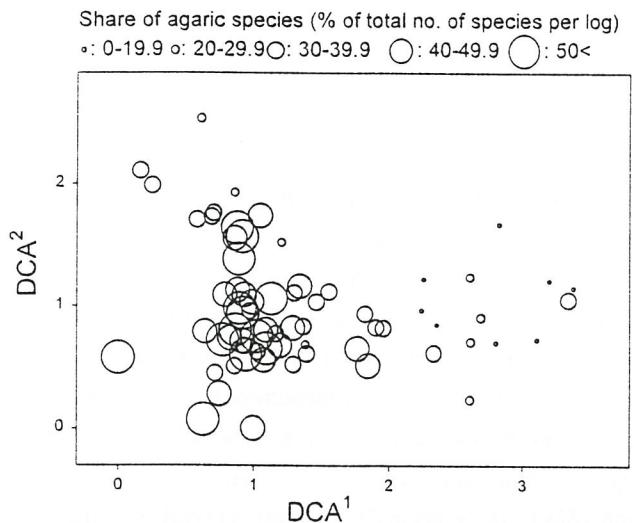
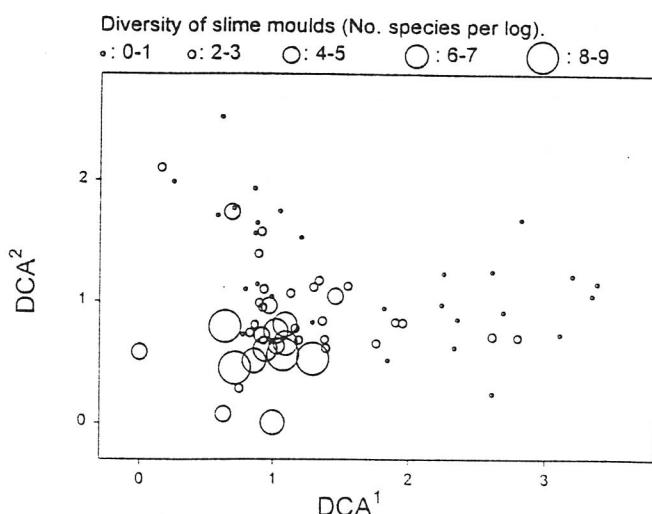
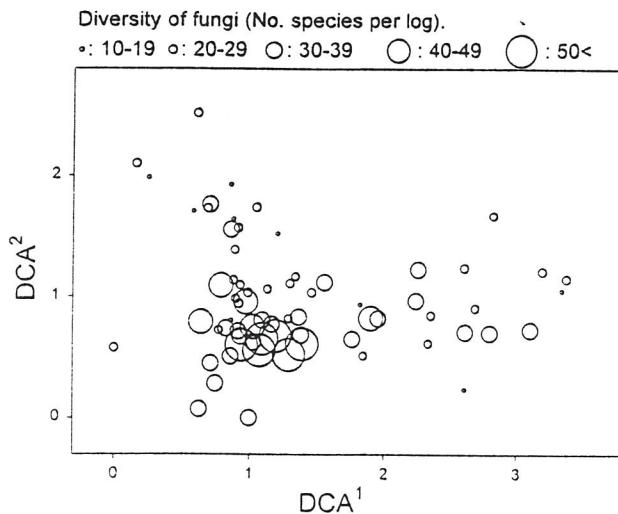


Fig. 11. Biplots showing the species diversity and the relative share of various morpho-groups in the DCA ordination diagrams. Axes are scaled in SD units.

	DCA1	DCA1#	DCA2	DCA2#	DCA3	DCA3#
<b>Species number of</b>						
Fungi	-0.00	0.20	-0.39****	-0.47****	-0.02	0.04
Slime moulds	-0.24**	0.18	-0.40****	-0.58****	-0.06	-0.03
<b>Share of</b>						
Pyrenomycetes	0.29***	0.11	0.05	0.08	0.08	0.04
Discomycetes	-0.19*	-0.13	-0.07	-0.07	0.30***	0.29**
Agarics	-0.40****	0.09	-0.11	-0.20	-0.10	-0.09
Corticoid fungi	0.11	-0.16	0.37****	0.43****	-0.19	-0.28
Polypores	0.37****	0.22*	-0.10	-0.10	0.02	0.05
Other basidiomycetes	0.24**	-0.14	-0.20*	-0.30**	-0.024	-0.00

Table 11. Kendall rank correlations between various species groups and DCA axes in the full dataset and in subset a(#). The two first rows consider raw species numbers, the others relative shares. \*\*\*\* P<0.0001; \*\*\* P= 0.0001 - <0.001; \*\* P= 0.001 - <0.01; \* P= 0.01 - < 0.05.

repeatedly from beech logs (Lange 1992, Willig & Schlegte 1995) and stumps (e.g. Kreisel 1961, Andersson 1997). Based on DCA, Renvall (1995), Høiland & Bendiksen (1996) and Lindblad (1997) even recognized a decay gradient to be the dominant factor in structuring communities of decomposer fungi on conifer logs.

On the mycelial level a distinct species turnover during decay has been described from studies of cut beech logs (Coates & Rayner 1985a-c, Chapela et al. 1988) and from studies of ash, oak and beech branches (Boddy, Bardsley & Gibbon 1987, Chapela & Boddy 1988c, Griffith & Boddy 1990). The understanding of wood decay as involving a marked species turnover in time, is thus well supported, and has in fact a longer history than indicated above (Shigo 1967, Käärik 1975). Many authors have described the process in terms of a succession (e.g. Kreisel 1961, Shigo 1967, Käärik 1975, Lange 1992, Renvall 1995), but most recently Boddy (1992) has argued against using this term deliberately. Boddy (1992) argues that the word succession implies an ordered stepwise change, whereas the true process of wood decay is complex and multidimensional, with a diverse array of 'optional' pathways. However, complexity and multidimensionality are features of many accepted successional processes (Walker & Chapin 1987) and I see no reason not to use the term also in the present case.

The gradient expressed along DCA2 is complex, as re-

flected by the axis/variable correlations (Table 9) and by the regression model, which includes several almost equally contributing variables (Table 10). In fact several almost equally strong models were obtained for the axis, based on other combinations of variables than in the presented model (results not shown). Accordingly, the presented model shall not be taken too literally in respect to specific included variables. Rather it implies that several different groups of variables should be considered in an ecological interpretation of DCA2.

One of two ruling parameters of the gradient seem to be the decay rate, as this variable compared to the other variables in the models has the strongest individual coefficient of determination. The variable seems to account for most of the explanatory power of log age, which is the variable, which has the most significant correlation coefficient to DCA2, when tested singly (Table 9). Log age and decay rate are naturally autocorrelated, since slowly decayed logs obtain older ages than fast decayed. The clear effect of decay stage in the model however implies that the substitution of log age by decay rate is not perfect, maybe because of the rather non-precise nature of the latter variable.

Another obvious key parameter of the gradient is related to the amount of externally induced microclimatic fluctuations. The parameter is in the model represented by wind exposure, which seems to account for the effects of

the variables distance to edge and sun exposure, which both are significantly correlated with DCA2, when tested singly (Table 9). Most wood decay fungi have optima at intermediate moisture levels, but considerable differences exist between different species of fungi; both in respect to their optima and tolerances (Boddy 1983, Boddy et al. 1985, Chapela & Boddy 1988b, Griffith & Boddy 1991b). Unfortunately, the effects of a fluctuating microclimate, as expressed along DCA2 has never been investigated. However, a strongly fluctuating microclimate must be considered a distinct stress factor, implying an altered community structure and a decreased decay rate.

The role of the last variable in the model, floristic deviation, is more difficult to interpret. The variable might reflect a long-term response to certain microclimatic or soil

related conditions, favouring other plant species than the very tolerant *Anemone nemorosa*.

DCA3 adds little to the description of overall community development of the studied logs. The model of the axis (Fig. 7; Table 10) is not very powerful and the low eigenvalue and gradient length scores (Table 7) indicate that the axis may partly represent noise or polynomial distortions (Økland 1990), an indication which is supported by the bad recover of the axis in the ordinations of subsets (Fig. 3). The axis seems to respond to a weak soil richness gradient, and it is somewhat surprising that the axis is so weakly expressed, as soil conditions have been found to influence the occurrence and fitness of several wood decaying fungi (Eriksson et al. 1973-1988, Rayner & Boddy 1988, Kuyper & de Vries 1990, Abdalla & Boddy 1996). Probably the gradient would be

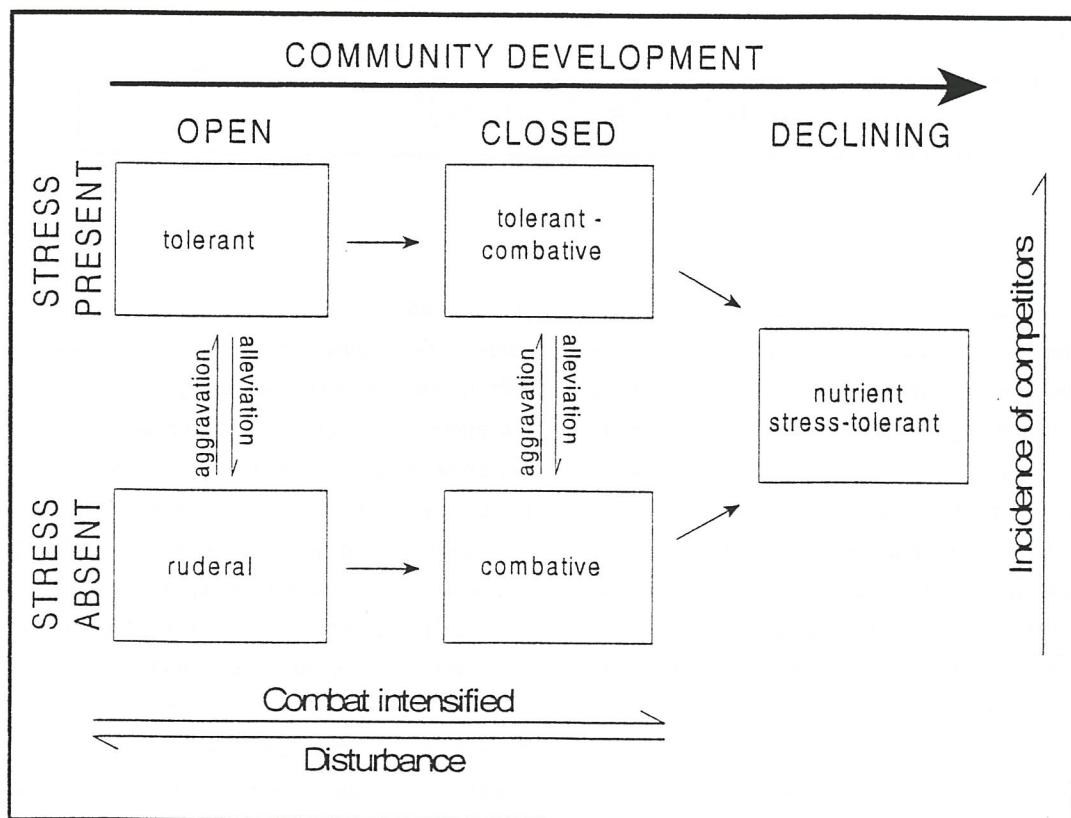


Fig. 12. A diagram of fungal community development pathways, slightly modified from Rayner & Webber (1984). The diagram describes the development from totally unoccupied woody resources to the left to a declining community stage, characterized by nutrient stress to the right.

more clearly expressed if logs distributed across a bigger spectre of soil types, than present at the locality, were studied. The increasing share of discomyctes along DCA3 (Table 11) is intriguing, but shall not be taken to literally in the light of the unreliability of the gradient.

#### *Overall community development*

A generalised diagram of fungal community development in organic substrata was presented by Cooke & Rayner (1984) and has since been simplified and discussed in respect to decaying wood (Rayner & Webber 1984, Rayner & Boddy 1988, Boddy 1992). The diagram, Fig. 12, is based on the theories of ecological strategies of Grime (1974, 1979) and invokes two ruling gradients; the first representing an increasing incidence of competition as decay proceeds, the second representing stress, e.g. imposed by an extreme microclimatic re-

gime. If DCA2 is accepted as representing a microclimatic stress gradient, the plot of DCA1 against DCA2 (Fig. 4) equals this model in a modified form.

The main modification concerns the initial open community development, which Rayner & Webber (1984) divided in two extreme pathways, depending on the amount of stress present. If various types of woody units are compared, this division makes good sense, as the stress regime in heartwood is very different from that of newly exposed sapwood, which again differs from that of attached dead branches (Rayner & Boddy 1988, Boddy 1992). When considering whole logs, however, the co-occurrence of several such stress related niches are likely to outrule the significance of stress between logs. At least the stress related DCA2 was in the present study only distinctly expressed among the older logs (Fig. 4; Table 8).

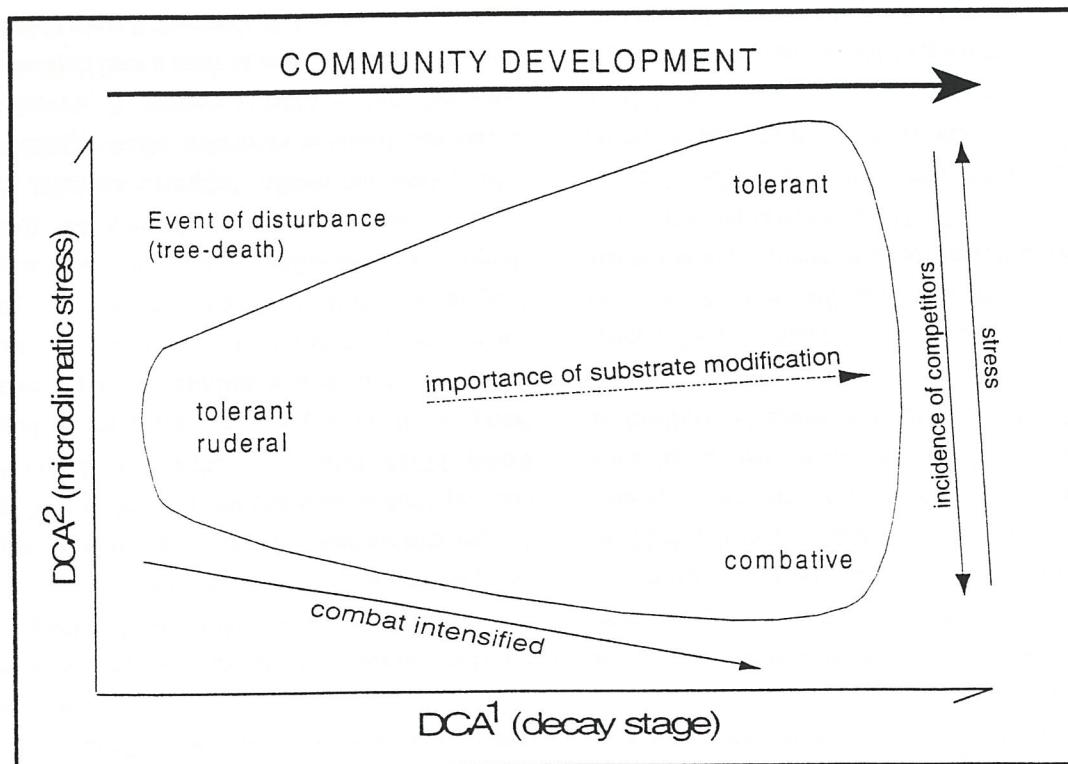


Fig. 13. A model of the fungal community development on the studied logs, based on the plot of DCA<sup>1</sup> against DCA<sup>2</sup> (Fig. 4), but laterally reversed. The model refers to the same ecological principles as the community diagram of Rayner & Webber (1984), (Fig. 12), but with substrate modification (pre-conditioning) added as an additional factor.

A second modification regards the other extreme of community development, which in the model of Rayner & Webber (1984) is implied to culminate in a predictable, declining, nutrient stress tolerant community. In contrast the present study implies that the fungal community structure becomes increasingly unpredictable as decay proceeds, as logs in the final phase of decay occur very wide spread in the ordination diagram (Fig. 8). A similar decay related increase in heterogeneity has been reported by Renvall (1995) and Høiland & Bendiksen (1996) from conifer logs, and was by Renvall (1995) explained to mirror an increase in the number of present micro-habitats in well decayed wood. Other possible explanations for the finding are listed by Høiland & Bendiksen (1996).

The two modifications result in a model somewhat opposite to the diagram by Rayner & Webber (1984) with the broadest, most differentiated part being expressed at late stages of decay (Fig. 13). The curvilinear prediction profile of log age in the regression model of DCA1 shows that the biggest amount of axis shift and hence species turnover occurs during the first half of the covered decay period. The wide spacing of logs along the younger right half of DCA1 (Fig 8) supports the finding. In combination with the two modifications this indicate that the overall course of succession shifts from being fast and predictable in early phases of decay to become increasingly slow, and unpredictable as decay proceeds.

#### *Distribution of species groups*

Along DCA1 a distinct turnover in combative ability is evident. Among the first species to obtain optima along the axis, *Nectria coccinea* and *Hypoxyylon fragiforme* have been reported as endophytic latent invaders in beech (Chapela & Boddy 1988b), and a similar strategy of 'specialised opportunism' (Rayner & Boddy 1988) seems to apply for *Inonotus nodulosus*, *Diatrype disciformis*, *Neobulgaria pura*, *Exidia glandulosa* and *E. nucleata*. All typically produce extensive fructifications within the very first years after tree death and most are selective for beech. Especially the first feature is highly characteristic of endophytic decay fungi (Rayner & Boddy 1988, Boddy 1994). The two species of *Exidia*

have a broader host selection (Hansen & Knudsen 1997), but other members of the genus, with a similar appearance, appears to be endophytic in ash and oak (Boddy & Rayner 1983, Boddy et al. 1987).

In general latent invaders have been reported to be poor competitors (Boddy et al. 1987, Chapela & Boddy 1988a,c) and their general early optima are hence expectable. Exceptions to this rule have, however, been reported (e.g. Boddy et al. 1987). Based on the present investigation *Coniophora puteana*, *Oudemansiella mucida*, *Eutypa spinosa* and *Hypoxyylon rubiginosum*, which has been reported as potential latent invaders by Chapela & Boddy 1988b, Hendry, Lonsdale & Boddy (1998) and Boddy et al. (1987) respectively, seem to have good combative or defensive abilities. All were found producing new sporocarps even on the oldest logs studied.

Judging the correlations shown on Table 11, polypores shows as a whole affinity to early phases of decay. Particularly early optima are obtained by *Bjekandera adusta*, *Datronia mollis*, *Polyporus brumalis*, *Trametes hirsuta* and *Trametes versicolor*, which together with the corticioid species *Phlebia radiata*, *P. tremellosa*, and *Stereum hirsutum* appear to form a well defined ecological group (cf. 'class 1' in Coates & Rayner (1985c)). The species often co-occur, *Trametes hirsuta*, however, has a strong preference for sun exposed wood. All are white rotters and several have been found to be very fast decayers, either experimentally (Worral, Anagnost & Zabel 1997) or under field conditions (Chapela et al. 1988, Willig & Schlegte 1995). Species of the group seem typically to replace ruderal unspecialized opportunists and latent invaders, but are themselves being replaced e.g. by cord-forming basidiomycetes a few years after establishment (Boddy & Rayner 1983, Coates & Rayner 1985c, Chapela & Boddy 1988c, Chapela et al 1988). In line with this, basidiomycetes forming mycelial cords, such as *Ramaria stricta*, *Clitocybe diatreta*, *Hypholoma fasciculare*, *Lycoperdon perlatum*, *L. pyriforme*, *Megacollybia platyphylla*, *Mutinus caninus*, *Phallus impudicus* and *Stropharia cyanea*, all have their optima in the 'older' half of the ordination diagram. Interestingly the optimum of the rhizomorphic *Armillaria gallica* (syn. *A. bulbosa* (Barla) Kile & Watling) is slightly earlier, which

supports the findings of Coates & Rayner (1985c), Chapela et al. (1988) and Dowson, Rayner & Boddy (1988) that *Armillaria* species in general are less combative than the true cord formers.

Cord formers are generally regarded as among the most combative of wood decomposers (e.g. Coates & Rayner 1985c, Boddy 1993). Under field conditions they have been found to cause a rapid decay (Chapela et al. 1988), but surprisingly this contrasts with *in vitro* experiments, in which several cord formers have been shown to have little or no ability to decay fresh wood (Dix & Cairney 1985, Tanesaka, Masuda & Kinugawa 1993, Worral et al 1997). The reason behind this paradox is probably that the cord formers depend on a preconditioning of the wood by earlier colonisers, before they can act as decayers. The understanding of DCA1 as representing a community trend solely related to the intensity of competition might thus represent a gross oversimplification. The role of substrate modification may be just as important, especially during the late phases of decay, as discussed by Renvall (1995) and by Holmer, Renvall & Stenlid (1997).

Most species of *Pluteus*, *Psathyrella* and *Mycena* have even later optima than the cord formers in the ordination diagram. Unfortunately these species, which are all agarics, have been included in very few experimental studies, and little is known about their wood degrading abilities and strategies. *Psathyrella piluliformis* (syn. *P. hydropnephila*) however was by Chapela et al. (1988) found able to replace several cord formers, and a strong combative ability may play an important role for the group in general. However, it is noteworthy that the isolation of *Pluteus* species is difficult, and depends on special nutritive media (Banerjee & Sundberg 1993, Banerjee 1994). This may on the one hand explain why the genus is not reported from studies based on isolation procedures (e.g. Chapela et al 1988), and suggests on the other hand, that members of the genus are highly dependent of a special nutritive environment, e.g. imposed by preconditioning by fungi occurring earlier in the succession.

Two ruling parameters, viz. decay rate and externally induced microclimatic stress, were recognized in the complex gradient underlying DCA2. Whereas the latter vari-

able acts fully independent of the fungal community structure, the relation between decay rate and community structure appears to be intricate. It is possible that both parameters are solely determined by the external micro-climatic regime, but the strong expression of decay rate in the model indicates, that this is not the case. The decay rate varies considerably among species of wood decaying fungi (Worral et al. 1997). In beech wood several pyrenomycetes, including *Xylaria hypoxylon* and *Nemania serpens* (syn. *Hypoxylon serpens*) have been found to be especially slow decayers, both experimentally (Boddy, Owens & Chapela 1989) and in the field (Chapela et al. 1988). Chapela et al. (1988) found that *X. hypoxylon* was always associated with very dry decay columns sealed by pseudosclerotial plates. Apparently the species is able actively to decrease the water content of occupied wood, probably as a defense against invasion of more combative fungi. The slow decay rate of the species seems to be a direct result of this strategy (Boddy et al. 1989, Boddy 1992). Similar strategies might be achieved also by other fungi forming pseudosclerotial plates, but might appear to be especially distributed among pyrenomycetes of which several have been found to be well adopted to water stress (Griffith & Boddy 1991b, Boddy 1992).

*Eutypa spinosa* and *Kretzschmaria deusta* (syn. *Ustulina deusta*) are known to cause a decay characterized by well developed pseudosclerotial plates (Pearce 1991, Hendry et al. 1998). Both species were in the present investigation recorded very frequently (Table 4) and in several logs appeared to occupy large volumes of wood, estimated from very extensive fructifications. In contrast the equally frequent *X. hypoxylon* appeared generally to be confined to small patches. Considered isolated, the decay rate is correlated negatively with the relative cover of *E. spinosa* (Table 5), but this correlation is not evident with respect to DCA2 (Table 9). The relative cover of *K. deusta* was not estimated, but the species has a rather high score on DCA2 (Fig. 9), and may have a significant effect on the decreasing decay rate expressed along the axis. This opens up for the somewhat speculative, but intriguing possibility, that the shift in community structure along DCA2 is caused not only by external micro-

climatic stress but also by microclimatic stress induced by pyrenomycetes, in particular *K. deusta*.

The most significant community trend along DCA2 is the decline in overall diversity accompanied by an increasing relative share of corticioid fungi (Table 11, Fig. 11). This trend may appear surprising as several, rare corticioid fungi are supposed to be highly dependend of a humid forest environment (Eriksson et al. 1973-1988, Larsson 1997). In general the group seems however to exhibit good adaptions to environments marked by microclimatic stress. adaptions include resistance to prolonged desiccation e.g. by clamydospore formation (Boddy 1992), resistent basidiomata regaining functionality only during humid periods (Eriksson et al. 1973-1988, Nunez & Ryvarden 1993, Nunez 1996), psychrophily, allowing growth and sporulation at low temperatures under snow or during moderately cold, humid winterperiods (Gilbertson 1973, Larsen, Jurgensen & Harwey 1981, Griffith & Boddy 1991b) and cryptic sporulation under logs and large branches where close soil contact creates a more stable, humid micro-environment (Høiland & Bendiksen 1996). The latter strategy appears to be obtained by several species with high scores on DCA2, such as *Gloiothele lactescens*, *Hypoderma praetermissum*, *Scopoloïdes rimosus*, *Styphella grilletii* and *Trechispora stevensonii*, and psychrophily appears to be a part of the strategy of *Brevicellicium olivascens*, *Hypodontia radula*, *Radulomyces confluens*, *Phlebia livida*, *Sistotrema sernanderi* and *Tulasnella violea*, all of which were found most frequently during the winther months. In both groups several members have fruitbodies adapted to repeated cycles of desiccation and rewetting, but this adaption seems to be especially prominent among corticioid fungi with low DCA1 scores, notably *Cylindrobasidium evolvens*, *Peniophora cinerea*, *P. incarnata* and *P. lycii*. These are all confined to the exposed bark and outermost sapwood of newly dead trees, an environment which offers little protection from the dessicating forces of wind and sun. Also *Schizophyllum commune*, which has appeared to be closely related to, or even congeneric with the corticioid genera *Auriculariopsis* (Nakasone 1996), is evidently well adapted to this habitat.

The diversity of slime moulds is negatively correlated with both DCA1 and DCA2 (Table 11; Fig. 11), which implies that the group has a strong partiality for well decayed, moist wood. This is not surprising since water availability has been stated to be of crucial importance for the group (Ing 1994). Interestingly the three species with highest DCA2 scores, viz. *Enteridium lycoperdon*, *Fuligo septica* and *Lycoperdon epidendrum*, all form large aethalia, in contrast to all other species included in the ordination. The significance of this finding remain unsolved.

Relatively little is known about the activity of slime moulds in decaying wood, but their role is most likely entirely indirect (Rayner & Boddy 1988, Ing 1994). Several species are able to digest fungi in artificial culture (Howard & Curie 1932) and digestion of fungal mycelia and sporocarps are also recorded in nature (Elliott & Elliott 1920, Ing 1994). Another important food resource of slime moulds is bacteria (Madelin 1984), which could explain the late peak of the group, since bacteria in general appears to occur abundantly in well decayed wood (Rayner & Boddy 1988).

Exclusion of slime moulds from the ordination resulted in a less distinct expression of DCA2 (Fig. 3), and in less powerfull correlations with environmental variables (results not shown). This indicate that slime moulds might be good indicators of the environmental characteristics of decaying wood, and a rich occurrence seems to be highly indicative of a humid wood environment. Whether a rich occurrence of slime moulds can be indicative of the biotic characteristics of wood is uncertain at present, but is likely since their biomass production in some logs appears to be considerable, estimated from the amounts of sporocarps.

#### *Sampling-methods.*

Several authors (e.g. Shigo 1967, Rayner & Todd 1979, Rayner & Boddy 1988) have seriously doubted the relevance of studies of wood decaying fungal communities, if based solely on the registration of fungal sporocarps. Most seriously it has been disputed whether the distribution of fungal sporocarps gives a realistic impression of how active mycelia are distributed within the wood. The

most thorough review on methods relevant for the study of fungal communities in decaying wood appears to be that of Boddy & Rayner (1988; chapter 3). For general purposes the authors recommend a methodology involving the cutting of wood samples to reveal the spatial distribution of mycelial individuals, based on the combined analysis of decay columns, zone lines, associated sporocarps and hyphal structures, the latter eventually inferred from cultures isolated from wood samples.

The application of such a methodology in a large scale study like the present is unfortunately unrealistic. Especially, since the amount of work needed to do this would be immense, but also for reasons of conservation. Fortunately, the shortcomings of the sole reliance of sporocarps seem to be partly outruled by the project scale, as the focus is moved from the consideration of individuals within single units, to an estimation of the relative distribution of species across a large number of units (e.g. logs).

Logs and other coarse woody debris appear in general as well spaced units on the forest floor. Each unit is gradually decomposed and is therefore a suitable habitat only for a limited period. Consequently, wood decaying fungi depends highly on spore dispersal, apart from cord forming and rhizomorphic basidiomycetes, which have an effective vegetative dispersal. In practice little is known about how mycelial activity and sporocarp production are correlated in time. However, it is a common experience that many wood decaying fungal species are very persistent in their production of sporocarps (pers. obs., Lange 1992, Renwall 1995). This indicate that mycelial activity and sporocarp production in many cases are concurring events.

With respect to macrofungi the main drawbacks by the solely sporocarp based approach, seems thus to be a modest lack of precision in the understanding of mycelial activity in time, and a potential underrepresentation of cord forming and rhizomorphic basidiomycetes. With respect to non-hymenomycetous fungi more serious drawbacks persist, though some attempts has been made to include these in solely non-destructive inventories (Hoiland & Bendiksen 1996, Penttilä & Kotiranta 1996). Microfungi are often richly represented in decaying

wood, apparently mostly in the first phases of decay, e.g. as endophytes (Shigo 1967, Chapela & Boddy 1988a, Griffith & Boddy 1990), in strongly decayed wood (Chapela et al. 1988, Rayner & Boddy 1988) or associated with insect activity (Shigo 1967, Swift & Boddy 1984). However, it remains uncertain to which degree they take active part in the primary decomposition of wood under normal conditions, and to which degree they may influence the overall community structure. It is also unclear to which degree fungi reported as microfungi in various studies, in fact are linked with teleomorph stages, producing conspicuous sporocarps under certain conditions, e.g. during wintertime, where the activity of field mycologist are generally limited. Future studies dealing with these aspects are highly welcome.

I wish to thank Thomas Læssøe for highly useful comments and improvements to the manuscript and Morten Christensen for fruitful discussions and nice hours in the field. Rasmus Ejrnæs is thanked for introducing me to the world of ordination. Henrik F. Gøtzsche, Nils Haltenberg, Seppo Huhtinen, Henning Knudsen, Ewald Langer, Thomas Læssøe, Ain Raitvää, Peter Roberts, Leif Örstadius and last but not least Jan Vesterholt are thanked for help with the determination of various critical specimens. Finally I want to thank the Foundation of Sorø Akademi for giving me the permission to work in Suserup Skov.

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*Ischnoderma resinosum* er for tiden kun kendt fra en dansk lokalitet – Suserup Skov. Her er arten til gengæld ganske udbredt.

At present *Ischnoderma resinosum* is only known from one Danish locality – Suserup Skov. Here the species occurs quite abundantly.

Photo: Jacob Heilmann-Clausen, 13/11 1998 (coll. JHC 98-117)

Appendix 1. Total artsliste. Arterne er inddelt i praktiske grupper af vekslende taxonomisk værdi. Med sikkerhed kunstige grupper er anført med anførelsestegn. I kolonnen benævnt "det," er det angivet, hvis andre end jeg har selv har været indblandet i bestemmelsesarbejdet. A1: Ain Raitviir (Tartu), EL: Ewald Langer (Tübingen), NH: Nils Hallenberg (Göteborg), HFG: Henrik F. Götzsche (København), HK: Henning Kudsen (København), JV: Jan Vesterholt (Vejle), LÖ: Leif Örstadius (Kristiansstad), SH: Seppo Huhtinen (Turku), PR: Peter Roberts (Kew), TL: Thomas Læssøe (København)

		Det.	Antal registreringer fordelt på indsamlingsdatoer												Frekvens af arter fordelt på stammealders-klasser (år)						
			24/2 1995	7-8/5 1995	29/5 1994	10/7 1994	23-24/8 1994	25/9 1994	1/10 1995	4/10 1995	15/10 1994	1/11 1995	14+17/12 1995	14+17/12 1995	1-4	5-9 1995	10-13 1995	14-16 1995	17-22 1995	23-27 1995	28-34 sum
Diatrypales																					
Diatrype disciformis	(Hoffm.: Fr.) Fr.	TL	2	3											0	0	0	0	0	5	
Diatrype flavovirens	(Pers.: Fr.) Fr.	TL	1	1											0	0	0	0	0	1	
Diatrype stigma	(Hoffm.: Fr.) Fr.	TL	5	4											0	0	0	0	0	1	
Eutypa spinosa	(Pers.: Fr.) Tul. & C.Tul.														10	10	8	8	7	62	
Eutypella quaternata	(Pers.: Fr.) Rappaz														0	0	0	1	1	2	
Hypocreales																					
Hypocre gelatinosa	(Tode: Fr.) Fr.	TL	1	1											1	0	0	1	1	4	
Hypomyces aurantius	(Pers.: Fr.) Tul. & C.Tul.	TL	2	2											2	0	1	3	0	0	
Nectria coccinea	(Pers.: Fr.) Fr.	TL	1	2											4	0	0	0	0	4	
Nectria episphaeria	(Tode: Fr.) Fr.	TL	1	1											0	0	0	0	1	1	
Nectria peziza	(Tode: Fr.) Fr.	TL	2	2											0	0	0	0	0	2	
Protocrea cfr. farinosa	(Berk. & Br.) Petch														0	0	1	0	0	1	
Leotiales																					
Arachnopeziza aurata	Fuckel	SH	1	1											0	0	0	0	1	3	
Arachnopeziza variegilosa	(Galán & Raitvii) Huhtinen														1	0	0	0	1	1	
Ascocyste cylindrium	(Tul.) Korf	AR	3	1											8	1	3	1	1	8	
Bisporella citrina	(Batsch: Fr.) Korf & S.Carp. s.l.	AR	1	1											10	2	4	3	0	10	
Cystopeltizella conorum	(Rehm) Srvcsek														0	0	0	0	0	1	
Dasyoscyphella nivea	(Hedw.: Fr.) Raitv.														0	0	0	0	2	3	
Haloscyphula fuckeli	Nannf.														2	0	0	0	0	5	
Lachnum brevipilosum	Baral														3	1	0	1	1	5	
Lachnum impudicum	Baral														15	2	3	8	3	22	
Lachnum virgineum	(Batsch: Fr.) P.Karst.														0	0	2	0	0	4	
Mollisia cinerea	(Batsch) P.Karst.														1	6	10	7	10	49	
Mollisia ligni	(Desm.) P.Karst.														1	0	1	0	0	6	
Neobulgaria pura	(Fr.) Petrak														4	5	1	0	0	2	
Ombrophila sp.															6	1	0	0	0	19	
Orbilia cf. alinea	Velten.														1	1	2	2	5	6	
Orbilia delicatula	(P.Karst.) P.Karst.														7	0	1	0	0	2	
Orbilia epipora	(Nyl.) P.Karst.														2	1	1	0	0	12	
Polydesmia priuosa	(Jerdon in Berk. & Br.) Boud.														1	1	0	1	0	0	
Tapesia lividofusca	(Fr.: Fr.) Rehm														11	1	1	1	4	1	



	24/2 1995	7-8/5 1994	29/5 1994	10/7 1994	23-24/8 1994	25/9 1995	1/10 1995	4/10 1995	15/10 1994	1/11 1995	14+17/12 1995	1-4 1995	5-9 1995	10-13 1995	14-16 1995	17-22 1995	23-27 1995	28-34 1995	sum
Armillaria mellea (Vahl.: Fr.) P.Kumm.																			1
Bolbitius reticulatus (Pers.: Fr.) Rick.																			7
Clitocybe diaireta (Fr.: Fr.) P.Kumm.																			21
Clitocybe nebularis (Batsch: Fr.) P.Kumm.																			9
Clitocybe phyllophila (Pers.: Fr.) P.Kumm.																			3
Clitopilus hobsonii (Berk.) P.D.Orton	1																		10
Collybia butyracea (Bull.: Fr.) P.Kumm.																			8
Collybia fusipes (Bull.: Fr.) Quél.																			1
Collybia personata (Bolt.: Fr.) P.Kumm.	1																		1
Collybia tuberosa (Bull.: Fr.) P.Kumm.	1																		2
Conocybe brunnea (J.E.Lange & K.Hner) ex Watling																			3
Conocybe sordida (Kühner ex) Kühner & Watling																			3
Conocybe tenera (Schaeff.: Fr.) Fayod																			3
Coprinus domesticus (Bolt.: Fr.) Gray																			1
Coprinus echinosporus Buller																			1
Coprinus lagopus (Fr.: Fr.) Fr.																			36
Coprinus micaceus (Bull.: Fr.) Fr.																			1
Coprinus xanthothrix Romagn.																			1
Cystolepiota adulterina (F.H.Möller) Bon																			1
Cystolepiota heteri (Boud.) Singer																			1
Entoloma dichroum (Pers.: Fr.) P.Kumm.																			1
Flammulaster muricatus (Fr.: Fr.) Watling																			1
Galerina nana (Petr.) Kühner																			3
Galerina unicolor (Fr.) Singer																			3
Gymnopilus junonius (Fr.: Fr.) P.D.Orton																			5
Hemimyces cucullata (Pers.: Fr.) Singer																			5
Henningssomyces candidus (Pers.: Fr.) O.K.																			5
Hydropus subalpinus (Höhn.) Singer																			5
Hypoloma fasciculare (Huds.: Fr.) P.Kumm.																			35
Inocybe petiginosa (Fr.: Fr.) Gillet																			1
Kuehneromyces mutabilis (Schaeff.: Fr.) Sing. & Smith																			15
Laccaria laccata (Scop.: Fr.) Berk. & Br.																			1
Lepiota aspera (Pers.: Fr.) Quél.																			6
Lepiota boertmannii Knudsen																			14
Lepiota cristata (Bolt.: Fr.) P.Kumm.																			4
Lepiota fulvella Rea																			1
Lepiota jacobii Vellinga & Knudsen																			2
Lepiota ochraceofulva P.D.Orton																			1
Lepiota flaccida (Sow.: Fr.) Pat.																			3
Lepista nuda (Bull.: Fr.) Cooke																			1
Macrolepiota rhacodes (Vitt.) Singer																			12
Marasmius alliaceus (Jacq.: Fr.) Fr.	4	1	21	1	3	27	6	14	1	8	6	7	6	6	8	4	12	40	

HK

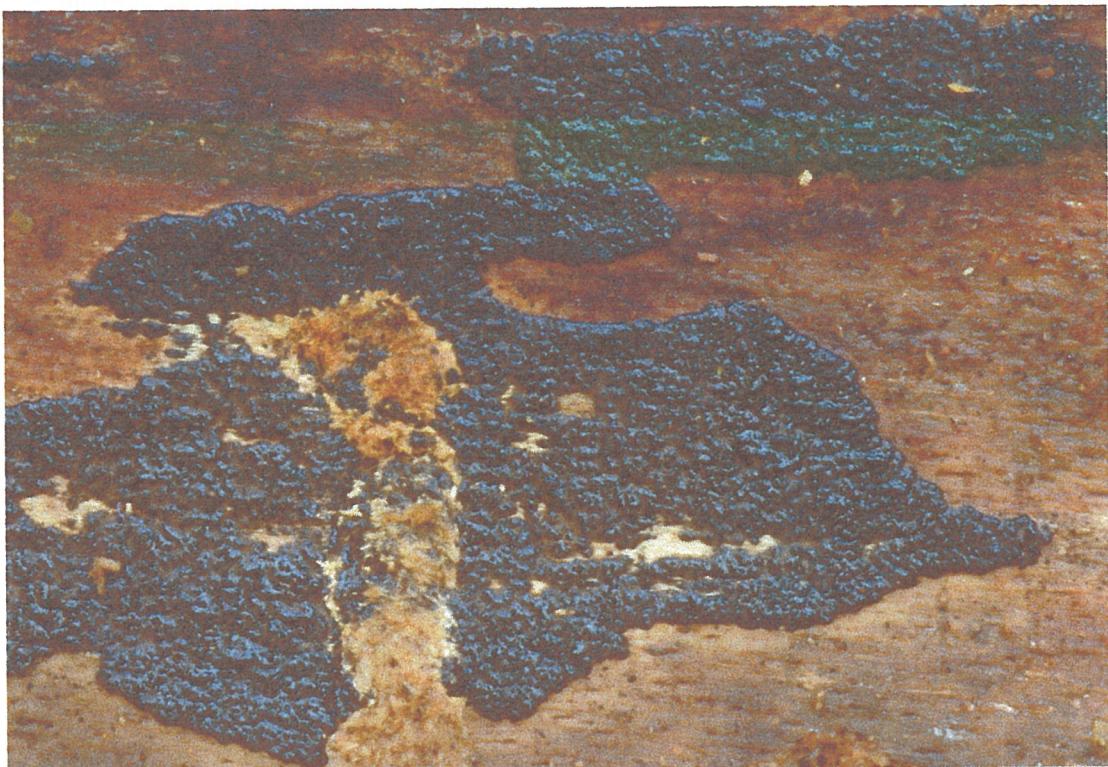
	24/2 1995	7-8/5 1994	29/5 1994	10/7 1994	23-24/8 1994	25/9 1995	1/10 1995	4/10 1994	15/10 1995	1/11 1995	14+17/12 1995	1-4 1995	5-9 1995	10-13 1995	14-16 1995	17-22 1995	23-27 1995	28-34 1995	sum
<i>Marasmius cohaerens</i> (Alb. & Schw.: Fr.) Cooke & Quél.																			1
<i>Marasmius rotula</i> (Scop.: Fr.) Fr.																			27
<i>Marasmius torquescens</i> Quél.																			9
<i>Marasmius wynnei</i> Berk. & Br.																			1
<i>Megacollybia platyphylla</i> (Pers.: Fr.) Kotl. & Pouz.																			1
<i>Melanophyllum aimatospermum</i> (Bull.: Fr.) Kreisel																			22
<i>Melanotus horizontalis</i> (Bull.) P.D.Orton																			2
<i>Mérismodes anomalus</i> (Pers.: Fr.) Singer																			2
<i>Micromphale brassicola</i> (Romagn.) P.D.Orton																			1
<i>Mycena amicta</i> (Fr.) Quél.																			3
<i>Mycena crocata</i> (Schrad.: Fr.) P.Kumm.																			35
<i>Mycena diosma</i> Kriegsteiner & Schwöbel																			6
<i>Mycena erubescens</i> Höhn.																			2
<i>Mycena galericulata</i> (Scop.: Fr.) Quél.																			2
<i>Mycena galopus</i> (Pers.: Fr.) P.Kumm.																			28
<i>Mycena haematopus</i> (Pers.: Fr.) P.Kumm.																			4
<i>Mycena hiemalis</i> (Osb.: Fr.) Quél.																			50
<i>Mycena pelianthina</i> (Fr.) Quél.																			3
<i>Mycena polygramma</i> (Bull.: Fr.) Gray																			2
<i>Mycena pura</i> (Pers.: Fr.) P.Kumm.																			17
<i>Mycena renati</i> Quél.																			11
<i>Mycena rosea</i> Gramberg																			4
<i>Mycena speirea</i> (Fr.: Fr.) Gillet																			4
<i>Mycena tintinabulum</i> (Fr.) Quél.																			4
<i>Omphalina epichysium</i> (Pers.: Fr.) Quél.																			8
<i>Oudemansiella mucida</i> (Schrad.: Fr.) Höhn.																			14
<i>Panellus serotinus</i> (Pers.: Fr.) Kühn.																			6
<i>Pholiota aurivillus</i> (Fr.) P.Kumm.																			2
<i>Pholiota squarrosa</i> (Weig.) Fr.: P.Kumm.																			2
<i>Pleurotus dryinus</i> (Pers.: Fr.) P.Kumm.																			10
<i>Pleurotus ostreatus</i> (Jacq.: Fr.) P.Kumm.																			10
<i>Pluteus cervinus</i> (Batsch) Singer																			8
<i>Pluteus chrysophaeus</i> (Schaeff.) Quél.																			37
<i>Pluteus ephebeus</i> (Fr.: Fr.) Gillet																			11
<i>Pluteus godeyii</i> Gill. ss. Lange																			2
<i>Pluteus inquinans</i> Romagn.																			22
<i>Pluteus nanus</i> (Pers.: Fr.) P.Kumm.																			5
<i>Pluteus phlebophorus</i> (Ditmar.: Fr.) P.Kumm.																			1
<i>Pluteus podospileus</i> Sacc. & Cub.																			0
<i>Pluteus salicinus</i> (Pers.: Fr.) P.Kumm.																			1
<i>Pluteus semibulbosus</i> (Lasch) Gill.																			3
<i>Pluteus thomsonii</i> (Berk. & Br.) Dennis																			0

	24/2 1995	7-8/5 1994	29/5 1994	10/7 1994	23-24/8 1994	25/9 1995	4/10 1995	15/10 1995	1/11 1995	14+17/12 1995	1-4 1995	5-9 1995	10-13 1995	14-16 1995	17-22 1995	23-27 1995	28-34 sum
Pluteus umbrinosus (Fr.) P.Kumm.																	5
Psathyrella candolleana (Fr.: Fr.) Maire																	1
Psathyrella corynoides P.D.Orton	LÖ																0
Psathyrella conopilus (Fr.: Fr.) A.Pears.																	0
Psathyrella fusca (Schum.) A.H.Smith																	1
Psathyrella lacrymabunda (Bull.: Fr.) Moser																	0
Psathyrella obtusata (Pers.: Fr.) A.H.Smith	LÖ	4	10	2	1	1	1	1	1	0	0	0	0	0	0	0	1
Psathyrella piluliformis (Bull.: Fr.) P.D.Orton																	0
Psathyrella populina (Britz.) Kits.																	0
Psathyrella pygmaea (Bull.: Fr.) Singer																	0
Psathyrella rostellata Örstadius	LÖ																1
Psathyrella spadiceogrisea (Schaeff.) Maire																	4
Psathyrella tephrophiyla (Romagn.) Bon																	1
Pseudodictyotibe cyathiformis (Bull.: Fr.) Singer																	8
Ramnicola centunculus (Fr.: Fr.) Wattl.																	1
Resupinatus trichotis (Pers.) Singer																	1
Ripartites tricholoma (Alb. & Schw.: Fr.) P.Karst.																	1
Stropharia cyanea (Bull.) Tuomikoski																	3
Tricholoma lascivum (Fr.) Gillet																	1
Tubaria furfuracea (Pers.: Fr.) Gillet																	2
Volvariella hypophysis (Fr.) Shaffer																	1
Xerula radicata (Rehm: Fr.) Dörfelt																	10
<b>Boletales</b>																	
Coniolephora arida (Fr.) Karst.																	0
Coniolephora puteana (Schum.: Fr.) P.Karst.																	0
“Corticiales”																	17
Athelia epiphylla Pers.																	1
Athelopsis glauicina (Boud. & Galz.) Parm.																	8
Botryobasidium botryosum (Bres.) J.Erikss.																	1
Botryobasidium conspersum (J.Erikss.)																	1
Botryobasidium laeve (J.Erikss.) Parm.																	29
Botryobasidium pruinatum (Bres.) J.Erikss.																	6
Botryobasidium subcoronatum (Höhn. & Litsch.) Donk																	5
Brevicelllicium olivascens (Bres.) Larss. & Hjortst.																	9
Bysomeserulus corium (Fr.) Parm.																	3
Ceriporia excelsa (Lund.) Parm.																	8
Ceriporia reticulata (Hoffm.: Fr.) Dom.																	38
Cristinia gallica (Piliat) Jülich																	2
Cristinia helvetica (Pers.) Parm.																	0
Cylindrobasidium evolvens (Fr.: Fr.) Jülich																	2
Gloeocystidiellum luridum (Bres.) Boid.																	7
Gloeohypochnicium analogum (Bourd. & Galz.) Hjortst.																	3

		7-8/5 1995	29/5 1994	10/7 1994	23-24/8 1994	25/9 1995	1/10 1995	4/10 1995	15/10 1994	1/11 1995	14+17/12 1995	1-4 1995	5-9 1995	10-13 1995	14-16 1995	17-22 1995	23-27 1995	28-34 sum
Gliothelie lactescens (Berk.) Hjortst.																		
Hypoderma aff. roseocremeum (Bres.) Donk	NH	1	1	1	1	3	2	0	0	0	3	2	3	2	0	1	6	19
Hypoderma argillaceum (Bres.) Donk	JV	2																
Hypoderma medioburiense (Buit) Donk																		
Hypoderma pallidum (Bres.) Donk																		
Hypoderma praetermissum (P.Karst.) J.Erikss. & Strid	JV	1	1	1	2	1	10	0	0	0	0	0	0	0	0	0	0	1
Hypoderma puberulum (Fr.) Wallr.																		
Hypoderma roseocremeum (Bres.) Donk																		
Hypoderma setigerum (Fr.) Donk																		
Hypodontia anguta (Fr.) J.Erikss.																		
Hypodontia aspera (Fr.) J.Erikss.																		
Hypodontia paradoxa (Fr.) E.Langer & Vesterh.																		
Hypodontia radula (Pers.: Fr.) E.Langer & Vesterh.																		
Hypodontia sambuci (Pers.) J.Erikss.	NH	10	3	2	1	1	1	1	0	0	0	0	0	0	0	0	0	6
Hypodontia subalutacea (P.Karst.) J.Erikss.	JV	1																16
Hypochnicium eichleri (Bres.) J.Erikss. & Ryvarden																		
Hypochnicium polonense (Bres.) Strid																		
Hypochnicium punctulatum (Cooke) J.Erikss.																		
Lindneria cir. flavula Parm.																		
Mycacia uda (Fr.) Donk																		
Peniophora cinerea (Pers.: Fr.) Cooke																		
Peniophora incarnata (Pers.: Fr.) P.Karst.																		
Peniophora lycii (Pers.) Höhn. & Litsch.																		
Peniophora nuda (Fr.) Bres.																		
Peniophora querina (Pers.: Fr.) Cooke																		
Phaneirochaete sordida (P.Karst.) J.Erikss. & Ryvarden																		
Phaneirochaete tuberculata (P.Karst.) Parm.																		
Phaneirochaete velutina (DC.: Fr.) P.Karst.																		
Phlebia livida (Pers.: Fr.) Bres.																		
Phlebia radiata Fr.: Fr.																		
Phlebia rufa (Pers.: Fr.) M.P.Christ.																		
Phlebia tremilloides (Schrad.: Fr.) Burds. & Nakas.																		
Phlebiella allantospora (Oberw.) Larss. & Hjortst.																		
Radulomyces confluens (Fr.: Fr.) M.P.Christ.																		
Schizophyllum commune Fr.: Fr.																		
Scopuloides rimosa (Cooke) Jülich																		
Sistotrema biggsiae Hallenb.																		
Sistotrema brinkmannii (Bres.) J.Erikss.	NH	16	1	1	3	5	4	1	0	1	0	0	0	0	0	0	0	4
Sistotrema coroniferum (Höhn. & Litsch.) Donk	JV, NH	3	1	2	3	2	0	1	0	0	0	0	0	0	0	0	0	6
Sistotrema octosporum (Schroet. ex Höhn. & Litsch.) Hallenb.	NH	11	2	1	3	7	4	2	0	0	0	0	0	0	0	0	0	20
Sistotrema sernanderi (Litsch.) Donk	NH	1				1	1	0	0	0	0	0	0	0	0	0	0	5
		2				1	3	0	0	0	0	0	0	0	0	0	0	2
																		5

	24/2 1995	7-8/5 1994	29/5 1994	10/7 1994	23-24/8 1994	25/9 1994	11/10 1995	4/10 1995	15/10 1995	17/11 1995	14+17/12 1995	1-4 1995	5-9 1995	10-13 1995	14-16 1995	17-22 1995	23-27 1995	28-34 1995	sum
<i>Sistotremastrum niveocremneum</i> (Höhn. & Litsch.) J Erikss.													0	1	0	0	0	2	
<i>Steccherinum ochraceum</i> (Pers.: Fr.) Gray													1	2	0	0	0	16	
<i>Stereum hirsutum</i> (Willd.: Fr.) Fr.													1	0	0	0	0	21	
<i>Subulicystidium longisporum</i> (Pat.) Parm.													3	1	2	0	0	7	
<i>Thanatephorus fusicporus</i> (J.Schroet.) Hauerslev & P.Roberts													1	1	2	0	1	12	
<i>Trechispora cohaerens</i> (Schw.) Jlich & Stalpers													0	0	2	0	1	3	
<i>Trechispora farinacea</i> (Pers.: Fr.) Lib.													2	0	0	0	2	9	
<i>Trechispora hymenocystis</i> (Berk. & Br.) K.H.Larss.													1	0	0	0	0	4	
<i>Trechispora microspora</i> (P.Karst.) Lib.													1	0	0	0	0	1	
<i>Trechispora praefocata</i> (Boud. & Galz.) Lib.													0	1	0	0	0	0	
<i>Trechispora stevensoni</i> (Berk. & Br.) Larss.													1	0	0	0	0	7	
<i>Xenasma pulverulentum</i> (Litsch.) Donk													0	0	0	0	0	1	
<b>Ganodermatales</b>													1	0	2	0	1	7	
<i>Ganoderma lipiensis</i> (Batsch) Atk.													0	0	0	0	0	1	
<i>Ganoderma pfeifferi</i> Bres.													0	0	0	0	0	22	
<b>Gomphales</b>													0	0	0	0	0	1	
<i>Ramaria stricta</i> (Pers.: Fr.) Quél.													0	0	0	0	0	15	
<b>Hericiales</b>													2	2	3	2	1	4	
<i>Hericium coralloides</i> (Scop.: Fr.) Pers.													0	0	2	0	0	2	
<b>Hymenochaetales</b>													0	0	0	0	0	0	
<i>Fuscoporia ferrea</i> (Pers.) G.Cunn.													1	0	0	0	0	5	
<i>Inonotus nodulosus</i> (Fr.) P.Karst.													3	4	2	0	0	6	
<b>"Polyporales"</b>													0	0	0	0	0	0	
<i>Antrodia hoehnelii</i> (Bres.) Niemelä													0	2	0	0	0	2	
<i>Antrodia semisupina</i> (Berk. & Curt.) Ryvarden & Johans.													1	0	0	0	0	2	
<i>Bjerkandera adusta</i> (Willd.: Fr.) P.Karst.													11	6	8	2	0	17	
<i>Bjerkandera fumosa</i> (Pers.: Fr.) P.Karst.													1	0	0	0	0	0	
<i>Ceriporiopsis gilvescens</i> (Bres.) Dom.													3	2	0	2	0	9	
<i>Datronia mollis</i> (Sommerf.: Fr.) Donk													0	1	0	0	0	6	
<i>Fomes fomentarius</i> (L.: Fr.) Fr.													6	1	1	0	0	38	
<i>Fomitopsis pinicola</i> (Swartz.: Fr.) P.Karst.													39	1	4	2	3	0	
<i>Ischnoderrma resinosum</i> (Schrad.: Fr.) P.Karst.													3	1	0	4	4	0	
<i>Meipilus giganteus</i> (Pers.: Fr.) P.Karst.													6	1	0	4	4	14	
<i>Physiporus sanguinolentus</i> (Alb. & Schw.: Fr.) Pilát													1	1	3	5	3	18	
<i>Physiporus vitreus</i> (Pers.: Fr.) P.Karst.													1	1	4	4	2	16	
<i>Polyporus badius</i> (Pers.) Schw.													1	0	0	0	0	0	
<i>Polyporus brumalis</i> (Pers.) Fr.: Fr.													1	2	1	0	0	3	
<i>Polyporus ciliatus</i> Fr.: Fr.													1	3	1	0	0	3	
<i>Polyporus squamosus</i> (Huds.: Fr.) Fr.													1	2	0	0	0	3	
<i>Polyporus tubercularis</i> (Pers.: Fr.) Fr.													1	1	0	0	0	3	
<i>Polyporus varius</i> (Pers.) Fr.: Fr.													3	3	6	1	0	0	

	24/2 1995	7-8/5 1995	29/5 1994	10/7 1994	23-24/8 1994	25/9 1995	1/10 1995	4/10 1995	15/10 1994	19/10 1995	14+17/12 1995	1-4 1995	5-9 1995	10-13 1995	14-16 1995	17-22 1995	23-27 1995	28-34 1995	sum
Skeletocutis nivea (Jungh.) Keller																			2
Trametes gibbosa (Pers.: Fr.) Fr.																			2
Trametes hirsuta (Wulfen: Fr.) Pilát.																			7
Trametes versicolor (L.: Fr.) Quél.																			20
Tyromyces chioneus (Fr.: Fr.) P.Karst.																			1
Tyromyces wynnei (Berk. & Br.) Donk																			1
<b>Thelephorales</b>																			4
Tomentella ferruginea (Pers.) Pat.																			1
Tomentella radiospora (P.Karst.) Rick																			1
Tomentella subtilicina (Ellis & Holway) Wakef.																			5
<b>Russulales</b>																			1
Lactarius subdulcis (Bull.: Fr.) Gray																			4
<b>MYXOMYCOTA</b>																			
Arcyria affinis Rostr.																			3
Arcyria cinerea (Bull.) Pers.																			4
Arcyria denudata (L.) Wetst.																			2
Arcyria incarnata (Pers.) Pers.																			24
Ceratiomyxa fruticulosa (Müll.) Macbr.																			4
Comatricha alta Preuss																			1
Comatricha nigra (Pers.) Schroet.																			2
Cribaria argillacea (Pers.) Pers.																			5
Enteridium lycopédon (Bull.) Farr.																			18
Fuligo septica (L.) Wiggers																			19
Lamproderma arcuoides (Sommerf.) Rost.																			3
Lycogala epidendrum (L.) Fr.																			35
Metatrichia floriformis (Schw.) Nann.-Brem.																			1
Metatrichia vesparium (Batsch) Nann.-Brem.																			2
Physarum cinereum (Batsch) Pers.																			1
Stemonitis axifera (Bull.) Macbr.																			7
Stemonitis fusca Roth																			6
Stemonitopsis hyperocea (Meylan) Nann.-Brem.																			1
Stemonitopsis typhina (Wiggers) Nann.-Brem.																			6
Symphytocarpus amaurochaetoides Nann.-Brem.																			2
Trichia contorta (Ditm.) Fr.																			1
Trichia persimilis P.Karst.																			1
Trichia scabra Roth.																			10
Trichia varia (Pers.) Pers.																			29
Tubifera ferruginea (Batsch) J.F.Gmel.																			1



*Nemania chestersii* er ikke tidligere angivet fra Danmark, men er en karakterart for flere af de allerbedste danske dødt-veds lokaliteter. Se iøvrigt under taksonomiske og floristiske noter.

*Nemania chestersii* is not officially reported from Denmark, but is an abundant species on several of the best Danish "dead wood localities".

Photo: Jacob Heilmann-Clausen, 13/11 1998 (JHC98-112)

## APPENDIX 2.

Taksonomiske og floristiske noter.

### Kærnesvampe

#### *Camarops tubulina*

Arten er ikke tidligere angivet fra bøg i Danmark, men er efterhånden fundet på dette substrat på en del sjællandske lokaliteter, samt i Stenderup Midtskov i Østjylland. Arten findes tillige på ædelgran og er fundet et par gange på denne vært i Danmark.

#### *Nemania atropurpurea*

Arten er ikke tidligere angivet fra Danmark, men forekommer dog ganske udbredt og på flere forskellige værter (Granmo & Læssøe in prep.). Den typiske habitat er blottet (uægte) kærneved af bøg, i modsætning til slægtens øvrige arter, der normalt findes på splintved.

#### *Nemania chestersii*

Arten er ikke tidligere angivet fra Danmark og er tilsyneladende sjælden og tilknyttet naturskove med større dødtvedsmængder. Den normale vært er bøg (Granmo & Læssøe in prep.), men arten kan sjældent findes også på andre substrater.

#### *Protocrea cfr. farinosa*

En indsamling i materialet er indtil videre henført under dette navn (se f.eks. Dennis 1968). Artsgruppen omkring dette taxon er dog langt fra udredt til bunds, og det er usikkert om navnet i dette tilfælde er brugt rigtigt (pers. komm. Thomas Læssøe).

### Skivesvampe

#### *Arachnopeziza variopilosa*

Arten er ikke tidligere angivet fra Danmark. Den blev først beskrevet af Gålan & Raitviir (1986), som *Protoungiularia variepilosa*. Huhtinen (1987) kombinerede arten ind i *Arachnopeziza* og citerer materiale af arten fra 2 lokaliteter i henholdsvis Spanien (holotype) og Tjekkiet. Gålan & Raitviir (1986), nævner ingen indsamlinger udover typen. Kendt fra én indsamling i materialet, som er bestemt af Seppo Huhtinen (Turku).

#### *Bisporella citrina* s.l.

Tre indsamlinger i materialet har afveget fra *B. citrina* ss.auct. ved at have markant kortere sække end normalt (< 100 um mod normalt 100-125 um (Breitenbach & Kränzlin 1981) / op til 135 um (Dennis 1968)).

Ain Raitviir (Tartu), der har studeret indsamlingen, har ikke umidelbart haft et alternativt navn at foreslå. Da de fleste bestemmelser af *B. citrina* er foregået i felten, uden indsamling af kontrolmateriale, har det desværre ikke været muligt at holde *B. citrina* ss.str. adskilt fra de afvigende kollektioner.

*Cystopezizella conorum*

Arten er beskrevet fra fyrekogler, og findes sjældent på andre substrater. Ain Raitviir der har bestemt materialets enlige kollektion, kender til to andre indsamlinger af arten fra råddent løvtræ; en fra Spanien og en fra Estland. I følge Jan Vesterholt (Vejle) er arten ikke ualmindelig på fyrrenåle i Danmark.

*Orbilia cfr. alnea.*

Denne og den følgende art er begge bestemt ved hjælp af Jan Vesterholts upublicerede skivesvampenøgler. Jeg har ikke fundet Vesterholts originalkilde til *O. alnea*. Svrcek (1954) synonymiserer arten med *O. xanthostigma* (Fr.) Fr., som Vesterholt udnøgler som en selvstændig art, adskillende sig bla. ved lidt bredere sporer. *O. acuum* Velen. er et andet muligt navn for arten. Denne art er dog beskrevet fra fyrrenåle, men er med nogen tøven angivet fra dødt ved hos Kirk & Spooner (1984). Slægten trænger kraftigt til en revision.

*Orbilia epipora*. Bestemmelsen af denne art er rimelig sikker. Arten er kendetegnet ved at ascii er delvis sammenhængende (danner et epithecium) og ved at sporerne er forholdsvis små og cylindriske (Svrcek 1954, Hansen & Knudsen 1999). *O. oreadum* Velen. som Vesterholt udnøgler selvstændigt, synes at stå meget nær. Arten adskiller sig navnligt fra *O. epipora* ved at sporene er svagt krumme. *O. epipora* er ikke tidligere angivet fra Danmark.

*Ombrophila* sp.

Kendt fra én indsamling i materialet, men senere samlet på bøgestammer i Stenderup Midtskov (Østjylland) samt i Lellinge Skovhusvænge (Sydsjælland). Desuden samlet af Thomas Læssøe (København) på en aske-stamme i Suserup Skov. Jeg har sendt materiale til Ain Raitviir, som har meddelt at han kender arten fra adskillige norske lokaliteter, men at han ikke har kunnet finde et navn for den. Formodentlig vil arten blive nybeskrevet af Ain Raitviir og Seppo Huhtinen indenfor en overskuelig fremtid.

*Peziza micropus*

Navnet er brugt forholdsvis ukritisk om en stor gulbrun, +/- stilket bægersvamp, der forekommer almindelig på bøgeved ikke bare i Suserup Skov, men også på andre lokaliteter med indslag af dødt ved. Arten tilhører en dårlig udredt gruppe indenfor slægten. Gruppen er pt. under udredning i forbindelse med et PhD projekt ved Karen Hansen, Københavns Universitet, og det er endnu uvist om *P. micropus* er det rette navn for arten.

Barksvampe

*Athelia epiphylla* s.l.

En af de mest almindelige barksvampe i Suserup Skov. Ifølge Eriksson & Ryvarden (1973) udgør *Athelia epiphylla* nærmere et arts kompleks end en enkel art, og den er da også hos Jülich (1984) splittet op i adskillige arter. Jeg har her ukritisk fulgt det brede arts-koncept hos Eriksson & Ryvarden (1973). Det er sandsynligt at enkelte angivelser i materialet dækker over andre arter i slægten, navnlig *A. neuhofii* (Bres.) Donk., der makroskopisk minder meget om *A. epiphylla*. Jeg har dog undersøgt talrige indsamlinger af *Athelia* fra Suserup Skov, uden at støde på andre arter end *A. epiphylla* s.l.

#### *Botryobasidium conspersum*

Det kønnede stadie af denne art kan ikke tilfredstillende adskilles fra den nærtstående *B. aureum* Parm. Begge arter danner gullige til brunlige pudeformede anamorfer tilhørende formslægten *Alysidium* Kunz., men med individuelle forskelle der tillader en uproblematisk artsadskillelse (Eriksson & Ryvarden 1973, Vesterholz i Hansen & Knudsen 1997). Kun *B. conspersum* er med sikkerhed kendt fra Suserup Skov, men jeg finder det sandsynligt at også *B. aureum* forekommer, på trods af arten er kendt fra uhyre få nordiske indsamlinger (Eriksson & Ryvarden 1973). Jeg er dog først blevet opmærksom på at det forholder sig således, flere år efter at indsamlingsarbejdet blev afsluttet, og har således ikke taget højde for problemet, hvilket jeg beklager.

#### *Hyphoderma aff. roseocremeum*

To indsamlinger henført under denne betegnelse afviger fra typisk materiale af arten ved at hyferne i context er amyloide og ved at basidierne er extremt lange (60-80 um, mod normalt 25-30(-35) um (Eriksson & Ryvarden 1975)). Desuden forekommer der spredte, svagt grenede hyphidier og sporerne er noget bredere end normalt (4-5 um mod normalt 3-4 um (Eriksson & Ryvarden 1975)).

Umiddelbart bedømte jeg indsamlingerne til at repræsentere en art indenfor slægten *Corticium*, der bl.a. er karakteriseret ved meget lange basidier og forekomsten af dendrohyphidier. Jeg sendte derefter en indsamling af arten til Nils Hallenberg (Göteborg), der bestemte den til *H. roseocremeum*. Dog havde han selv sin tvivl om bestemmelsen og havde selv været omkring *Corticium* s.l. (Der hos Hansen & Knudsen 1997 henføres til en helt anden orden end *Hyphoderma*). Det er nu planen at få arten i kultur, for at fastslå artens rette idenditet. Ifølge Nils Hallenberg (pers. komm.) er der sandsynligvis tale om et ubeskrevet taxon.

#### *Hyphodontia sambucina* s.l.

Sporerne og cystiderne angives hos denne art til at måle henholdsvis 4.5-6(-7) x (3-) 3.5-4 (-4.5) um og 20-50 x 3.5-5 um hos Eriksson & Ryvarden (1976), mens Langer (1994) giver målene 4.5-7 x 3.5-4.5 um (sporer) 19-33 x 3.5-5 µm (cystider). Til sammenligning målte jeg sporerne på materialets enlige indsamling til 6-8 x 4-5 µm, cystiderne til 40-60 x 4-7 µm; dvs. klart I overkanten. Nils Hallenberg (pers. komm.), som har set indsamlingen, og konfirmeret bestemmelsen, betegner *H. sambucina* som et artskompleks, der kun kan afklares ved hjælp af omfattende parrings-studier.

#### *Lindtneria* cfr. *flava*

Slægten *Lindtneria* indeholder ifølge den senest publicerede nogle 11 arter (Bernicchia & Ryvarden 1998). En gennemgang af litteraturen om slægten åbenbarer at hovedparten af disse er beskrevet ud fra enkelt-indsamlinger, og der er ikke publiceret revisioner af slægten baseret på en gennemgang af et større antal indsamlinger.

Hymenoforets udformning er en karakter der især på det seneste er blevet tillagt afgørende betydning ved adskillelsen af arter indenfor slægten (Hjortstam 1987, Larsen & Bernicchia 1990, Hansen & Knudsen 1997, Bernicchia & Ryvarden 1998) på trods af at det aldrig er undersøgt hvorvidt dette er holdbart. Alt i alt finder jeg at det er uhyre vanskeligt at gennemskue, hvor mange af de beskrevne arter, der reelt repræsenterer forskellige taxa.

Da jeg så fandt en *Lindtneria* som ikke passede på nogle af de anerkendte arter indenfor slægten, slog jeg efter nogen overvejelse koldt vand i blodet, og valgte ikke at nybeskrive endnu en tvivlsom art. Min indsamling afviger fra samtlige anerkendte arter i slægten ved de ganske små ellipsoidiske sporer (6-7 x 4-4.5 µm) og fra størsteparten af dem ved det glatte hymenie. *Lindtneria flava* og *L.*

*leucobryophila* (P. Henn.) Jülich er tilsvneladende de arter der kommer tættest på min indsamling. Begge arter, især sidstnævnte, er dog beskrevet med betydeligt større sporer. *L. flava* afviger desuden ved at være beskrevet med et poroidt hymenium. Da jeg finder det fornuftigt at ligge større vægt på mikroskopiske forskelle end på makroskopiske, har jeg valgt at anføre min kollektion som *L. cfr. flava*. Erast Parmasto (pers. comm.) mener udfra en beskrivelse, at min indsamling kan repræsentere "hans" art. I øvrigt anser jeg det for sandsynligt at *L. flava* og *L. leucobryophila* baserer sig på en og samme art, hvilket i fald min antagelse holder stik, vil give priorititet til navnet *L. leocubryophila*.

#### *Phlebia livida*

Eriksson et al. (1981) såvel som Vesterholt i Hansen & Knudsen (1997) angiver sporerne hos denne art til at måle 5-6 x 2-2.5 um, mens Christiansen (1959) angiver sporemålene 4-5 x 2 um. De indsamlinger jeg har tjekket har generelt haft sporer der målte ca. 4.5 x 2 um; dvs. De har passet perfekt med angivelserne hos Christiansen (1960) men mindre godt med de andre nævnte kilder.

#### *Sistotrema biggsiae*

Ikke tidligere angivet fra Danmark, og ikke udnøglet hos Hansen & Knudsen (1997). Tilhører artsgruppen omkring *S. brinkmannii*. Beskrevet udfra nordamerikansk materiale af Hallenberg (1984) på baggrund af parringsforsøg, understøttet af morfologiske karakterer. Materialets enlige indsamling er bestemt af Nils Hallenberg.

#### *Sistotrema brinkmannii*

Arts-gruppen omkring *S. brinkmannii*, blev revideret af Hallenberg (1984) på baggrund af omfattende parrings-forsøg og morfologiske studier. Arbejdet resulterede i adskillige nybeskrivelser. Kærne-arten fremstår dog stadig efter denne revision som et arts kompleks, snarere end som en veladskilt art (Hallenberg pers. komm.). I hvert fald én af materialets indsamlinger adskilte sig fra typisk *S. brinkmannii*, ved et helt glat hymenie.

#### *Tulasnella tomaculum*

Arten er ikke angivet som kendt fra Danmark hos Hansen & Knudsen (1997). Jan Vesterholt (pers. komm.) kender dog arten fra Høstemark Skov i Nordjylland. I Suserup Skov er arten tilsvneladende ganske udbredt, og er også samlet på ved af ask. Arten blev beskrevet af Roberts (1993), der angiver arten som almindelig.

#### Poresvampe

#### *Ceriporia excelsa*

Både hos Ryvarden & Gilbertson (1993) og Vesterholt i Hansen & Knudsen (1997) adskilles *C. excelsa* fra *C. viridians* på større porer og mere rødlige til pink farver. Hos sidstnævnte kilde anvendes dog også frugtlemets udformning som nøglekarakter; ofte vidt udbredt hos *C. viridians* - små og afgrænset hos *C. excelsa*. De fleste af mine indsamlinger har placeret sig lidt "mellemtostole" i forhold til disse kriterier, dog er jeg ikke i tvivl om at de alle har repræsenteret en og samme art. Mine specimens har generelt været ret små, med porer af intermediær størrelse og med pink til rødviolette farver, dog evt. blandet med creme til grønlige nuancer (sidstnævnte angives som typiske for *C. viridians*). Flest karakterer har således peget i retning af *C. excelsa*, hvorfor jeg konsekvent har valgt at bruge dette navn. Jeg kan dog kun tilslutte mig Ryvarden & Gilbertson (1993), der i noterne til *C. viridians* skriver at krydsningsforsøg er nødvendige for at afklare afgrænsningen de to arter imellem.

## Bladhatte

### *Armillaria gallica*

*A. gallica* tilhører en yderst vanskelig gruppe indenfor slægten. Gruppens arter er kendtegnede bla. ved en +/- kølleformet stok. Parringsforsøg er reelt nødvendige for en sikker artsbestemmelse indenfor gruppen, men materiale fra Suserup Skov passer generelt bedst på *A. gallica*, ifølge bestemmelseskriterierne angivet hos Marxmüller & Printz (1985; som *A. bulbosa*) og Watling i Hansen & Knudsen (1992).

Marxmüller & Printz (1985) fandt at *A. gallica* er meget almindelig i Danmark – og klart mere almindelig end de to mest oplagte forvekslingsmuligheder *A. borealis* (art A) og *A. cepistipes* (art B). Det stemmer godt overens med en senere dansk undersøgelse (Koch 1991), hvor der af de tre nævnte arter kun blev konstateret *A. gallica*, selv om flere afvigende kollektioner blev undersøgt. *A. borealis* og *A. cepistipes* er ifølge Koch (1991) slet ikke belagt med danske materiale. På den baggrund har jeg valgt konsekvent at bruge navnet *A. gallica* for honningsvampe af den køllestokkede type, uden forsøg på at udskille afvigende former i andre taxa vha. parringsforsøg mv.

### *Clitocybe diatreta*

Denne art, der forekommer ganske almindeligt på nedbrudte bøgestammer i Suserup Skov, har voldt mig en del kvaler. Indenfor slægten tillægges økologi en del betydning i afgrænsningen af arter (Gulden i Hansen & Knudsen 1992, Bas et al. 1995), og flere arter angives som decideret vedboende. Det gælder bl.a. *Clitocybe lignatilis* (Pers.: Fr) Karst., *C. truncicola* (Peck) Sacc. og *C. subbulbipes* Murril. Fra starten gjorde jeg meget for at henføre mine fund til et af disse taxa, men uden succes. De mikroskopiske karakterer var helt forkerte.

Da jeg derefter i stedet prøvede at bestemme mit materiale som en førne-nedbryder gik det meget bedre. Ved hjælp af nøglen hos Kuyper i Bas et al. (1995) nåede jeg utvetydigt frem til navnet *C. diatreta*, som passer på alle karakterer, undtagen økologi. Jeg ser dog ikke stor grund til at tvivle på rigtigheden af bestemmelsen på grund af denne afvigelse, da også andre typiske førne-nedbrydere indenfor slægten af og til danner frugtlegemer på bøgestammer, i hvert fald i Suserup Skov. Det gælder bl.a. for *C. phyllophila* og *C. nebularis*.

### *Galerina unicolor* s.l.

Petersen & Vesterholt (1990) udnøgler hele tre vedboende arter af *Galerina* med ring (*Galerina marginata* (Batsch) Kühner, *G. unicolor* og *G. autumnalis* (Peck) Smith & Singer), som navnlig adskilles på hathudsmorfologi. De ca. 10 indsamlinger, jeg har tjekket fra Suserup Skov har alle passet bedst på *G. unicolor*. Jeg har dog siden konferereret sagen med Gro Gulden (Oslo), som overfor mig har sagt, at hun tvivler på at det er rimeligt at anderledes mere end en art i gruppen, hvilket i øvrigt delvist fremgår af hendes nøgle til slægten hos Hansen & Knudsen (1992). Anses de tre navne som synonymer, har *G. unicolor* tilsyneladende prioritet, da det som ifølge Gulden i Hansen & Knudsen (1992) som det eneste er sanktioneret af Fries. Jeg har valgt at bruge navnet ukritisk for alle "store" træboende hjelmhatte med ring, uden studier af hathudsmorfologi mv. hos alle indsamlinger.

### *Lepiota boertmanii*

Kendt fra én indsamling i materialet, som venligst blev bestemt af Henning Knudsen (København). Hermed kendt fra to danske lokaliteter (Se Rald et al. 1992). Indsamlingen blev gjort i træsmuld på fugtig muld, tæt klinet op ad den tykke del af en nedbrudt stamme.

### *Lepiota ochraceofulva*

Denne i Danmark meget sjældne art er kendt fra to indsamlinger i materialet, men er alt i alt fundet ganske udbredt i Suserup Skov. Se i øvrigt Rald & Heilmann-Clausen (1993).

### *Omphalina epichysium*

Tilsyneladende ikke kendt fra andre danske lokaliteter end Suserup, men her ret udbredt. Hos Hansen & Knudsen (1992) er arten ikke angivet som kendt fra Danmark og det anses som tvivlsomt om den er arts-forskellig fra *O. oniscus*. Jeg finder dog at de to taxa med hensyn til økologi (*O. oniscus* forekommer i tørvemoser) er så forskellige at det er rimeligt at give dem begge artsrang, så længe intet andet er bevist. Mig bekendt findes der ikke et evt. analogt eksempel på én art med præference for begge de nævnte substrater. *O. epichysium* er meget almindelig i tempererede asiatiske naturskove (Thomas Læssøe pers. komm.).

### *Pluteus chrysorrheus*

Tilhører den uhyre vanskelige gruppe af skærmhatte omkring *P. phlebophorus*. Jeg har brugt navnet for indsamlinger med varme, ret lyse gulbrune hatfarver og en +/- cocnagfarvet stok (+/- som tavle 72D hos Lange (1937)). Jeg er dog ikke overbevidst om at denne form virkelig er artsforskellig fra *L. phlebophorus*.

### *Pluteus ephabeus*

Navnet dækker nærmere over et arts kompleks end en art. Her er fulgt det meget brede arts konsept hos Vellinga i Bas et al. (1990) og Døssing i Hansen & Knudsen (1992). Følges Orton (1986), som har en meget snæver artsopdeling, passer materialets indsamlinger bedst på *P. murinus* Bres.

### *Pluteus godeyi*

Kendt fra én indsamling i materialet. Betragtes af Vellinga i Bas et al. (1990) som et synonym af *P. cinereofuscus* Lange, men adskilles hos Døssing i Hansen & Knudsen (1992) på at være mindre og hygrofan, dog med en note om at der sandsynligvis blot er tale om en lille form af *P. cinereofuscus*. At jeg har valgt at bruge navnet *P. godeyi* skal ikke tages som udtryk for en kritisk stillingstagen til artsafgrænsningen, blot at min indsamling passede bedst på dette navn i Hansen & Knudsen (1992). Interresant nok har jeg aldrig set typisk *P. cinereofuscus* i Suserup Skov.

### *Pluteus nanus*

Ifølge Vellinga i Bas et al. (1990) er denne art let at adskille fra arterne omkring *P. phlebophorus* på grund af hathudens afvigende struktur. Jeg har haft ret svært ved at erkende denne forskel, og har baseret mine bestemmelser på makroskopiske karakterer, navnlig hatfarve og statur. Ifølge mine erfaringer er *P. nanus* generelt mindre, mere kortstokket og betydeligt mørkere på hatten end *P. phlebophorus*-gruppens arter. Derimod finder jeg ikke hattens struktur (radiærrynket eller glat) som af nogle (fx. Petersen & Vesterholt 1990, Døssing i Hansen & Knudsen 1992) angives som skillekarakter mellem *P. nanus* og *P. phlebophorus* pålidelig.

Else Vellinga har ved en tidligere lejlighed bestemt materiale fra Suserup Skov til *P. nanus*.

*Pluteus phlebophorus*

"Hovedarten" i *P. phlebophorus*-gruppen og som sådan mest defineret udfra negative karakterer. Mit koncept af arten stemmer rimeligt godt overens med det der anvendes af Vellinga i Bas et al. (1990). I modsætning til hende lægger jeg dog ikke vægt på, hvorvidt hatten er rynket eller ej, da jeg finder denne karakter ustabil. Se også *P. chrysophaeus*.

*Pluteus semibulbosus*

Af Vellinga i Bas et al. (1990) synonymiseret med *P. plautus*. Jeg finder dog at denne disposition er udtryk for en overforsimpling, selv om jeg dog må medgive at det ofte er meget vanskeligt at skelne mellem de mange arter der accepteres i gruppen omkring *P. plautus*, ikke mindst hos Orton (1986). Alle specimens jeg har set i Suserup Skov har dog været af samme type (ofte forholdsvis kortstokkede og uregelmæssige med lyse farver; hat creme til lyst grålig brun eller honninggul. - Meget lig Lange 1937: tab 71E og Phillips 1981: 119). *P. plautus* s.str. beskrives traditionelt som en meget mørk art (c.f. Orton 1986, Døssing i Hansen & Knudsen 1992. Hos Hansen & Knudsen (1992) nøgles uden problemer ud i *P. semibulbosus*.

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Appendix 3. Registreringer af arter på stammer, sorteret efter frekvens (antal stammer) og stammenummer. Bemærk at arter fundet på to eller færre stammer ikke er medtaget i ordinationen.

frekvens	B3NE-B1	B3SW-B11	B3SW-B13	B4SE-B8	C2NE-B1	C2NE-B8	C3NE-B3	C3NW-B7	C3NW-B8	C3SE-B12	B3	C4NW-B1	C4NW-B3	C4SE-B9	D2NE-B16	D2NE-B12	D2NE-B8	D2NE-B3	D2NE-B8	D2NE-B20	D2NE-B4	D2NE-B8	D3NE-B1	D3NE-B2	D3NE-B5	D3NE-B6	D3NW-B1
ara van	1																										
arc affi	1																										
arm meli	1																										
ath glau	1																										
aur aur	1																										
bas caes	1																										
bje fumo	1																										
bot boit	1																										
coi fusi	1																										
col pero	1																										
com alla	1																										
con sord	1																										
cop ech	1																										
cop lago	1																										
cop xant	1																										
cys adul	1																										
cys cono	1																										
cys hei	1																										
dac sil	1																										
das nive	1																										
dia flav	1																										
dia stig	1																										
ent dich	1																										
exi eflu	1																										
fla mur	1																										
gal nana	1																										
gan pfei	1																										
gio lun	1																										
hetero	1																										
hyp aspe	1																										
hyp eich	1																										
hyp medi	1																										
hyp samb	1																										
ino peti	1																										
lac lacc	1																										
lac papy	1																										
lep boer	1																										
lep jaco	1																										
lep ruda	1																										
lin flav	1																										
lop lurg	1																										
lyc echi	1																										
mar coha	1																										
mar wynn	1																										
mel hori	1																										
micr bras	1																										
mol lign	1																										
myc uda	1																										
nec epis	1																										
ombrop	1																										
pen nuda																											
pen quer	1																										
pha sord	1																										
pha tube	1																										
pha velu	1																										
phi alia	1																										
phy cine	1																										

Appendix 3. Registreringer af arter på stammer, sorteret efter frekvens (antal stammer) og stammenummer. Bemærk at arter fundet på to eller flere stammer ikke er medtaget i ordinatoren.

frekvens	B3NE-B	B3SW-B	B3SW-B	B4SE-B	C2NE-B	C2NE-B	C3NE-B	C3NW-	C3SE-B	C3SE-B	C3SW-B	C4NW-	C4SE-B	D2NE-B	D2NE-B	D2NE-B	D2NW-	D2SE-B	D3NE-B	D3NE-B	D3NW-
ple dryi	1																				
plu gode	1																				
plu podo	1																				
pol badi	1																				
protocre	1																				
psa cand	1																				
psa cono	1																				
psa frus	1																				
psa fusc	1																				
psa lacr	1																				
psa popu	1																				
psa pygm	1																				
psa spad	1																				
psa leph	1																				
ram cent	1																				
res tric	1																				
rip tric	1																				
sis ligg	1																				
sis obio	1																				
ste hype	1																				
lap livi	1																				
iom ferr	1																				
iom radi	1																				
re micr	1																				
re prae	1																				
ri cont	1																				
ri flor	1																				
ri lasc	1																				
ri pers	1																				
tub ferr	1																				
ui eich	1																				
tyr chio	1																				
tyr wynn	1																				
vol hypo	1																				
xen pulv	1																				
ant hoeh	2																				
anti semi	2																				
arc inca	2																				
bot laev	2																				
cam tubu	2																				
co tube	2																				
com nigr	2																				
con and	2																				
corticu	2																				
cri gall	2																				
cri helv	2																				
eua quat	2																				
hyp argu	2																				
hyp pall	2																				
hyp polo	2																				
lep ochr	2																				
mel haem	2																				
mer anom	2																				
met vesp	2																				
myc erub	2																				
myc poly	2																				
rec pezi	2																				

Appendix 3. Registreringer af arter på stammer, sorteret efter frekvens (antal stammer) og stammenummer. Bemærk at arter fundet på to eller færre stammer ikke er medtaget i ordinatenionen.

frekvens	B3NE-B	B3SW-B	B3SY-B	B4SE-B	C2NE-B	C3NE-B	C3NW-	C3SE-B	C3SE-B	C3SW-B	C4NW-	C4NW-	D2NE-B	D2NE-B	D2NE-B	D2NE-B	D2NE-B	D3NE-B	D3NE-B	D3NE-B	D3NW-
orb aine	2																				1
orb epis	2																				1
phi rufa	2																				1
phi squa	2																				1
phi ephie	2																				1
sis nive	2																				1
sis octo	2																				1
ska nive	2																				1
sym amau	2																				1
tra gibb	2																				1
tub furf	2																				1
ara aura	3																				1
arc cine	3																				1
bys coni	3																				1
cli phyl	3																				1
con brun	3																				1
con tene	3																				1
cop domne	3																				1
exi nud	3																				1
gio anal	3																				1
gym junio	3																				1
hem cucc	3																				1
hya fuck	3																				1
hyp coha	3																				1
hyp punc	3																				1
hyp suba	3																				1
iam arcy	3																				1
mut cani	3																				1
myc amic	3																				1
myc hiem	3																				1
pen cine	3																				1
pen lyci	3																				1
phi thom	3																				1
pol brum	3																				1
pol cili	3																				1
pol squa	3																				1
pol tube	3																				1
sis coro	3																				1
sir cyan	3																				1
tul loma	3																				1
uth fusi	3																				1
arc denu	4																				1
exi glan	4																				1
her cora	4																				1
hyp aura	4																				1
hyp gelia	4																				1
hyp rose	4																				1
iac subd	4																				1
iac virg	4																				1
lep fulv	4																				1
myc galio	4																				1
myc rose	4																				1
nyc speli	4																				1
nec cocc	4																				1
rem atra	4																				1
phi inqu	4																				1
phi semi	4																				1

Appendix 3. Registreringer af arter på stammer, sorteret efter frekvens (antal stammer) og stammenummer. Bemærk at arter fundet på to eller færre stammer ikke er medtaget i ordinationen.

Appendix 3. Registreringer af arter på stammer, sorteret efter frekvens (antal stammer) og stammenunder. Bemærk at arter fundet på to eller flere stammer ikke er medtaget i ordinationen.

Appendix 3. Registreringer af arter på stammer, sorteret efter frekvens (antal stammer) og stammenummer. Bemærk at arter fundet på to eller færre stammer ikke er medtaget i ordinationen.

	frekvens	B3NE-B	B3SW-B	B3SW-B	B4SE-B	C2NE-B	C3NE-B	C3NW-	C3SE-B	C3SE-B	C3SW-B	C4NW-	C4NW-	D2NE-B	D2NE-B	D2NE-B	D2NE-B	D2NE-B	D2NE-B	D3NW-
bot cons	29	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
scu scut	29	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
tri vari	29	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
hyp fasc	35	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
lyc epid	35	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
myc croc	35	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
cap mica	36	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
lyc pyri	36	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
ath epip	37	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
plu cerv	37	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
cer reti	38	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
fom forme	38	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
mar alli	40	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
xyl hypo	42	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
am gall	44	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
kre deus	45	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
mol cine	49	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
myc haem	50	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
eut spin	62	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1

Appendix 3. Registreringer af arter på stammer, sorteret efter frekvens (antal stammer) og stamnummer. Bemærk at arter fundet på to eller færre stammer ikke er medtaget i ordinatenionen.

frekvens	D3NW-B4	D3NW-B5	D3NW-B8	D3SE-B12	D3SW-B3	D3SW-B4	D4NE-B1	D4NW-B11	D4SE-B13	E2NE-B11	E2NE-B13	E3NE-B16	E3NW-B5	E3NW-B8	E3SE-B10	E3SW-B2	E3SW-B4	E4SW-B7	F2NE-B9	F2NW-B1	F3NW-B3	F3NW-B7	
ara vani	1																						
arc affi	1																						
arm mel																							
ath glau	1																						
aur aur	1																						
bas caes	1																						
bie fumo	1																						
bot botr	1																						
col fusi	1																						
col pero	1																						
com alia	1																						
con sord	1																						
cop echii	1																						
cop lago	1																						
cop xant	1																						
cys adul	1																						
cys cono	1																						
cys helii	1																						
dac sili	1																						
das nive	1																						
dia flav	1																						
dia stig	1																						
ent dich	1																						
exi erfu	1																						
fla mun	1																						
gal nana	1																						
gan plei	1																						
gio lunii	1																						
helero	1																						
hyp aspe	1																						
hyp eich	1																						
hyp medi	1																						
hyp samb	1																						
ino peti	1																						
lac iacc	1																						
lac papy	1																						
tep boer	1																						
tep iaco	1																						
lep nuda	1																						
lin lav	1																						
lop turq	1																						
lyc echii	1																						
mar coha	1																						
mar wynn	1																						
mel horii	1																						
micr bras	1																						
mol lign	1																						
myc iuda	1																						
nec epis	1																						
ombrop	1																						
pen nuda	1																						
pen quer	1																						
pha sord	1																						
pha tube	1																						
pha velu	1																						
phi alla	1																						
phy cine	1																						

Appendix 3. Registreringer af arter på stammer, sorteret efter frekvens (antal stammer) og stammenummer. Bemærk at arter fundet på to eller færre stammer ikke er medtaget i ordinationen.

Frekvens	D3NW	D3NW-	D3SW	D3SW-	D3SE	D3SE-B	D3SW-B	D3SW-B	D4NE	D4NE-B	E2NE	E2NE-B	E3SE	E3SE-B	E3SW	E3SW-B	E3NW	E3NW-B	F2NE	F2NE-B	F3NW	F3NW-B
pie dry	1																					
plu gode	1																					
plu podo	1																					
pol badi	1																					
protoce	1																					
psa cand	1																					
psa cono	1																					
psa frus	1																					
psa fusc	1																					
psa lacr	1																					
psa popu	1																					
psa pygm	1																					
psa spad	1																					
psa leph	1																					
ram cent	1																					
res tric	1																					
rip tric	1																					
sis biog	1																					
sis oblo	1																					
ste hype	1																					
tap livi	1																					
tom terr	1																					
tom radi	1																					
tre micr	1																					
tre prae	1																					
tri cont	1																					
tri flor	1																					
tri lasc	1																					
tri pers	1																					
tub ferr	1																					
tul eich	1																					
tyr chio	1																					
yr wynn	1																					
voi hypo	1																					
xen pulv	1																					
ant hoeh	2																					
ani semi	2																					
arc inca	2																					
boot baev	2																					
cam tubu	2																					
col tube	2																					
com nigr	2																					
con and	2																					
conficiu	2																					
cni gall	2																					
cri helv	2																					
eut quat	2																					
hyp argu	2																					
hyp pali	2																					
hyp polo	2																					
hyp ochi	2																					
mei haem	2																					
mer anom	2																					
met vesp	2																					
myc enub	2																					
myc poly	2																					
rec pezi	2																					

Appendix 3. Registreringer af arter på stamme, sorteret efter frekvens (antal stammer) og stammenummer. Bemærk at arter fundet på to eller færre stammer ikke er medtaget i ordinatenen.

	Drekvens	D3NW-	D3NW-	D3NW-	D3NW-	D3SE-B	D3SW-B	D3SW-B	D4NE-B	D4NW-	D4SE-B	E2NE-B	E2NE-B	F2NE-B	F2NW-B	F3NW-B	F3NW-B
orb alne	2																
orb epis	2																
phi rula	2																
phi squa	2																
phi sphæ	2																
sis nive	2																
sis octo	2																
ske nive	2																
sym amau	2																
tra glibb	2																
tub furf	2																
ara aura	3																
arc cine	3																
bys cori	3																
cli phyl	3																
con brun	3																
con tene	3																
cop dome	3																
exi nuci	3																
gio anal	3																
gym junio	3																
hem cucc	3																
hya fuck	3																
hyp coha	3																
hyp punc	3																
hyp suba	3																
lam arcy	3																
mut cani	3																
myc amic	3																
myc hiem	3																
pen cine	3																
pen lyci	3																
phi thom	3																
pol brun	3																
pol cili	3																
pol equa	3																
pol tube	3																
sis coro	3																
stir cyan	3																
tui loma	3																
utu tusi	3																
arc denu	4																
exi gian	4																
her cora	4																
hyp aura	4																
hyp gelia	4																
hyp rose	4																
iac subd	4																
iac virg	4																
lep tulv	4																
myc galø	4																
myc rose	4																
myc spei	4																
nec cocc	4																
phi atro	4																
phi inqu	4																
phi semi	4																

Appendix 3. Registreringer af arter på stammer, sorteret efter frekvens (antal stammer) og stammenummer. Bemærk at arter fundet på eller færre stammer ikke er medtaget i ordinatenionen.

	frekvens	D3NW-	D3NW-	D3SE-B	D3SW-B	D3SW-B	D3SW-B	D4NE-B	D4NW-	D4SE-B	E2NE-B	E3NE-B	E3NW-B	E3NW-B	E3SE-B	E3SW-B	E3SW-B	E3SW-B	F2NE-B	F2NE-B	F3NW-B	F3NW-B	F3NW-B
psa rost	4																						
scy rimo	4																						
sty gril	4																						
sty subn	4																						
tre hymne	4																						
tui viol	4																						
bot subc	5	1																					
ctr argi	5																						
dia disc	5		1																				
fus ferr	5																						
hen cand	5																						
hyd suba	5																						
lac brev	5																						
myc peli	5																						
pha impu	5																						
plu ranu	5																						
plu umbr	5																						
sch comm	5			1																			
sis seen	5	1																					
lori subl	5																						
bot prui	6																						
cal corn	6																						
dat moll	6			1																			
tom pini	6																						
hyp argi	6																						
hyp para	6		1																				
hyp prae	6																						
ino nodu	6																						
lep aspe	6																						
myc dios	6																						
nem serp	6																						
neo pura	6																						
phi trem	6																						
pho aur	6																						
ste tusc	6																						
ste lyph	6																						
bol reli	7																						
cyl aev	7																						
lep cris	7																						
myc tint	7																						
ste axif	7																						
ste rugo	7																						
tra hirs	7																						
tre fan	7																						
tre mese	7																						
tre slev	7	1																					
asc cyli	8																						
cer exca	8		1																				
coi buty	8																						
omp epic	8																						
pan sero	8																						
pen inca	8	1																					
plu sali	8																						
psa pliu	8																						
bre oliv	9																						
cam poly	9																						
cer golv	9																						

Appendix 3. Registreringer af arter på stammer, sorteret efter frekvens (antal stammer) og stammenummer. Bemærk at arter fundet på to eller færre stammer ikke er medtaget i ordinationen.

frekvens	D3NW-1	D3NW-1	D3SW-B	D3SW-B	D3SW-B	D4NE-B	D4NW-	D4SE-B	E2NE-B	E3NE-B	E3NE-B	E3NW-B	E3NW-B	E3SW-B	E3SW-B	E3SE-B	E3SW-B	E3SW-B	E3NW-B	F2NW-B	F3NW-B	F3NW-B
cli neb	9	1							1	1						1	1				1	1
hyp sei																						
mar lorq	9	1																				
nem ches	9	1																				
pse cyat	9																					
ite cona	9																					
bis citr	10																					
cli hobs	10																					
phi livi	10																					
ple osie	10																					
tri seab	10																					
xer radi	10																					
myc rena	11																					
phi fagi	11																					
plu chry	11																					
mac mac	12	1																				
pol priui	12	1																				
sub long	12																					
lyc perl	13																					
phi radi	13																					
pol vari	13																					
isc resi	14	1																				
lep flacc	14																					
oud mucu	14																					
phy vitt	14																					
hyp frag	15	1																				
hyp rubi	15																					
kue muta	15	1																				
ram stri	15	1																				
hyp radu	16																					
phy sang	16																					
ste ochr	16																					
bje adus	17																					
con pule	17																					
hyp pube	17	1																				
myc pura	17	1																				
xylo poly	17	1																				
ent lyco	18	1																				
mer giga	18	1																				
fui septi	19																					
glo lact	19	1																				
orb deli	19	1																				
psa obliu	19	1																				
rad conf	20	1																				
sis brin	20																					
tra vers	20	1																				
cli diat	21	1																				
ste hirs	21	1																				
gan los	22	1																				
lac impu	22	1																				
meg pliat	22	1																				
plu phile	22	1																				
cer fruc	24																					
gal unic	24																					
pez micr	26	1																				
mar rotu	27	1																				
myc gate	28																					

Appendix 3. Registreringer af arter på stammer, sorteret efter frekvens (antal stammer) og stammenummer. Bemærk at arter fundet på to eller flere stammer ikke er medtaget i ordinationen.

Appendix 3. Registreringer af arter på stammer, sorteret efter frekvens (antal stammer) og stammenummer. Bemærk at arter fundet på to eller færre stammer ikke er medtaget i ordinationen.

	F3SW- B8	F4SW- B10	G3NE- B13	G3NW- B9	G3SW- B1	G3SW- B3	H3NE- B10	H3NE- B6	H3NW- B2	H3NW- B1	I3NW- B4	I3NW- B5	I3SE-B4	I3SE-B9	I3SW-B4
ara van	1														
arc affi	1														
arm mell	1														
ath gau	1														
aur aur	1														
bas caes	1														
bije fumo	1														
bot botr	1														
col fusi	1						1								
col pero	1														
com alla	1														
con sord	1														
cop ech	1														
cop lago	1														
cop xant	1														
cys adul	1														
cys cono	1														
cys heli	1														
dec stili	1														
das nive	1														
dia flav	1														
dia stig	1														
ent dich	1														
exi enfu	1														
fla muri	1														
gal nana	1														
gan piei	1														
gio un	1														
hetero	1														
hyp aspe	1														
hyp eich	1														
hyp medi	1														
hyp samb	1														
ino peli	1														
lac lacc	1														
lac papy	1														
lep boer	1														
lep jaco	1														
lep nuda	1														
lin flav	1														
lop lurg	1														
lyc ech	1														
mar coha	1														
mar wynn	1														
mel hori	1														
mic bras	1														
mol lign	1														
myc uda	1														
nec eps	1														
ombrop	1														
pen nuda	1														
ren quer	1														
pha sord	1														
pha tubo	1														
pha velu	1														
phi alia	1														
phy cine	1														

Appendix 3. Registreringer af arter på stammer, sorteret efter frekvens (antal stammer) og stammenummer. Bemærk at arter fundet på to eller færre stammer ikke er medtaget i ordinatenionen.

	frekvens	F3SW-B	F4SW-B	G3NE-B	G3NW-	G3SW-B	G3SW-BH3NE-B	H3NE-B	H3NW-	I3NW-B	I3NW-B	I3SE-B9	I3SE-B4	I3SW-B4
ple dry	1													
plu gode	1													
plu podo	1			1										
pol badi	1													
protocre	1													
psa cand	1													
psa cono	1													
psa fus	1													
psa fusc	1													
psa lacr	1													
psa popu	1													
psa pygm	1													
psa spad	1													
psa leph	1													
ram cent	1													
res tric	1													
rip tric	1													
sis bligg	1													
sis oblo	1													
ste hype	1													
tap livi	1													
tom ferr	1													
tom radi	1													
tre minor	1													
tre pree	1													
tri cont	1													
tri flor	1													
tri lasc	1													
tri pers	1													
tub ferr	1													
tul eich	1													
tyr chio	1													
tyr wynn	1													
vo hypo	1													
xen pulv	1													
ant hoeh	2													
ant semi	2													
arc inca	2													
bot baev	2													
cam tubu	2													
coi tube	2													
com nigr	2													
con and	2													
corticu	2													
cri gall	2													
cn helv	2													
eut quat	2													
hyp argu	2													
hyp pall	2													
hyp polo	2													
lep ochr	2													
mel taem	2													
mer aniom	2													
met vesp	2													
myc erub	2													
myc poly	2													
nec pezi	2													

Appendix 3. Registreringer af arter på stammer, sorteret efter frekvens (antal stammer) og stammenummer. Bemærk at arter fundet på to eller færre stammer ikke er medtaget i ordinatenionen.

	Frekvens	F3SW-B	F4SW-B	G3NE-B	G3NW-	G3SW-B	G3SW-B/H3NE-B	H3NW-	H3NW-B	H3NW-B	H3NW-B	H3NW-B	H3SE-B9	I3SW-B4
orb alne	2													
orb epis	2													
phi rufa	2													1
pho squa	2													
phi sphé	2													
sis nive	2													
sia octo	2													
ske nive	2													
sym amau	2													
tra gibb	2													1
tub turf	2													
ara aura	3													
arc cine	3													
bys con	3													
cli ptyl	3													
con brun	3													
con tene	3	1												
cop dome	3													
exi nuci	3													
gio anal	3													
gym junio	3													
hem cucc	3													
nya fuck	3													
hyp coha	3													
hyp punc	3													
hyp suba	3													
iam arcy	3													
mut cani	3													
myc amic	3													
myc hiem	3													
pen cine	3													
pen lyci	3													
phi thom	3													
pol brum	3													
pol cili	3													1
pol squa	3													
pol tube	3													1
sis coro	3													
sir cyan	3													
sil loma	3													
uh fusi	3													
arc denu	4													
exi glan	4													
her cora	4													
hyp aura	4													
hyp gelia	4													
hyp rose	4													1
iac subd	4													
iac virg	4													
lep fulv	4													
myc gallo	4													
myc rose	4													
myc spie	4													
nec cocc	4													
rem alto	4													1
phi inqu	4													
phi semi	4													

Appendix 3. Registreringer af arter på stammer, sorteret efter frekvens (antal stammer) og stammenummer. Bemærk at arter fundet på to eller færre stammer ikke er medtaget i ordinationen.

	F3SW-B	F4SW-B	G3NE-B	G3NW-	G3SW-B	G3SW-B/H3NE-B	H3NE-B	H3NW-	I3NW-B	I3NW-B	I3NW-B	I3SE-B	I3SE-B	I3SW-B
psa rost	4								1					
scō imo	4								1					
sty gril	4								1					
sty subn	4													
tre rhyme	4													
tui viol	4													
bot subc	5									1				
cn̄ argi	5													
dia disc	5													
fus ferr	5								1					
hen cand	5	1								1				
hyd suba	5													
lac brev	5													
mys peili	5	1												
pha impu	5													
plu nanu	5	1												
plu umbr	5	1												
sch comm	5													
sis sem	5													
torn subl	5								1					
bot priū	6													
cal com	6													
dat moll	6													
torn pini	6								1					
hyp argi	6													
hyp para	6													
hyp prae	6													
ino nodu	6								1					
lep aspe	6													
myc dios	6													
nem serp	6													
neo pura	6								1					
phi trem	6	1												
pho aur	6													
ste tusc	6													
ste lyph	6													
bol reii	7													
cyl eav	7								1					
lep cris	7													
myc lant	7													
ste axif	7	1												
ste rugo	7													
tra his	7													
tre fari	7													
tre mese	7								1					
tre stav	7													
asc cyli	8								1					
cer exce	8								1					
col bulv	8													
omp epic	8	1												
pan sero	8								1					
pen inca	8													
plu sali	8													
psa pilu	8													
bre oliv	9	1												
cam poly	9													
cer gilv	9													

Appendix 3. Registreringer af arter på stammer, sorteret efter frekvens (antal stammer) og stammenummer. Bemærk at arter fundet på to eller færre stammer ikke er medtaget i ordinaten.

Frekvens	F3SW-B	F4SW-B	G3NE-B	G3NW-	G3SW-B	G3SW-B	H3NE-B	H3NW-	H3NW-B	I3NW-B	I3NW-B	I3SE-B9	I3SW-B4
cli nebu	9	1										1	1
hyp seti	9	1										1	1
mar loriq	9				1								
nem ches	9	1											
pse cyat	9												
tre coha	9												
bis citr	10	1										1	1
cli hobs	10											1	1
phi livi	10								1				
pile osie	10								1			1	
tri seab	10								1				
xer radi	10											1	
myo rena	11								1			1	
phi fagi	11								1				1
plu chry	11												
mac mac	12											1	
pol pnu	12				1							1	1
sub long	12	1										1	
phi radi	13		1						1				
pol vari	13	1										1	
isc resi	14	1											
lep nacc	14												
oud muci	14								1			1	1
phy vlt	14	1											
hyp frag	15	1	1	1					1				1
hyp rubi	15	1										1	
kue muta	15	1											
ram stri	15												
hyp radu	16	1										1	1
phy sang	16		1	1									
ste ochr	16		1	1									1
bje acus	17		1	1					1				1
con pule	17								1			1	1
hyp pube	17	1			1				1			1	1
myc pura	17	1							1			1	1
xy poly	17			1					1				
ent lyco	18						1					1	
mer giga	18											1	
ful sept	19												
gio lact	19												
orb deli	19												
psa ootu	19												
rad conf	20								1			1	1
sis bin	20	1										1	
tra vers	20		1									1	1
cli diat	21		1	1					1			1	1
ste hirs	21		1	1								1	
gan lips	22		1									1	
lac impu	22	1											
meg piat	22								1			1	1
plu phle	22	1										1	
cer fruc	24	1										1	
gal unc	24								1				
pez micr	26	1										1	
mar rotu	27								1				
myc gate	28	1							1			1	1

Appendix 3. Registreringer af arter på stammer, sorteret efter frekvens (antal stammer) og stammenummer. Bemærk at arter fundet på to eller færre stammer ikke er medtaget i ordinationen.

	F3SW-B	F4SW-B	F3NE-B	G3NW-	G3SW-B	B1H3NE-B	H3NE-B	H3NW-B	I3NW-B	I3NW-B	I3SE-B	I3SE-B	I3SW-B
bot cons	29	1			1		1		1		1		1
scu scut	29	1			1		1		1		1		1
tri van	29	1			1		1		1		1		1
hyp fasc	35	1			1		1		1		1		1
lyc epid	35	1			1		1		1		1		1
myc eroc	35	1			1		1		1		1		1
cop mica	36	1			1		1		1		1		1
lyc pyri	36				1		1		1		1		1
ath epip	37						1		1		1		1
plu cerv	37	1					1		1		1		1
cer reti	38	1					1		1		1		1
fom forme	38	1					1		1		1		1
mar alli	40	1					1		1		1		1
xyl hypo	42	1					1		1		1		1
arm gall	44						1		1		1		1
kre deus	45						1		1		1		1
mol cine	49	1					1		1		1		1
myc haem	50	1					1		1		1		1
eut spin	62	1					1		1		1		1

#### Appendix 4. Variabelscorer for samtlige stammer medtaget i ordinationen.

Appendix 4. Variabelscorer for samtlige stammer medtaget i ordinationen.

	D2NE-B8	D2NW-B20	D2SE-B4	D3NE-B1	D3NE-B2	D3NE-B5	D3NW-B1	D3NW-B4	D3NW-B5	D3SE-B12	D3SW-B1	D3SW-B3	D3SW-B4	D3SE-B1	D3NW-B1	DANE-B1	D3NW-B8	D3SE-B11	D3SE-B1	D3NW-B1
DCA1	223,7478	96,72174	15,80413	310,5948	176,0605	138,9038	89,27057	85,71695	279,9225	99,57628	119,0126	233,2131	91,80602	87,68096	104,3657	70,93513	82,21997	85,39236		
DCA2	96,39819	95,58926	210,3806	72,18427	64,73701	60,63694	98,02598	155,3644	69,50249	0	67,20736	61,13186	94,16202	113,2926	173,7722	44,61137	73,55215	192,638		
DCA3	133,8882	45,91608	89,60825	74,23377	83,30868	67,07516	176,69004	84,6374	106,0672	116,6493	171,1835	8,34406	77,88853	55,71993	50,62612	92,92442	99,67258	26,87848		
fractures	3	2	3	2	3	3	0	1	1	4	1	1	1	1	1	1	4	4	1	
soil contact	70	100	80	90	90	100	50	30	80	100	70	100	70	100	70	100	100	100	100	
moss cover	5	5	10	5	5	5	10	5	5	0	10	10	5	5	5	5	5	5	5	
bark cover	80	0	90	80	60	0	10	90	20	10	90	0	0	0	10	0	20	5	0	
erytia cover	20	30	10	10	20	10	40	30	5	40	10	0	10	10	40	40	30	30	20	
plant cover	90	90	40	90	90	90	80	100	100	90	90	50	100	100	90	70	70	50	50	
decay stage	1	4	5	1	2	3	5	4	2	3	1	4	4	4	4	4	4	4	5	
log age	5	31	4	7	9	27	20	7	15	13	4	27	27	27	27	27	27	27	27	
dbh	90	115	120	130	132	120	130	114	104	112	100	100	80	100	144	100	112	100	112	
dist. to edge	60	60	15	100	110	130	130	120	110	110	110	105	90	70	65	75	15	55		
plant diversity	4	1	3	1	2	1	2	0	0	0	10	30	10	0	10	0	1	1	1	
floristic deviance	0	0	20	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
bole forks	2	2	1	1	2	2	0	1	1	1	2	2	0	2	0	2	0	2	3	
soil humidity	3	3	4	3	2	2	2	3	3	2	3	3	3	3	2	2	3	4	3	
wind exposure	2	2	2	3	3	3	4	4	4	4	4	4	3	3	2	2	3	3	2	
sun exposure	2	3	1	2	3	4	3	3	3	3	3	3	5	3	2	2	5	4	5	
soil type																				
loamtil	loamtil	loamtil	loamtil	loamtil	loamtil	loamtil	loamtil	loamtil	loamtil	loamtil	loamtil	loamtil	loamtil	loamtil	loamtil	loamtil	loamtil	loamtil	loamtil	
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Appendix 4. Variabelscorer for samtlige stammer medtaget i ordinationen.

	E2NE-B11	E2NE-B12	E3NE-B13	E3NE-B16	E3NW-B2	E3NW-B5	E3NW-B8	E3SE-B10	E3SE-B4	E3SW-B2	E3SW-B7	E4NW-B1	F2NE-B7	F2NE-B9	F2NW-B1	F3NW-B3	F3NW-B5	F3NW-B7	F3NW-B8
DCA1	102,0213	261,0073	85,96357	184,1914	91,40102	133,8997	107,1527	78,7499	93,9187	189,9385	87,28396	128,7587	260,9599	138,1251	85,63653	62,39252	74,39278		
DCA2	74,83033	23,45268	80,14588	50,93615	71,24533	116,311	55,58676	72,03576	59,8272	82,15681	57,60583	163,9562	81,75935	123,654	67,92207	50,52287	7,0206	28,04924	
DCA3	86,05448	129,087	75,81738	64,73048	86,00787	60,03586	65,33133	145,8854	106,7471	88,22461	40,88825	0	109,7064	111,9407	124,1586	134,2707	37,14113	91,04132	
fractures	4	0	2	1	5	3	4	1	5	2	1	3	1	3	4	3	3	3	3
soil contact	100	60	100	100	80	70	80	100	90	100	100	100	100	100	50	60	90	100	100
moss cover	10	5	30	10	5	10	50	30	10	0	5	5	0	0	5	10	10	5	
bark cover	10	100	0	80	10	40	50	0	5	70	0	10	0	100	20	60	0	40	
eytyna cover	30	30	10	5	20	40	10	0	60	70	0	0	40	10	50	20	20	10	10
plant cover	80	100	90	80	100	90	90	100	100	70	70	70	70	70	90	100	90	100	100
decay stage	4	1	5	2	4	3	3	4	3	2	5	5	4	1	3	3	5	3	3
log age	20	31	3	4	16	11	13	27	16	7	20	31	20	2	13	13	16	15	
dbh	118	90	104	100	144	134	132	140	138	108	106	110	74	98	106	120	120	110	
dist. to edge	45	50	135	120	110	130	135	100	115	90	135	15	40	40	60	130	120	105	
plant diversity	2	2	1	1	2	2	1	3	3	2	1	1	3	2	1	4	1	1	
floristic deviance	10	10	0	0	10	0	0	0	0	20	0	0	20	10	0	0	0	0	0
bole forks	3	1	1	0	2	3	3	1	3	1	0	0	0	1	1	2	2	1	
soil humidity	4	4	3	3	3	2	3	3	3	3	3	3	3	3	2	3	4	3	
wind exposure	4	4	4	4	5	4	5	4	5	4	5	2	3	2	2	4	4	4	
sun exposure	4	4	3	2	4	2	4	4	5	2	4	3	2	3	2	3	3	2	
soil type	lac sand	loamill	loamill	loamill	loamill	loamill	loamill	lac sand	lac sand	loamill	loamill	lac clay	lac clay	lac clay	loamill	loamill	loamill	loamill	
log type	broken in 1-7 m. height	broken in 8-15 m. height	broken in 1-7 m. height	broken in 8-15 m. height	broken at root neck	broken at root neck	broken at root neck	broken in 1-7 m. height	broken in 1-7 m. height	broken at root neck									
decay rate	0	0	2	0	0	0	0	2	0	-1	0	2	0	2	0	0	0	0	2
ane ran	10	10	0	0	0	0	0	0	5	10	0	0	0	0	0	0	0	0	
ane nem	70	90	80	80	100	90	90	80	80	80	70	50	60	90	100	90	100	100	
cor bul	0	0	0	0	0	0	0	0	0	0	20	0	0	0	5	0	0	0	
men per	0	0	0	0	10	5	0	0	0	0	0	0	0	0	5	0	0	0	
ran fic	0	0	0	0	0	0	0	5	10	0	0	0	0	0	5	0	0	0	
lam gal	0	0	0	0	0	0	0	0	0	0	0	0	20	10	0	0	0	0	
gal odo	0	0	0	0	0	0	0	0	0	0	0	5	0	0	0	0	0	0	
vio rin	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
other plants	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	

Appendix 4. Variabelscorer for samtlige stammer medtaget i ordinationen.

	F3SW-B8	G3NE-B13	G3NW-B9	G3SW-B3	H3NE-B10	H3NE-B6	H3NW-B2	H3NW-B1	I3NW-B7	I3NW-B5	I3SE-B4	I3SE-B3	I3NW-B4	I3NW-B1	I3SE-B9	I3SW-B4
DCA1	102.4444	269.111	235.4405	282.3632	57.5112	78.41554	92.78378	337.7209	60.2856	112.7817	129.8811	155.2431	69.72221	334.236		
DCA2	62.30295	90.257784	84.38643	166.7326	170.4183	109.086	109.2068	114.6503	252.9482	105.9167	80.2053	110.762	111.6674	176.0561	104.8492	
DCA3	74.05123	103.8455	119.5228	94.33161	182.8238	127.9042	57.46194	107.134	97.249686	41.64962	33.86673	70.34692	78.75471	17.97541	81.08933	
fractures	4	1	2	3	2	3	3	3	0	1	3	1	4	2	1	
soil contact	100	50	40	50	100	100	90	30	60	50	100	100	80	100	100	
moss cover	5	5	10	10	10	20	10	0	5	10	10	0	5	5	0	
bark cover	10	30	90	100	20	5	0	90	10	10	10	10	40	0	0	100
elytra cover	40	30	5	30	30	10	10	50	40	40	40	20	20	30	0	
plant cover	100	100	90	90	100	100	90	80	90	50	30	70	50	90	90	
decay stage	4	2	2	1	3	4	4	1	3	4	4	4	3	4	1	
log age	20	5	7	3	16	20	27	2	31	31	20	20	12	31	2	
dbh	110	80	106	110	84	168	120	90	86	100	120	110	130	105	126	
dist. to edge	105	60	50	120	40	50	60	45	75	60	70	80	25	50	20	
plant diversity	1	1	5	4	5	3	2	2	6	3	2	2	3	2	4	
floristic deviance	0	0	80	50	40	10	10	0	70	10	0	0	0	0	0	
bole forks	2	0	1	2	0	2	2	1	1	1	1	1	3	2	50	
soil humidity	3	2	4	3	5	4	2	1	2	2	3	2	4	2	2	
wind exposure	4	3	4	5	3	3	3	2	2	2	2	1	2	1	2	
sun exposure	4	2	5	3	3	5	3	2	3	3	3	3	2	2	1	
soil type	lac sand	sandhill	lac sand	loamhill	lac sand	sandhill										
log type	broken in 1-7 m. height	broken at root neck	broken in 8-15 m. height													
decay rate	1	0	0	0	0	0	0	0	0	0	0	0	0	0	-2	
ane ran	0	0	20	30	10	10	0	0	20	5	0	0	5	0	0	0
ane nem	100	100	10	40	20	90	80	80	20	40	30	70	50	90	40	
cor bul	0	0	0	0	0	0	0	10	0	5	5	5	0	5	0	
mer per	0	0	0	20	10	0	0	0	5	0	0	0	0	0	0	
ran fic	0	0	50	5	20	5	0	0	20	0	0	0	0	0	0	
lam gal	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
gal odo	0	0	0	0	0	0	0	0	0	0	0	0	5	0	40	
vio fin	0	0	0	0	0	0	0	0	5	0	0	0	0	0	5	
other plants	0	0	2	1	0	0	0	0	2	0	0	0	0	0	0	0

